

# The effects of diazinon on pituitary–gonad axis and ovarian histological changes in rats

Habibollah Johari<sup>1</sup> Ph.D., Mehrdad Shariati<sup>2</sup> Ph.D., Shahram Abbasi<sup>2</sup> M.Sc., Esfandyar Sharifi<sup>2</sup> M.Sc., Hamid Reza Askari<sup>2</sup> M.Sc.

1 Department of Biology, Islamic Azad University, Darab Branch, Darab, Iran.

2 Department of Biology, Islamic Azad University, Kazeron Branch, Kazeron, Iran.

Received: 11 October 2009; accepted: 26 April 2010

## Abstract

**Background:** Diazinon is a widely used Organophosphate insecticide, which is applied against plant pests. This compound has various side effects because it acts as an acetyl cholinesterase enzyme inhibitor.

**Objective:** The aim of present study was to investigate the effects of diazinon on pituitary–gonad axis and ovarian histological changes in rats.

**Materials and Methods:** In total, 50 female wistar rats were divided into 5 groups of 10, including control, sham and experimental groups I, II and III which orally received 50, 100 and 150 mg/kg/bw diazinon for 14 days respectively. Diazinon was administered orally, and 24 hours after the last treatment, blood samples were taken from the heart, centrifuged and sera were evaluated for the concentrations of estrogen, progesterone and gonadotropins via RIA method. In addition, ovaries were removed, fixed and studied with steriological methods.

**Results:** The results show no significant changes in body weight among various groups; while, ovarian weight in experimental group III decreased significantly ( $p < 0.05$ ). Also, no significant changes in the levels of LH, FSH and estradiol hormones were observed. In contrast, the concentration of progesterone showed a significant decrease in all experimental groups compared with the control group. The results showed no significance difference in the mean number of primary, secondary and graffian follicles but there was a significant decline in the mean number of corpus luteum in experimental group receiving 150 mg/kg diazinon ( $p < 0.05$ ).

**Conclusion:** One can conclude that oral administration of diazinon could have adverse effect on progesterone hormone level as well as hazardous effects on ovarian tissues and reproductive processes.

**Key words:** Diazinon, Progesterone, Estrogen, Ovary, Rat.

## Introduction

Exposure to pesticides may be a major cause of various disorders and reproductive dysfunction in humans and animals. Organophosphates are a group of widely used insecticides in agriculture. A German chemist investigated the efficiency of organophosphate as insecticides (1). One of this insecticide family is an acid phosphoric esters called diazinon. This compound is a colourless

fluid, with the boiling temperature of 84°C, 12 hours half life (in animals body) and 300 mg/kg lethal dose in rats (2, 3). Diazinon has acquired a wide usage in farms and households against noxious insects. Organophosphate insecticides are considered only slightly toxic to vertebrates. However there are evidences of greater effects on reproductive function (4). In animals, the absorption of diazinon takes place mainly through digestive system, skin and inhalation; its adverse effects are exerted through the inhibition of acetyl cholinesterase in neurons. Toxic symptoms of diazinon in human include: diarrhea, nausea, excessive salivary secretion, agitation and alertness (5). Degradation and excretion of diazinon occur in liver and kidney by the oxidation and hydrolysis:

### Corresponding Author:

Habibollah Johari, Department of Biology, Islamic Azad University of Darab, Islamic Azad University Avenue, P.O.BOX : 74817 – 83143, Darab, Iran.

**Email:** johari@Iaudarab.ac.ir

its oxidation is carried out by p450 enzyme, and its excretion is done rapidly; in rats for example, 50% is excreted in urine in 12 hours (6). The results of some studies indicate that diazinon and malathion used in commercial formulation could be toxic to *in vitro* fertilization and embryo development (3). Similarly, the insecticide methyl is a reproductive toxicant in the male rat and causes deterioration in the structural integrity of the reproductive organs and also the biochemical parameters in the epididymis (7). A research in 2009 have showed that diazinon treatment of male mice for 30 days brought about significant reduction in sperm counts and spermatogenesis, Leydig and sertoli cells and a decrease in serum testosterone concentration (8). In addition, diazinon can induce testicular cancer and cause a decline in sperm density (9). Some studies have demonstrated that diazinon increases the risk of cancer incidence, especially breast cancer in female farmers (10). It also causes deformation in sperm chromatin, morphology and viability (11). On the other hand, a research on Chinese hamster showed that organophosphates induces sister chromatid exchange (12). Since diazinon has adverse effects on various organ systems specially reproductive system, and since a relatively high percentage of female farmers are exposed to diazinon directly, we tried to investigate the effects of this compound on ovary and female sex hormones in order to highlight its probable undesirable influences and increase awareness in those at risk and thereby exerting more regulatory and control procedures on the usage of this insecticide.

### Materials and methods

Fifty adult female rats with the average weight of  $165 \pm 5$  grams, obtained from animals' house of Islamic Azad University of Kazeroon, were randomly divided into 5 groups of 10 animals each: a control, a sham and three experimental groups (I, II and III). All groups were kept separately in special polycarbonate cages under the same environmental conditions of 22-25°C and 12h lights and dark cycle with unlimited access to water and dry rat food. The experimental groups I, II and III received 50, 100 and 150 milligrams diazinon per kilograms live weight per day (mg/kg/day) respectively. The control group was left untreated and the sham received equivalent amount of distilled water as solvent. For 14 days, diazinon was administered orally via syringes equipped with feeder. Before the start of the experiment, all the rats were sexually co-cycled in

a lab lacking male rats; co-cycling was brought about by injection of 100 micrograms steradiol valerte dissolved in 0.2 milliliter olive oil and after 42 hours 500 micrograms progesterone dissolved in 0.2 milliliter olive oil was also injected. To ascertain the co-cycling vaginal smear was prepared from all animals. To prepare vaginal smear vagina was washed with physiologic serum several times, and was spread on a slide. The slides were examined under light microscope after Gimmsa staining (13, 14).

Twenty four hours after the last treatment, all the rats were weighted, anesthetized by ether and blood samples were taken from the heart directly. In order to separate sera, blood samples were centrifuged at the rate of 5000 RPM for 10 min; serum levels of LH, FSH, estrogen and progesterone were determined by radioimmunoassay method using special rat kits (Specteria, Finland) (15, 16). For histological studies, ovaries were removed, weighted, and placed in Boin's solution and after dehydration stepwise in increasing ethanol samples were cleared by xylene and undergone routine histological processes. 5 µm sections were then prepared and stained by Hematoxilin-Eosine dyes. Primary, secondary and graffian follicles, and corpus luteum in each group were counted (13, 14, 17).

### Statistical analysis

The data were statistically analyzed using SPSS software; mean concentrations of hormones and the ovarian cell counts of various groups were compared with the control using the ANOVA and t-test methods, and  $p < 0.05$  was considered as a significant difference.

### Results

There were no significant differences in the body weight among various groups; except in experimental group III (dosage of 150 mg/kg) which showed a decrease in body weight (Table I). In addition, the results of hormones assay, demonstrated that the serum concentrations of LH, FSH and estrogen comparable in experimental groups; while the progesterone hormone level decreased significantly in all experimental groups ( $p < 0.05$ ) (Table II). Histological studies of the ovarian tissues showed no significant difference in the mean number of primary, secondary and graffian follicles but there was a significant decline in the mean number of corpus luteum in experimental group 3 (Tables III and IV). Also, we found signs of degeneration in corpus luteum (Figures 1-4).

**Table I.** Mean body and ovary weights levels of FSH, LH, estrogen and progesterone in experimental and control groups.

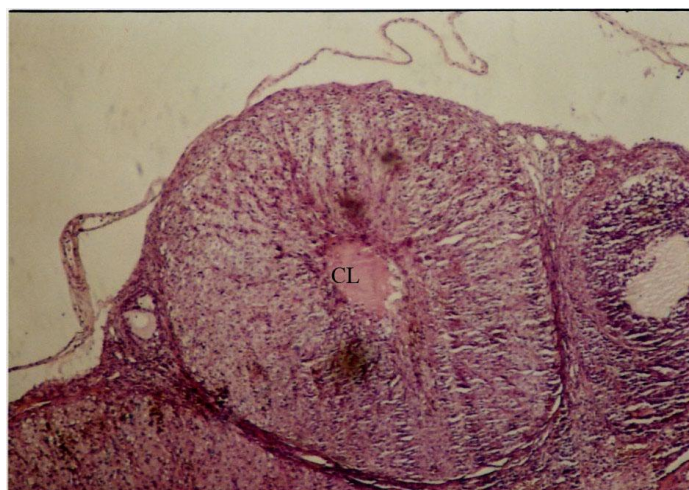
Groups	Control	Sham	I	II	III
Body weight (g)	165.3±3.76	176.6±5.02	170.6±3.12	168.2±3.34	163.9±4.57
Right ovary weight (mg)	107.6±9.38	105.1±1.73	104.2±5.33	92.4±5.61	82.1±6.97*
Left ovary weight(mg)	$\bar{X} \pm SEM$ 103.8±4.09	100.03±4.98	91.3±4.86	93.0±3.67	81.7±4.01*
FSH (mIU/ml)	0.23±0.03	0.25±0.02	0.26±0.03	0.32±0.04	0.27±0.05
LH (mIU/ml)	0.3±0.02	0.28±0.02	0.31±0.03	0.35±0.05	0.3±0.02
Progesterone ( pg/ml)	11.02±1.73	9.57±1.95	5.98±1.52*	5.70±1.51*	3.25±0.58*
Esterogen (pg/ml)	41.50±5.31	31.93±4.4	30.88±2.7	38.63±4.63	41.66±4.75

\* Indicates significant difference between control and experimental groups (p<0.05).

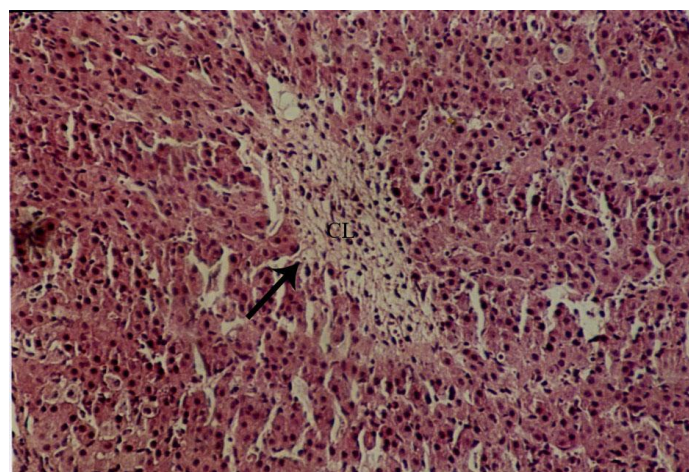
**Table II.** Mean number of primary, secondary, graffian follicles and corpus luteum in right and left ovaries in experimental and control groups.

Groups	Control	Sham	I	II	III
Primary follicle of right ovary	333±0.92	2.83±0.60	3.00±0.37	2.67±0.56	2.50±0.67
Primary follicle of left ovary	2.83±0.48	3.50±0.43	3.33±0.56	2.83±0.54	3.17±0.48
Second follicle of right ovary	1.83±0.31	1.50±0.22	1.67±0.33	1.50±0.22	1.33±0.21
Second follicle of left ovary	$\bar{X} \pm SEM$ 1.67±0.21	1.50±0.22	1.17±0.31	1.33±0.42	1.00±0.26
Graffian follicle of right ovary	0.17±0.05	0.17±0.06	0.33±0.06	0.17±0.09	0.33±0.07
Graffian follicle of left ovary	0.50±0.22	0.50±0.08	0.33±0.09	0.33±0.07	0.33±0.07
Corpus luteum of right ovary	2.50±0.50	1.83±0.31	2.00±0.37	2.00±0.45	1.33±0.21*
Corpus lateum of left ovary	2.17±0.60	2.00±0.45	2.67±0.49	1.33±0.21	0.67±0.21*

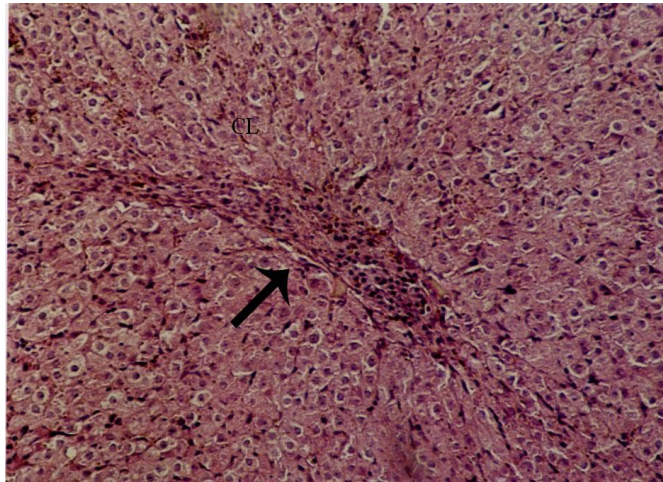
\* Indicates significant difference between control and experimental groups (p<0.05).



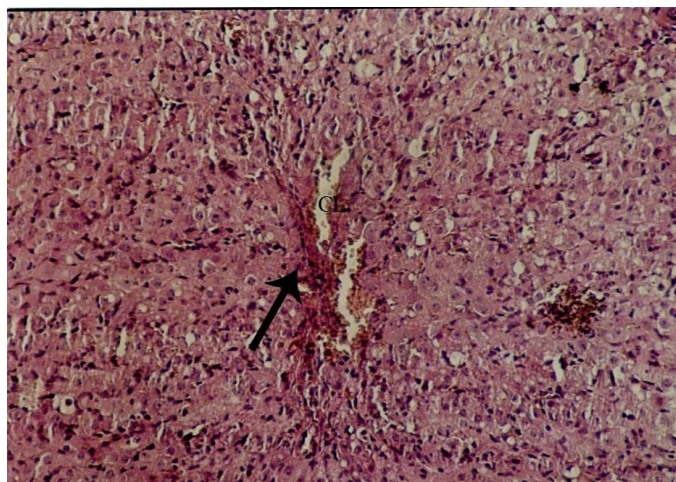
**Figure 1.** Corpus luteum in control group (H&E staining ; x 4 )



**Figure 2.** Corpus luteum in experimental group 1 (H&E Staining ; x10 ). The pointer shows cellular changes , deformation and degeneration in corpus luteum. Effects of diazinon on pituitary – gonad.



**Figure 3.** Corpus luteum in experimental group 2 ( H&E Staining ; x10 )



**Figure 4.** Corpus luteum in experimental group 3 (H&E Staining; x10). The pointer shows cellular changes, deformation and degeneration in corpus luteum

### Discussion

Organophosphates affect mammalian brain development through a variety of mechanisms beyond their shared property of cholinesterase inhibition. The idea is that, different organophosphates target multiple pathways (involved in neural cell development) and they deviate in key aspects that may contribute to disparate neurodevelopmental outcomes (18). Several studies show that organophosphates pesticides exert several effects on mammalian spermatogenesis (8).

Nevertheless, their effects on oogenesis are less studied. After 24-h exposure to 250 microM malathion or 900 nM diazinon, oocytes survival was significantly decreased, and the negative effect of these insecticides on the regulation of genes encoding proteins involved in transcription (BP75), translation (ribosomal protein S5), and mitochondrial function (cytochrome oxidase

subunits I and III), provide evidence for OP insecticides as toxicants for mammals oocytes during the early oogenesis (19).

According to our finding, diazinon has no effect on body weight. In 2003, it was demonstrated that feeding baboon with low concentrations of diazinon for 26 months caused no significant changes in body weight, and concentrations of serum lipids and cholesterol lipoprotein (20). In 1980, the effects of 0.1-15 ppm diazinon on rats were investigated for 92 days, and it was shown to have no influence on diet (21). Our study showed that diazinon significantly decreases ovarian weight and mean number of corpus luteum. Like other Organophosphates, diazinon can increase ACTH; this may promote protein catabolism in ovary which could lead to ovarian weight lost (21). Our investigation shows that diazinon decreases progesterone level but it doesn't induce significant changes in gonadotropins and estrogen concentrations. According to different studies,

organophosphates significantly reduce plasma level of progesterone (23), by the possible mechanism of corticosterones elevation that lead to a decrease in  $3\beta$ -HSD activity which is necessary for progesterone generation (24). Most cholesterol used in steroidogenesis, originate from intracellular sources; indeed, this is the restricting step in steroidogenesis; in fact, a few proteins like Steroidogenic active poly peptide (SAP) and Peripheral Benzo diazpins receptor (PBR) act as cholesterol regulators. However, the protein most studied in this respect is STAR. Most organophosphates are able to interfere in STAR protein gene expression, and reduce its level; thereby, decrease progesterone generation (25, 26). According to some studies, prolactin secretion is controlled by dopamine. On the other hand, some organophosphates, like Dichlorvos, stimulate dopamine level and hence, lead to a decrease in prolactin concentration. Since prolactin can elevate the number of LH receptors, its decrease can reduce the quantity of LH receptors which are involved in progesterone synthesis (27 and 28). Some organophosphates inhibit G-protein activities and could lead to inactivation of LH receptors (29); hence, it may reduce progesterone level.

Our histological studies show signs of apoptosis in corpus luteum of experimental groups. According to some reports, diffusion of  $Ca^{++}$  could activate caspase 12 located on endoplasmic reticulum (30). Likewise, diazinon and other organophosphates induce  $Ca^{++}$  release from intracellular structures and promote caspase 12 activation; this could be the possible mechanism for apoptosis incidence in corpus luteum induced by diazinon (31). On the other hand, diazinon can cause apoptosis through mechanism of ATP decline and thus, bring about changes in mitochondrial membrane potential (32). Some organophosphates activate caspase 3 (33), which is another important apoptotic enzyme. Diazinon can accelerate the synthesis of Interleukin II which in turn, elevate prostaglandins; thereby, causing corpus luteum degeneration (34). Prolactin is an important hormone involved in protection of corpus luteum in many species. However, organophosphates decrease prolactin (35, 36) and through prolactin reduction, they could shorten corpus luteum lifetime.

### Conclusion

Our data showed that, 14 days application of diazinon insecticide can cause a reduction in serum progesterone concentration and ovary

degeneration; therefore, it is strongly recommended that protective measures should be considered before application of diazinon.

### Acknowledgement

The authors wish to thanks the research office of the Islamic Azad University of Kazeroun, Iran for its financial supports.

### References

1. Allison DF, Pamela JL, Angela SH, Bethany LY, Rondell AB, Jett DA. Mechanisms of organophosphate insecticide- Induced airway hyperreactivity. *Am J Physiol lung Cell Mol Physiol* 2004; 286: 963 – 969.
2. Ogut A, Uzunhisarcikli M, Kalender S, Durak D, Bayrakdar F, Kalender Y. The effects of organophosphate Insecticide diazinon on malondialdehyde levels and myocardial cells in rat heart tissue and protective role of vitamine E. *Pesticide Biochemistry and Physiology* 2006; 86: 93 -98.
3. Ducolomb Y, Casas E, Valdez A, Gonzales G, Altamirano – Lozano M, Betancourt M. In vitro effect of malathion and diazinon on oocyte fertilization and embryo development in porcine. *Cell Biol Toxicol* 2009; 25: 623 – 633.
4. Casas E, Bonilla E, Ducolomb Y, Betancourt M. Differential effects of herbicides atrazine and Fenoxaprop – ethyl, and malathion, on viability and maturation of porcine oocytes in vitro. *Toxicol In Vitro* 2010; 24: 224 – 230.
5. Vettorazzi G, Vettorazzi PM. Safety evaluation of chemicals in Food : toxicological data profiles for pesticides 1. Carbamate and organophosphorus insecticides used in agriculture and public health. *Bull World Health Organ* 1975; 52: 1 – 61.
6. Cantor KP, Blair A, Everett G, Gibson R, Burmeister LF, Brown LM, et al. Pesticides and other agricultural risk factors for non – Hodgkin' s lymphoma among men in IoWa and Minnesota. *Cancer Res* 1992; 52: 2447– 2455.
7. Prashanthi N, Narayana K, nayanatara A, Chandra kumar HH, Bairy kl, D'Souza UJ. The reproductive toxicity of the organophosphate pesticide o, o – dimethyl 0 – 4 – nitrophenyl phosphorothioate (methyl parathion) in the male rat. *Folia Morphol* 2006; 65: 309 – 321.
8. Fattahi E, Parivar K, Jorsaraei SGA, Moghadamnia AA. The effects of diazinon on testosterone, FSH and LH levels and testicular tissue in mice. *IJRM* 2009; 7: 59 – 64.
9. Sharpe RM, Skakkebaek NE. Are oestrogens Involved in falling sperm counts and disorders of the male reproductive tract ? *Lancet* 1993; 341: 1392–1395.
10. Gandhi R, Snedeker SM. Pesticides and Breast Cancer Risk, An Evaluation of Chlorpyrifos. Institute for Comparative and Environmental. *Toxicology* 1999; 26: 112 – 134.
11. Pina - Guzman B, Solis – Heredia MJ, Quintanilla – Vega B. Diazinon alters sperm chromatin structure in mice by phosphorylating nuclear protamines. *Toxicol Appl Pharmacol* 2005; 202: 189 – 198.
12. Nishio A, Uyeki EM. Induction of sister chromatid exchange in Chinese hamster ovary cells by organophosphate Insecticides and their oxygen analogs. *J Toxicol Environ Health* 1981; 8: 939 – 946.

13. Tallan SK, Walton JS. Effect of oxytocin on follicular development and duration of the estrous cycle in heifers. *Therigenology* 2000; 53: 951 – 962.
14. Roshangar L, Soleimani Rod J, Nikpoo P, Sayyah Melli M. Effect of oxytocin injection on folliculogenesis, ovulation and endometrial growth in mice. *Iranian Journal of Reproductive Medicine* 2009; 7: 91 – 95.
15. Pidoux G, Gerbaud P, Tsatsaris V, Matpeau O, Ferreira F, Meduri G, et al. Biochemical characterization and modulation of LH / CG – receptor during human trophoblast differentiation. *J Cell Physiol* 2007; 212: 26 – 35.
16. Pegel H, Towns R, Nair A, Menon KM. A novel mechanism for the modulation of luteinizing hormone receptor mRNA expression in the rat ovary. *Mol Cell Endocrinol* 2005; 233: 65 – 72.
17. Howard CV, Reed MG. Unbiased stereology three–dimensional measurement in microscopy. 2nd Ed. Oxford: Bios Scientific Publishers; 2005.
18. Slotkin TA, Seidler FJ. Comparative developmental neurotoxicity of organophosphates in vivo: transcriptional responses of pathways for brain cell development cell signaling, cytotoxicity and neurotransmitter systems. *Brain Res Bull* 2007; 72: 232 -274.
19. Bonilla E, Hernandez F, Cortes L, Mendoza M, Mejia J, Carrillo E, et al. Effects of the insecticides malathion and diazinon on the early oogenesis in mice in vitro. *Environ Toxicol* 2008; 23: 240 – 245.
20. Ibrahim NA, EL – Gamal BA. Effect of diazinon , an organophosphate insecticide, on plasma lipid constituents in experimental animals . *J Biochem Mol Biol* 2003; 36:499-504.
21. Davies DB, Holub BJ. Toxicological evaluation of dietary diazinon in the rat. *Arch Environ Contam Toxicol* 1980a; 9: 637 – 650.
22. Gupta M, Bagchi GK, Gupta SD, Sasmal D, Chatterjee T, Dey SN. Changes of acetylcholine , catecholamines and amino acid in mice brain following treatment with Nuvacron and Furadan. *Toxicology* 1984; 30: 171 – 175.
23. Parkash N, Narayana K, Murthy GS, Moudgal NR, Honnegowda. The effect of malathion, an organophosphate, on the plasma FSH, 17 beta – estradiol and progesterone concentrations and acetylcholinesterase activity and conception in dairy cattle. *Vet Hum Toxicol* 1992; 34: 116 – 119.
24. Valli G, Sudha S, Ravi Sankar B, Govindarajula P, Srinivasan N. Altered corticosterone status impairs steroidogenesis in the granulosa and thecal cells of Wistar rats. *J Steroid Biochem Mol Biol* 2000; 73: 153 – 158.
25. Speroff BL, Glass RH, Kase NG. Clinical Gynecologic Endocrinology and Infertility. 6<sup>th</sup> Ed. Philadelphia: lippincott Williams Wilkins; 1999.
26. Walsh LP, MC Cormick C, Martin C, Stocco DM. Roundup Inhibits steroidogenesis by disrupting steroidogenic acute regulatory (STAR) protein expression. *Environ Health Perspect* 2000; 108: 769 – 776.
27. Chambers RD. Fluorine in organic chemistry. 2<sup>nd</sup> Ed. UK: Black well publishing; 2004.
28. Goodman HM. Basic – medical endocrinology. 2<sup>nd</sup> Ed. New York: Raven Press; 1994.
29. Zou LM, Li SY, Zhang J. Effect of organophosphorus insecticides on G protein – coupled receptor kinase – 2 mediated phosphorylation of M<sub>2</sub> muscarinic receptors. *Zhonghua Lao Dong Wei Sheng Zhi Ye Bing Za Zhi* 2006; 24: 352 – 325.
30. Nakagawa T, Yuan J. Cross – talk between two cysteine protease families : activation of caspase – 12 by calpain in apoptosis. *J Cell Biol* 2000; 150: 887 – 894.
31. Kauffman FC, Davis LH, Whittaker M. Activation of glycogen phosphorylase in rat pheochromocytoma PC 12 cells and isolated hepatocytes by organophosphates . *Biochem Pharmacol* 1990; 39: 347 – 354.
32. Nakagawa Y, Moore G. Role of mitochondria membrane permeability transition in p – hydroxybenzoate ester – induced cytotoxicity in rat hepatocytes. *Biochem Pharmacol* 1999; 58: 811 – 816.
33. Wang X. The expanding role of mitochondria in apoptosis. *Gene & Development* 2001; 15: 2922 – 2933.
34. Aluigi MG, Angelini C, Falugi R, Fossa P, Genever L, Gallus PG, et al. Interaction between organophosphate compounds and cholinergic functions during development. *Chemico – Biological Interactions* 2005; 157: 305 – 316.
35. Johnson MH, Everitt BJ. Essential reproduction. 6<sup>th</sup> Ed. Massachusetts: Blackwell publishing; 2007.
36. Smallridge RC, Carr FE, Fein HG. Diisopropylfluorophosphate ( DFP ) reduces serum prolactin, thyrotropin , luteinizing hormone , and growth hormone and increases adrenocorticotrophin and corticosterone in rats :Involvement of dopaminergic and somatostatinergic as well as cholinergic pathways. *Toxicol Appl Pharmacol* 1991; 108: 284 – 295.