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**Original article**

# The effect of various concentrations of myo-inositol in culture medium on development of bovine embryos

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**Abstract**

**Background:** Progress achieved in culture media formulations have resulted led to an improvement in maintaining the mammalian embryo in culture throughout the preimplantation and pre-attachment period.

**Objective:** The objective of this study was to evaluate the effect of various concentrations of myo-inositol during in vitro fertilization of bovine oocytes on subsequent embryo development.

**Materials and Methods:** Bovine cumulus oocytes complexes (COCs) were matured in vitro at 39$^{o}$C, in humidified 5% $CO_2$ atmosphere for 22-24 h. The COCs were co-incubated with epididymal spermatozoa of post mortem bulls in modified TALP medium for 22-24hr. The fertilization medium used was: 1) TALP medium without myo-inositol (control); 2) control+0.02 g/l myo-inositol; 3) control+0.03 g/l myo-inositol; 4) control+0.04 g/l myo-inositol. Zygotes were cultured in vitro for 8 days when the ratios of in vitro embryo development of the hatched blastocysts were assessed and compared with the control group ($p<0.05$).

**Results:** The presence of 0.04 g/l myo-inpsitol significantly improved overall morula and blastocyst rates (46.94%) compared to control (32.19%), but there was no difference in the percentage of embryos successfully developed to the morula and blastocyst stage when different levels of myo-inositol were used (46.94, 36.36 and 37.33% respectively). The mean percentage of cleavage rate was not significantly affected by treatments.

**Conclusion:** These results suggest that, addition of 0.04 g/l myo-inositol in TALP medium is more beneficial for subsequent bovine embryonic development.

***Key words:*** *Bovine, Myo-inositol, Fertilization, Embryo development.*

## Introduction

Myo-inositol is formed from glucose-1-phosphate in a $NAD^+$-catalyzed oxidation/ reduction reaction. It is a cyclic alcohol with 6 hydroxyl groups, one of many stereoisomers (1). Myo-inositol plays an important role in cell morphogenesis and cytogenesis, lipid synthesis, structure of cell membranes and cell growth (2).

The effects of vitamins have been tested on oocytes maturation and preimplantation embryo culture systems and some benefits on oocytes maturation and embryo development have been observed. The addition of inositol, choline and pantothenate to chemically defined medium, improved hatched blastocyst of hamsters embryos (3).

Rabbit blastocysts required inositol, pyrodoxal, riboflavin and niacinamide for expansion and growth. The most drastic reduction in expansion was caused by the omission of inositol (4). Inositol is essential for rabbit blastocyst expansion, cell proliferation and protein synthesis and show that the effect on rabbit blastocyst expansion (5).

Addition of citrate and myo-inositol improved blastocyst development rates in defined medium to comparable to those obtained with serum, BSA or co-culture (6). Improvement in preimplantation embryo development has been reported with myoinositol uptake from culture medium in studies with a number mammalian species (7, 8).

However, on the basis of our knowledge, there is not much information about the effect of various concentrations of myo-inositol on in vitro fertilization and embryo development in bovine. The objective of this study was to determine if supplementation of various concentrations of myo-inositol in TALP fertilization medium could subsequent in vitro embryo development.

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## Materials and methods

### Chemicals

Unless mentioned otherwise, growth factors and other chemicals were purchased from Sigma (USA), plastics from Falcon (UK), hormones (eCG and hCG) from Intervet (Netherlands). All the media were incubated at 39°C under humidified atmosphere of 5% $CO_2$ in air for 1 hr prior to use.

### Oocyte collection

Ovaries were transported from the local abattoir to laboratory in 0.9% saline at 30-35°C. They were washed three times with warmed 0.9% saline. COCs were aspirated from 2-6 mm follicles using a 20-gauge needle attached to a 10 ml syringe. COCs were collected in TCM HEPES medium supplemented with 14.24 mg/ml heparin and 50 μg/ml gentamicin.

### In vitro maturation

COCs with at least three layers of nonexpanded cumulus and homogeneous cytoplasm were selected and washed three times in TCM HEPES and once in TCM-199 maturation medium supplemented with 0.23 mmol/l sodium pyruvate, 50 ng/ml epidermal growth factor (EGF), 10 IU/ml equine chorionic gonadotrophin (eCG), 10 IU/ml human chorionic gonadotrophin (HCG) and penicillin/streptomycin (100 U/ml penicillin, 100 μg/ml streptomycin). Oocytes were matured in groups of 10 per 50 μl droplets in culture dishes under 10 ml mineral oil. Maturation proceeded for 22-24h at 39°C and 5% $CO_2$ in air with 100% humidity (9).

### In vitro fertilization

At the end of the maturation period, oocytes were briefly placed in TCM199 maturation medium supplemented with 500 IU/ml hyaluronidase, followed by gentle pipeting to dissociate surrounding cumulus cells and washed three times in fertilization medium.

Oocytes (10-15) were placed into a 100 µl drop to which sperm cells were added 5 min later. An epididymal spermatozoa was collected from post mortem bulls and kept at room temperature for up to 2 h, then washed in TALP, centrifuged twice at 200×g for 5 min. The medium used for in vitro fertilization was TALP, supplemented with 6 mg/ml of BSA-FFA, 0.23 mmol/l sodium pyruvate, 10 µg/l heparin, 20 µmol/l penicillamine, 10 µmol/l hypotaurine, 1 µmol/l epinephrine (PHE) as originally described by Parrish *et al* (10), in this study sperms and oocytes co-incubated without or with different concentration of myo-inositol.

Treatment groups were: I) TALP medium without myo-inositol (Control); II) Control+0.02 g/l myo-inositol; III) Control+0.03 g/l myo-inositol; IV) Control+0.04 g/l myo-inositol. Sperms were counted in a hemacytometer and checked for acceptable motility (i.e., at least 80% progressively motile); then oocytes were inseminated by addition of sperm to result in a final concentration of $1.0\times10^6$ motile sperm per milliliter. Sperm and oocytes were incubated under mineral oil at 39°C under humidified atmosphere of 5% $CO_2$ for 22-24h.

### In vitro culture

Embryo culture took place in modified SOF under mineral oil in a humidified atmosphere of 5% $CO_2$ and 5% $O_2$ and 90% $N_2$ at 39°C. Between 15 and 22-24 h after insemination, presumptive zygotes were denuded of surrounding cumulus cells by repeated pipetting in SOF HEPES and subsequently washed three times in SOF HEPES before being transferred to the culture droplets (50 µl) in groups of 10-15 embryos. Cleavage was assessed after 48 h of culture, and the number of embryos developing to the morula and blastocyst stages was assessed on day 8.

To prevent toxic accumulation of ammonium as a result of amino acid degradation, SOF medium was replaced every 48 h. In this study, we used a two-culture system. The first system (SOFC1) medium contained 0.8% BSA crystallized, MEM nonessential amino acids, 1 mM glutamine, 1.5mM glucose, and 10 µM EDTA for the first 48 h. Then, the medium was replaced by the second system (SOFC2) containing 0.8% BSA crystallized, MEM nonessential amino acids, MEM essential amino acids, 3mM glucose, and 1mM glutamine for the remaining 6 days of culture (9).

### Statistical Analysis

This experiment was replicated 5 times and COCs were randomly allocated into each treatment group. The data were analyzed by

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one-way ANOVA using SPSS (11.5, 2004) program. Duncan's multiple range test was used to test the differences between treatments. P-value <0.05 indicated as a significant difference.

## Results

In the present study, there were no differences between treatments in the percentage of embryos successfully developing to the cleavage stage, expressed as a percentage of total inseminated oocytes (Table I). The presence of 0.04 g/l myo-inositol significantly increased the percentage of morula and blastocyst (p<0.05) as compared with control (46.94 and 32.19; respectively, Table I). No significant difference was also observed in morula and blastocyst rates between 0.02, 0.03 and 0.04 g/l myo-inositol (36.36%, 37.33% and 46.94%; respectively).

**Table I.** Effect of different concentrations of myo-inositol added during IVF of bovine oocytes on subsequent embryonic development.

| Treatments* | Oocytes (n) | Percentage cleaved | Morula and blastocyst/ cleaved (%)** |
|---|---|---|---|
| Treatments I | 135 | 66.55 ± 4.35 | 32.19±3.49[b] |
| Treatments II | 120 | 66.66 ± 4.42 | 36.36±2.35[ab] |
| Treatments III | 128 | 61.38 ± 4.45 | 37.33±2.98[ab] |
| Treatments IV | 117 | 62.59 ± 4.83 | 46.94±6.48[a] |

Values within a column with different superscripts (a, b) are significantly different, (p<0.05).
* Treatments I: TALP medium without myo-inositol (Control).
Treatments II: Control + 0.02 g/l myo-inositol.
Treatments III: Control + 0.03 g/l myo-inositol.
Treatments IV: Control + 0.04 g/l myo-inositol.
** Sum of the morula and blastocysts per cleaved oocytes.

## Discussion

Myo-inositol has long been known as an essential growth factor for mammalian cells (11). Inositol is essential for the synthesis of certain membrane phospholipids in cells and it is possible that at the blastocyst stage it becomes necessary for the synthesis of phospholipid components of new membranes (12, 13). Inositol may play an essential role in the formation of pronuclei during in vitro fertilization (3). Rabbit blastocysts required inositol, pyridoxal, riboflavin and niacinamide for expansion and growth (4, 14).

Kane showed that the omission of inositol, pyridoxine, riboflavin, and niacinamide resulted in large statistically significant decreases in blastocyst expansion but omission of $B_{12}$ resulted in a significant increase in blastocyst expansion indicating that the level present in F10 is toxic to rabbit blastocysts (14). Funahashi *et al* reported that the addition of myo-inositol in maturation medium of procine oocytes had no significant effect on cytoplasmic maturation of oocytes (15).

They concluded that, this may be due to a difference in the effective order because supplementation with 12 mM myo-inositol increased the osmolality of maturation medium by only 6 mOsM (milli osmole). In the present study, the addition of myo-inositol (0.04 g/l) in fertilization medium (TALP) containing BSA significantly increased blastocyst development as compared with control.

This finding did not agrees with previous data from Holm *et al* who reported no effect of myo-inositol on blastocyst development in SOF containing citrate and BSA (6). Nevertheless, these author a stimulating effect of myo-inositol in medium containing polyvinyl alcohol (PVA) and reported that minute concentrations of myo-inositol could be present in commercial BSA preparations. Another study reported there was higher developmental ability of embryos cultured in chemically defined medium with myo-inositol, EGF and modification of energy substrate composition (16).

Kane used high concentrations (75 μm) of myo-inositol to culture medium and found a significant effect on rabbit blastocyst

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expansion (17). Fahy *et al* indicated that inositol is essential for rabbit blastocyst expansion, cell proliferation and protein synthesis (18). Chiu *et al* showed that, myo-inositol has a positive effect on meiotic maturation and developmental competence of maturing mouse oocytes in the absence of exogenous gonadotrophins (19). In addition, previous work a from our laboratory showed that addition of myo-inositol as compared to MEM vitamins may have beneficial effect on blastocyst rate in sheep (8).

Results from the present study agree with previous works showing that the addition of myo-inositol to maturation and culture medium increased blastocyst rate. In conclusion, present study indicated that under the defined conditions, responses to supplementation of various concentrations of myo-inositol to fertilization medium depended on the stage of embryo development (3, 4, 14, 19).

The presence of 0.04 g/l myo-inositol in fertilization medium increased morula and blastocyst rate as compared to control groups. It is possible, lower concentrations of myo-inositol were ineffective or not sufficient to IVF and subsequent embryonic development. However, further studies needed to determine whether the presence of myo-inositol in fertilization medium in high concentrations could further enhance fertilization and subsequent embryo development or not.

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## References

1. Michal G. Biochemical phathways. Berlin: Spektrum; 1999.
2. Downes CP. The cellular function of myo-inositol. *Biochem Soc Trans* 1989; 17: 259-268.
3. Kane MT, Bavister BD. Vitamin requirements for development of eight-cell hamster embryos to hatching blastocysts in vitro. *Biol Reprod* 1988; 39: 1137-1143.
4. Daniel JC Jr. Vitamins and growth factors in the nutrition of rabbit blastocysts in vitro. *Growth* 1967; 31: 71-77.
5. Fahy MM, Kane MT. Incorporation of [3H] inositol in to phosphoinositides and inositol phosphates by rabbit blastocysts. *Mol Reprod Dev* 1993; 34: 391-395.
6. Holm P, Booth PJ, Schmidt MH, Greve T, Callesen H. High bovine blastocyst development in a static in vitro production system using SOFaa medium supplemented with sodium citrate and myo-inositol with or without serum-proteins. *Theriogenology* 1999; 52: 683-700.
7. Hynes AC, Sreenan JM, Kane MT. Uptake and incorporation of myo-inositol by bovine preimplantation embryos from two-cell to early blastocyst stages. *Mol Reprod Dev* 2000; 55: 265-269.
8. Kafilzadeh F, Karami Shabankareh H, Soltani L, kazemine Jasemi V. Comparison of the effects of myo-inositol alone or in combination with MEM vitamins on embryo development of sanjabi sheep. *J Iran Anat Sci* 2011; 8: 189-196.
9. Gandhi AP, Lane M, Gardner DK, Krisher RL. A single medium supports development of bovine embryo throughout maturation, fertilization and culture. *Hum Reprod* 2000; 15: 395-401.
10. Parrish JJ, Susko-Parrish JL, Winer MA, First NL. Capacitation of bovine sperm by heparin. *Biol Reprod* 1988; 38: 1171-1180.
11. Eagle H, Oyama VI, Levy M, Freeman AE. Myo-Inoshol as an essential growth factor for normal and malignant human cells in tissue culture. *J Biol Chem* 1957; 226: 191-205.
12. Hawthorne JN. Inositol phospholipids. In: Hawthorne JN, Ansell GB. Phospholipids. Amesterdam: Elsevier Biomedical Press; 1982.
13. Vance DE. Phospholipid metabolism in eukaryotes. In: Vance DE, Vance JE, editors. Biochemistry of lipids and membranes. Menlo Park, CA: The Benjamin/ Cummings Publishing Co. Inc; 1986.
14. Kane MT. The effects of water-soluble vitamins on the expansion of rabbit blastocysts in vitro. *J Exp Zool* 1988; 245: 220-223.
15. Funahashi H, Kim NH, Stumpf TT, Cantley TC, Day BN. Presence of organic osmolytes in maturation medium enhances cytoplasmic maturation of porcine oocytes. *Biol Reprod* 1996; 54: 1412-1419.
16. Lim KT, Jang G, Ko KH, Lee WW, Park HJ, Kim JJ, et al. Improved in vitro bovine embryo development and increased efficiency in producing viable calves using defined media. *Theriogenology* 2007; 67: 293-302.
17. Kane MT. Effects of the putative phospholipid precursors, inositol, choline, serine and ethanolamine, on formation and expansion of rabbit blastocysts in vitro. *J Reprod Fertil* 1989; 87: 275-279.
18. Fahy MM, Kane MT. Inositol stimulates DNA and protein synthesis, andexpansion by rabbit blastocysts in vitro. *Hum Reprod* 1992; 7: 550-552.
19. Chiu TY, Rogers MS, Briton-Jones C, Haines C. Effects of myo-inositol on the in-vitro maturation and subsequent development of mouse oocytes. *Hum Reprod* 2003; 18: 408-416.

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