

The point mutations of mitochondrial tRNA threonine and proline in idiopathic repeated pregnancy loss

Seyed Mohammad Seyedhassani^{1,2} M.D., Massoud Houshmand¹ Ph.D., Seyed Mehdi Kalantar² Ph.D., Abbas Aflatoonian² M.D., Glayol Modabber¹ M.Sc., Fatemeh Hadipour³ M.D., Mohammad Hossein Fallahzadeh² Ph.D.

1 National Institute for Genetic Engineering and Biotechnology, Tehran, Iran.

2 Research and Clinical Center for Infertility, Shahid Sadoughi University of Medical Sciences, Yazd, Iran.

3 Special Medical Center, Tehran, Iran.

Received: 30 January 2010; accepted: 14 March 2010

Abstract

Background: Mitochondrial transfer RNAs (tRNA) genes are essential components of protein biosynthesis. These genes are hotspots for mutations. These mutations are associated with a wide spectrum of human disease. Many genetic factors are known in assessment of repeated pregnancy loss (RPL).

Objective: The aim of this study was analysis of tRNA^{Thr} and tRNA^{Pro} in women with RPL.

Materials and Methods: The nucleotide variations of threonine and proline were investigated in 96 women with idiopathic repeated pregnancy loss. The related mitochondrial area was amplified using a polymerase chain reaction (PCR). The PCR products were demonstrated by 2% agarose gel electrophoresis, and all the positive samples were purified and verified by an automated DNA sequencing method.

Results: The sequence analysis revealed 4 mutations in tRNA^{Thr}. These mutations were A15907G in 2 cases (2.08%), A15924G in 3 cases (3.12%), G15928A in 10 cases (10.42%) as the most common mutations and G15930A in 3 cases (3.12%) as a novel mutation. Also, the result of tRNA^{Pro} sequencing showed the T15972C mutation in 1 woman (1.04%) as a novel mutation.

Conclusion: These tRNAs mutations can alter their steady state level and affect the structure of tRNAs. It results in protein synthesis defects and, in turn, mitochondrial dysfunction. The mutations of these genes may help in the assessment of RPL. Further study of an expanded series of these tRNA mutants is recommended to describe their etiologic role in idiopathic RPL.

Key words: tRNA, Mitochondrial mutation, Repeated pregnancy loss.

Introduction

Pregnancy loss is the most common complication of pregnancy and can be defined as the unplanned spontaneous loss of pregnancy

Corresponding Author:

Massoud Houshmand, Medical Genetic Department, National Institute of Genetic Engineering and Biotechnology (NIGEB), Tehran, Iran.

E-mail: massoudh@nigeb.ac.ir, housh62@yahoo.com

before the fetus be able to survive extra uterine. Traditionally, repeated pregnancy loss (RPL) has been defined as at least three or more consecutive recognized pregnancy loss. Advances in the detection of early pregnancy revealed that about 70% of human conceptions fail to achieve viability (1); but clinically recognized pregnancy loss will occur in 15% of cases before 20 weeks of gestations (2). About 1 in 300 couples and 0.5-2% of women involved in RPL (3).

Many etiological factors are known for RPL. Usually, most females with a chief complaint of RPL will be evaluated under the care of gynecologist for these possible causes. These factors are responsible for about 50% of RPL. They include uterine anomalies, chromosomal aberrations, infectious conditions and endocrine dysfunction such as luteal phase deficiency and hypothyroidism (4, 5). Genetic factors are actively being sought in idiopathic cases. Association studies have been done to help understand the role of candidate single genes toward the fetal loss. Until now, some polymorphisms have been suggested that increase the chance of pregnancy loss in women (6-14).

A recent study revealed a higher frequency of mitochondrial DNA (mtDNA) variations in women with RPL (15). Mitochondria are the bioenergetics and metabolic centers of the cells. During a process of high-energy consumption such as cell proliferation and development, the role of mitochondria and genome condition and competency are more important. This role is conducted through oxidative phosphorylation by producing ATP (16). Dysfunction in the mitochondrial respiratory chain causes a various group of progressive incurable diseases leading to severe disability and premature death (17). Increasingly, it is supposed that mitochondrial dysfunctions can cause oocyte wastage and early fetal loss by changing the activation of apoptotic process (18).

The double strand circular mitochondrial DNA consists of 16,569 base pairs encoding 37 total genes in human: 22 tRNA, 2 rRNA, and 13 peptide genes (19). Since the first description of pathogenic mutations in the mitochondrial genome, over 200 disease-correlated point mutations and rearrangements have been found in association with a variety of mitochondrial cytopathies (19). More than half of these mutations have been located in tRNA genes that constitute 9% of the entire mitochondrial genome (20).

Thus, mitochondrial tRNA genes are hotspots for mitochondrial pathogenesis and contribute in a disproportionate way to the etiology of disorders caused by mitochondrial DNA mutations, which is conceivable due to their central role in mitochondrial protein synthesis. Previously, the significant difference in the prevalence of spontaneous abortions was shown in the diabetic

RPL patients with tRNA leucine mutation at position 3243 (21). In comparison, a little less than half of the mitochondrial mutations affect protein coding genes, which comprises 68% of the entire mitochondrial genome (22). We are describing mutations on tRNA threonine and proline in women with idiopathic RPL.

Materials and methods

This research was an analytical descriptive study. In total 96 women were been diagnosed as idiopathic RPL at a primary stage of evaluation. They were referred between September 2006 and June 2008 to the Research and Clinical Center for Infertility, Yazd University of Medical Sciences, Yazd, Iran. All of these women had past history of three or more consecutive pregnancy loss before 20 weeks of gestation. They had no history of live birth delivery.

The known causes of RPL were evaluated. Diagnostic work up was consisting of uterine sonography, TORCH infections study (Toxoplasmosis, Rubella, Cytomegalovirus, Herpes Simplex virus type II and Listeria), and assessment of hormonal status, IgM and IgG anticardiolipin, antiphospholipids antibodies and paternal and maternal karyotypes. The cases were explained and encouraged for consent and taking part in this study. The study was approved by ethic committee.

A Flexigene blood DNA kit (DNA fast, QIAGEN, Cat. No. 51204) was used for isolating DNA from the blood samples based on the manufacturer's protocol. The extracted DNA was kept at 4°C. One primer pairs was used for amplifying the proline and threonine tRNAs coding regions. The 5' end primer of this region (5'-ATC ATT GGA CAA GTA GCA TC- 3') from nucleotide 15791 to 15810, and the 3' end primer of it (5'-GCT CCG GCT CCA GCG TCT CG-3') from nucleotide 91 to 110 was used to amplify this genome.

The reaction mixture for PCR contained 7 pmol of each primer, 1 unit of Taq polymerase (Cinnagene, Iran), each dNTP (Cinnagene, Iran) at a final concentration of 200 µM, and 2.5 µl PCR buffer at a final volume of 25 µl by distilled water. The reactions were done in thermal cycler (ASTECC- Japan). The PCR condition was as initial denaturation at 95°C for 5min, then 35 cycles of denaturation at 94°C for 60s, annealing at 58°C for

60s, extension at 72°C for 35s, and then final extension at 72°C for 5min. The PCR products were evaluated on 2% agarose gel and then these fragments were purified and sequenced by MacroGen Company (Seoul, South Korea). The published revision of Cambridge reference sequence (<http://www.mitomap.org/>) was used for comparing the results by the Chromas and Clustal X program. The sequence variants not found in the corresponding record of MITOMAP and other human databases were defined as novel variations.

Results

Our data showed that the Mean±SD age of the women with RPL was 28.73±5.86 years and the mean±SD for the gestational age at the time of miscarriages was 10.35±3.75 weeks. These women had the history of 3 to 11 miscarriages (median 3).

PCR primers from both sides of the threonine and proline tRNA genome were used to amplify the related mitochondrial genome sequence. Direct automated sequencing of the PCR-amplified mtDNA was done.

The sequence analysis of threonine tRNA revealed 4 mutations (Table I) (figure 1).

Table I: Characteristics of proline tRNA mutations in RPL women.

Gene mutation	tRNA domain	Number	Percent (%)
A15907G	D- stem	2	2.08
A15924G	Anticodon stem	3	3.12
G15928A	Anticodon stem	10	10.42
G15930A ^a	V-loop	2	2.08

^a Novel mutation.

Among them, G15930A mutation was novel at the V loop in 2 women. Substitution of G to A at nucleotide 15928 was more common and seen in 10 females (10.42%).

The other two mutations were A15907G and A15924G that were found in 2 and 3 women respectively. One mutation was found in proline tRNA at nucleotide 15972 in one patient, which change T to C. This mutation was located in D-Loop Domain of this tRNA and has not been reported yet (Figure 2).

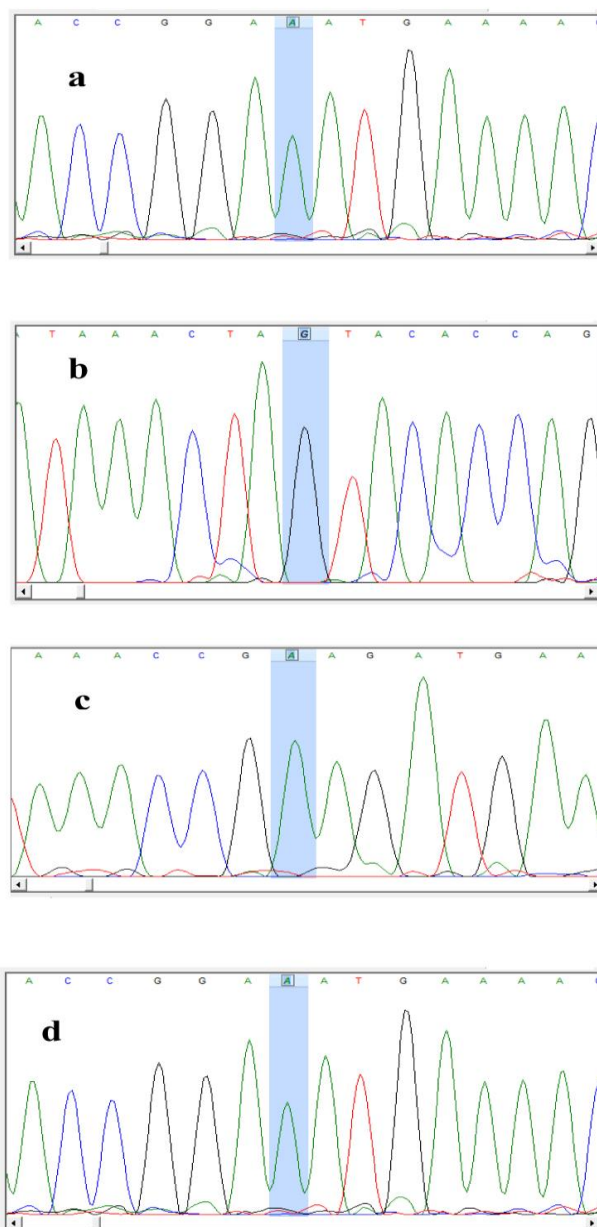


Figure 1. Results of direct sequencing of tRNA^{Thr} gene. a: A15907G, b: A15924G, c: G15928A, d: G15930A.

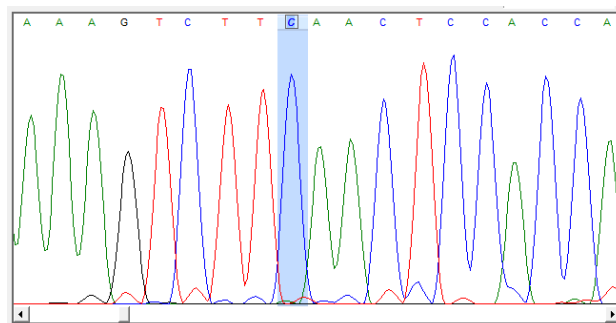


Figure 2. T15972C novel mutation in tRNA^{Pro} gene.

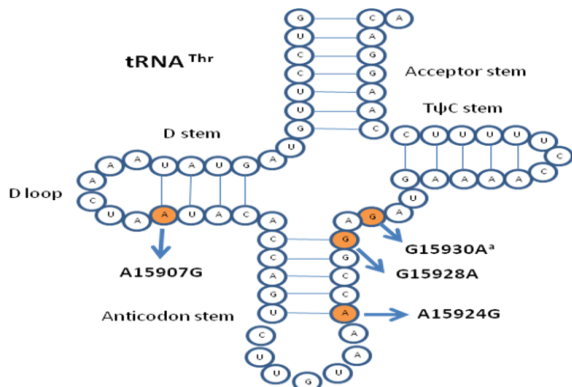


Figure 3. Location of the mutations in human mitochondrial tRNA^{Thr} in women with RPL: the locations of mutations; ^a novel mutation.

Discussion

The tRNAs are small ribonucleic acids (62 to 95 nucleotides) and essential components of protein synthesis because they function to transport amino acids to the ribosome, match them to the codons of mRNAs and facilitate their protein biosynthesis (23).

In this study tRNA threonine and proline were investigated. In total, A15907G, A15924G, G15928A and G15930A substitutions among 65 nucleotides of threonine tRNA, but only one T15972C substitution in 67 nucleotides of proline tRNA were found.

Mitochondrial tRNA^{Thr} mutations

The A15907G mutation is located at the D-stem domain of tRNA^{Thr} (Figure 3). As our knowledge this mutation is not reported in RPL patients or in human-related diseases. In one study, this mutation was reported in 2% of a normal control group as a variation (24). The A15924G in anticodon stem of tRNA^{Thr} was seen in 3 women with RPL. This mutation is reported in some diseases such as: mitochondrial encephalopathy (25), Parkinson's disease (26), Idiopathic cardiomyopathy (27) and fatal infantile respiratory enzyme deficiency (28). The other mutation in anticodon stem was G15928A that was seen in 10 women and was the most common mutation in tRNA^{Thr} in our study. Also, the recent mutation is reported in Parkinson's disease (26) and multiple sclerosis patients with severe optic involvement (29). The nucleotides in anticodon are the least affected bases because it has been identified only one mutation in one tRNA affects any of the three bases necessary for decoding (30). Substitutions at these locations could cause cellular outcomes too severe to sustain cell growth. The G15930A in V-loop or accessory stem was seen in 2 cases. Although it seems the

effect of this substitution is less than the others, however, identifying its role needs more evaluation. The other mutations which are reported previously consisting of: T15908C in deafness-associated 12S rRNA A1555G mutation (31), G15915A in mitochondrial encephalomyopathy (32, 33), A15923G in lethal infantile mitochondrial myopathy (25), fatal infantile respiratory enzyme deficiency (28), newborn cardiopulmonary arrest (34), G15927A in Parkinson's disease (26), multiple sclerosis patients with severe optic involvement (29), deafness-associated 12S rRNA A1555G mutation (35), G15950A in Parkinson's disease (36) and A15951G in LHON-associated ND4 G11778A mutation (37).

Mitochondrial tRNA^{Pro} mutation

The T15972C mutation is found at the D-loop domain of tRNA^{Pro}. There are several reports of this area as A15965G mutation in Parkinson's disease (37), C15975T in late-onset ataxia, retinitis pigmentosa, deafness, leukoencephalopathy and complex I deficiency (39), C15990T in myopathies (40), G15995A in cystic fibrosis (41) and T16002C as a novel mutation (42). However, T15972C is seen in 1 woman with RPL and is not reported yet. Two pathways for inducing the mtDNA including tRNA mutations are considerable. It is demonstrated that the disorder of mtDNA can be induced by the defects of nuclear DNA (43). Another promising pathway is associated with the reactive oxygen species (ROS). The mitochondrial genome is extremely susceptible to damages from continuous exposure to ROS. It has been suggested that the ROS is produced endogenously from mitochondrial respiratory chain and have been considered to be involved in the increased ratio of point mutant mtDNA (44). It is thought that the condition is probably induced by the inhibition of the repair system for ROS mediated damage to mtDNA, detoxification of ROS, or increase in ROS production and might be possible causes for tRNA point mutations of mtDNA. The various aspects of tRNAs function and the effect of their different mutations have been evaluated. Disease-related point mutations could potentially influence mitochondrial tRNA and affect their primary, secondary, and tertiary structure. It leads to protein synthesis defects and, in turn, mitochondrial dysfunction. Ultimately, these disturbances result in cellular dysfunction which is more important in cell proliferation and development. Embryo as a main material in protein synthesis and related functions can be more sensitive to these alterations leading to wastage.

Conclusion

Further study of an expanded series of these tRNA mutants is recommended in order to create a consensus or framework. It would permit the description of the cellular and phenotypic effects of tRNA mutations in related diseases. Because of difficulties to get aborted materials, it was focused on parents. More studies are necessary to clarify and show the primary or secondary role of tRNA mutations in embryonic development.

Acknowledgment

The authors would like to thank IVF teams of Yazd Research and Clinical Centre for Infertility, the laboratory team of National Institute of Genetic Engineering and Biotechnology and also Yazd clinic of genetics for their collaboration in this study.

References

- Edmonds DK, Lindsay KS, Miller JF, Williamson E, Wood PJ. Early embryonic mortality in women. *Fertility and sterility* 1982; 38: 447-453.
- Warburton D FF. Spontaneous abortion rate in man: data from reproductive histories collected in a medical genetics unit. *Am J Hum Genet* 1963: 1-25.
- Wilcox AJ, Weinberg CR, O'Connor JF, Baird DD, Schlatterer JP, Canfield RE *et al.* Incidence of early loss of pregnancy. *The New England journal of medicine* 1988; 319: 189-194.
- Hatasaka HH. Recurrent miscarriage: epidemiologic factors, definitions, and incidence. *Clinical obstetrics and gynecology* 1994; 37: 625-634.
- Clifford K, Rai R, Watson H, Regan L. An informative protocol for the investigation of recurrent miscarriage: preliminary experience of 500 consecutive cases. *Human reproduction (Oxford, England)* 1994; 9: 1328-1332.
- Tempfer C, Unfried G, Zeillinger R, Hefler L, Nagele F, Huber JC. Endothelial nitric oxide synthase gene polymorphism in women with idiopathic recurrent miscarriage. *Human reproduction (Oxford, England)* 2001; 16: 1644-1647.
- Pietrowski D, Bettendorf H, Riener EK, Keck C, Hefler LA, Huber JC *et al.* Recurrent pregnancy failure is associated with a polymorphism in the p53 tumour suppressor gene. *Human reproduction (Oxford, England)* 2005; 20: 848-851.
- Goodman C, Goodman CS, Hur J, Jeyendran RS, Coulam C. The association of Apoprotein E polymorphisms with recurrent pregnancy loss. *Am J Reprod Immunol* 2009; 61: 34-38.
- Lee HH, Hong SH, Shin SJ, Ko JJ, Oh D, Kim NK. Association study of vascular endothelial growth factor polymorphisms with the risk of recurrent spontaneous abortion. *Fertility and sterility* 2010; 93: 1244-1247.
- Govindaiah V, Naushad SM, Prabhakara K, Krishna PC, Radha Rama Devi A. Association of parental hyperhomocysteinemia and C677T Methylene tetrahydrofolate reductase (MTHFR) polymorphism with recurrent pregnancy loss. *Clinical biochemistry* 2009; 42: 380-386.
- Topalidou M, Effraimidou S, Farmakiotis D, Papadakis E, Papaioannou G, Korantzis I *et al.* Low protein Z levels, but not the intron F G79A polymorphism, are associated with unexplained pregnancy loss. *Thrombosis research* 2009; 124: 24-27.
- Rull K, Nagirnaja L, Ulander VM, Kelgo P, Margus T, Kaare M *et al.* Chorionic gonadotropin beta-gene variants are associated with recurrent miscarriage in two European populations. *The Journal of Clinical Endocrinology and Metabolism* 2008; 93:4697-4706.
- Sata F, Yamada H, Yamada A, Kato EH, Kataoka S, Saijo Y *et al.* A polymorphism in the CYP17 gene relates to the risk of recurrent pregnancy loss. *Mol Hum Reprod* 2003; 9: 725-728.
- Firouzabadi RD, Ghasemi N, Rozbahani MA, Tabibnejad N. Association of p53 polymorphism with ICSI/IVF failure and recurrent pregnancy loss. *Aust N Z J Obstet Gynaecol* 2009; 49: 216-219.
- Kaare M, Gotz A, Ulander VM, Ariansen S, Kaaja R, Suomalainen A *et al.* Do mitochondrial mutations cause recurrent miscarriage? *Mol Hum Reprod* 2009; 15: 295-300.
- Dumollard R, Duchon M, Carroll J. The role of mitochondrial function in the oocyte and embryo. *Current Topics in Developmental Biology* 2007; 77:21-49.
- DiMauro S, Schon EA. Mitochondrial DNA mutations in human disease. *American Journal of Medical Genetics* 2001; 106: 18-26.
- Van Blerkom J. Mitochondria in human oogenesis and preimplantation embryogenesis: engines of metabolism, ionic regulation and developmental competence. *Reproduction (Cambridge, England)* 2004; 128: 269-280.
- Anderson S, Bankier AT, Barrell BG, de Bruijn MH, Coulson AR, Drouin J *et al.* Sequence and organization of the human mitochondrial genome. *Nature* 1981; 290: 457-465.
- Kogelnik AM, Lott MT, Brown MD, Navathe SB, Wallace DC. MITOMAP: a human mitochondrial genome database--1998 update. *Nucleic Acids Research* 1998; 26: 112-115.
- Levinger L, Giege R, Florentz C. Pathology-related substitutions in human mitochondrial tRNA(Ile) reduce precursor 3' end processing efficiency in vitro. *Nucleic Acids Research* 2003; 31: 1904-1912.
- Yanagisawa K, Uchigata Y, Sanaka M, Sakura H, Minei S, Shimizu M *et al.* Mutation in the mitochondrial tRNA(leu) at position 3243 and spontaneous abortions in Japanese women attending a clinic for diabetic pregnancies. *Diabetologia* 1995; 38: 809-815.
- Florentz C, Sohm B, Tryoen-Toth P, Putz J, Sissler M. Human mitochondrial tRNAs in health and disease. *Cell Mol Life Sci* 2003; 60: 1356-1375.
- Schimmel PR. Recent results on how aminoacyl transfer RNA synthetases recognize specific transfer RNAs. *Molecular and Cellular Biochemistry* 1979; 25: 3-14.
- Houshmand M, Larsson NG, Holme E, Oldfors A, Tulinius MH, Andersen O. Automatic sequencing of mitochondrial tRNA genes in patients with mitochondrial encephalomyopathy. *Biochimica et Biophysica Acta* 1994; 1226: 49-55.
- Brown MD, Torroni A, Shoffner JM, Wallace DC. Mitochondrial tRNA(Thr) mutations and lethal infantile mitochondrial myopathy. *Am J Hum Genet* 1992; 51: 446-447.

27. Simon DK, Mayeux R, Marder K, Kowall NW, Beal MF, Johns DR. Mitochondrial DNA mutations in complex I and tRNA genes in Parkinson's disease. *Neurology* 2000; 54: 703-709.
28. Ozawa T, Tanaka M, Sugiyama S, Ino H, Ohno K, Hattori K et al. Patients with idiopathic cardiomyopathy belong to the same mitochondrial DNA gene family of Parkinson's disease and mitochondrial encephalomyopathy. *Biochemical and Biophysical Research Communications* 1991; 177: 518-525.
29. Yoon KL, Aprille JR, Ernst SG. Mitochondrial tRNA (thr) mutation in fatal infantile respiratory enzyme deficiency. *Biochemical and Biophysical Research Communications* 1991; 176: 1112-1115.
30. Mayr-Wohlfart U, Paulus C, Henneberg A, Rodel G. Mitochondrial DNA mutations in multiple sclerosis patients with severe optic involvement. *Acta Neurologica Scandinavica* 1996; 94: 167-171.
31. Moraes CT, Ciacci F, Bonilla E, Ionasescu V, Schon EA, DiMauro S. A mitochondrial tRNA anticodon swap associated with a muscle disease. *Nature Genetics* 1993; 4:284-288.
32. Young WY, Zhao L, Qian Y, Li R, Chen J, Yuan H et al. Variants in mitochondrial tRNA^{Glu}, tRNA^{Arg}, and tRNA^{Thr} may influence the phenotypic manifestation of deafness-associated 12S rRNA A1555G mutation in three Han Chinese families with hearing loss. *Am J Med Genet A* 2006; 140: 2188-2197.
33. Nishino I, Seki A, Maegaki Y, Takeshita K, Horai S, Nonaka I et al. A novel mutation in the mitochondrial tRNA(Thr) gene associated with a mitochondrial encephalomyopathy. *Biochemical and Biophysical Research Communications* 1996; 225: 180-185.
34. Seki A, Nishino I, Goto Y, Maegaki Y, Koeda T. Mitochondrial encephalomyopathy with 15915 mutation: clinical report. *Pediatric Neurology* 1997; 17:161-164.
35. Yoon KL, Ernst SG, Rasmussen C, Dooling EC, Aprille JR. Mitochondrial disorder associated with newborn cardiopulmonary arrest. *Pediatric Research* 1993; 33: 433-440.
36. Chen B, Sun D, Yang L, Zhang C, Yang A, Zhu Y et al. Mitochondrial ND5 T12338C, tRNA (Cys) T5802C, and tRNA (Thr) G15927A variants may have a modifying role in the phenotypic manifestation of deafness-associated 12S rRNA A1555G mutation in three Han Chinese pedigrees. *American Journal of Medical Genetics* 2008; 146: 1248-1258.
37. Grasbon-Frodl EM, Kosel S, Sprinzl M, von Eitzen U, Mehraein P, Graeber MB. Two novel point mutations of mitochondrial tRNA genes in histologically confirmed Parkinson disease. *Neurogenetics* 1999; 2: 121-127.
38. Li R, Qu J, Zhou X, Tong Y, Hu Y, Qian Y et al. The mitochondrial tRNA(Thr) A15951G mutation may influence the phenotypic expression of the LHON-associated ND4 G11778A mutation in a Chinese family. *Gene* 2006; 376: 79-86.
39. Da Pozzo P, Cardaioli E, Malfatti E, Gallus GN, Malandrini A, Gaudiano C et al. A novel mutation in the mitochondrial tRNA(Pro) gene associated with late-onset ataxia, retinitis pigmentosa, deafness, leukoencephalopathy and complex I deficiency. *Eur J Hum Genet* 2009; 17: 1092-1096.
40. Ionasescu VV, Hart M, DiMauro S, Moraes CT. Clinical and morphologic features of a myopathy associated with a point mutation in the mitochondrial tRNA (Pro) gene. *Neurology* 1994; 44: 975-977.
41. Wong LJ, Liang MH, Kwon H, Bai RK, Alper O, Gropman A. A cystic fibrosis patient with two novel mutations in mitochondrial DNA: mild disease led to delayed diagnosis of both disorders. *American Journal of Medical Genetics* 2002; 113: 59-64.
42. Seneca S, Ceuterik-De Groote C, Van Coster R, De Meirleir L. A novel mitochondrial transfer RNA proline mutation. *Journal of Inherited Metabolic Disease* 2000; 23: 853-854.
43. Zeviani M, Servidei S, Gellera C, Bertini E, DiMauro S, DiDonato S. An autosomal dominant disorder with multiple deletions of mitochondrial DNA starting at the D-loop region. *Nature* 1989; 339: 309-311.
44. Ozawa T. Genetic and functional changes in mitochondria associated with aging. *Physiological Reviews* 1997; 77: 425-464.