# The role of autoimmunity in premature ovarian failure

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#### Abstract

Premature ovarian failure (POF) is a heterogeneous syndrome with several causative factors. Autoimmune mechanisms are involved in pathogenesis of 4-30 % of POF cases. The present review focuses on the role of autoimmunity in the pathophysiology of POF. The evidences for an autoimmune etiology are: demonstration of ovarian autoantibodies, the presence of lymphocytic oophoritis, and association with other autoimmune disorders. Several ovarian antigenic targets have been identified in POF patients. The oocyte seems to be the most often targeted cell. Lymphocytic oophoritis is widely present in POF associated adrenal insufficiency. Addisons disease is one of the most common autoimmune disorders associated with POF. Early detection of this potentially life threatening disease was recommended in several studies.

The gold standard for detecting autoimmune POF is ovarian biopsy. This procedure is not recommended due to unknown clinical value, expense, and risks. Several immunoassays have been proposed as substitute diagnostic tools. Nevertheless, there is no clinically proven sensitive and specific serum test to confirm the diagnosis of autoimmune POF or to anticipate the patient's chance of developing POF or associated diseases.

Some authors suggested the possible effects of immuno-modulating therapy on the resumption of ovarian function and fertility in a selected group of autoimmune POF patients. However, in most instances, this treatment fails to reverse the course of the disease.

Numerous studies illustrated that standard treatment outcome for infertility is less effective in the presence of ovarian autoimmunity. The antibody-induced damage

could be a pathogenic factor. Nevertheless, the precise cause remains obscure.

**Key words:** Premature ovarian failure, Premature ovarian insufficiency, Autoimmune oophoritis, Autoimmunity.

### Introduction

remature ovarian failure (POF) is a term to define the women younger than 40 years of age who present with amenorrhea lasting more than 4 months and hypoestrogenichypergonadotropic serum profile (follicle stimulating hormone (FSH) levels≥ 40 mIU/mL on two occasions) (1-5). This disorder is characterized by anovulation, amenorrhea, sex steroid deficiency, and infertility (1). The most common presentation is secondary amenorrhea and the main consequences are infertility and psychological stress (6-8). However, POF is an important cause of secondary amenorrhea and infertility, some patients may continue to ovulate and conceive "premature (7-10). Therefore. ovarian insufficiency" and "premature ovarian dysfunction" are more accurate phrases to

reflect reversible nature of this condition (11-14). The disease affects 0.3-1% of general population (6, 15-17). Follicle depletion and follicle dysfunction are two main etiological mechanisms (9, 18). Follicle depletion can be the consequence of a primitive reduced pool of oocvtes or an accelerated follicular atresia (10). FSH-receptor mutation, inappropriate luteinization related to low follicular count, and autoimmune oophoritis were suggested as the causes of follicle dysfunction (9, 19). The etiology of the disease varies in different patients: chromosomal/genetic abnormalities, metabolic/enzymatic factors, autoimmunity, infections. environmental toxins. and iatrogenic influences involve in development of the disease (1, 10, 20-22). Nevertheless, the precise cause is undetermined in a significant portion of patients. These cases were classified as idiopathic (2, 23-28). Based on clinical observations, immunological data,

and histological findings, autoimmunity may be the pathogenic mechanism in 30% of cases of idiopathic POF (23). The following article presents the role review of autoimmunity in pathogenesis of POF. obtained by MEDLINE, EMBASE, Pub Med, Google Scholar, the Cochrane Library, and hand searches of pertinent references of English literature on POF and autoimmunity, cited between the January 2000 and December 2013. Reference lists of review articles, relevant trials, immunology and gynecology textbooks, and abstracts of scientific meetings were also searched. The literature search was performed using PubMed keywords: premature ovarian failure, premature ovarian insufficiency, premature ovarian dysfunction, hyper gonadotropic hypogonadisem, autoimmune oophoritis, and autoimmunity.

### Autoimmunity and POF

The lack of a high sensitive and specific test has precluded accurate estimation of the prevalence of autoimmune type of POF (23, 29). Nevertheless, depended on some studies results, autoimmunity is responsible for approximately 4-30% of POF cases (5-33). The autoimmune involvement is base on the presence of antiovarian antibodies (AOAs), the histological evidences of lymphocytic oophoritis, and association with other autoimmune disorders (2, 18, 23, 34-42).

## AOAs

The exact mechanism of autoimmunity in pathophysiology of this disorder remains probably genetic obscure: the or environmental factors initiate the activation of immune system (5, 23). A review of literature regarding POF with autoimmune involvement reveals that the presence of autoantibodies directed against ovarian tissues and elements. and their targets in cellular and molecular levels hold important issues (23, 35, 43-45). Nevertheless, it should be mention that alteration in cellular immunity such as macrophage and dendrite cells abnormalities, change in CD4+/CD8+ ratio, as well as inappropriate expression of class II MHC antigens by granulose cells has been seen in POF patients (28, 46-48).

Vallotton and Forbes were the first to detect AOAs in sera of POF patients (49). After that, numerous studies showed antibody-mediated mechanism for ovarian involvement (2, 18, 35, 36, 45). The presence of these antibodies is in association with increased risk of ovarian failure independent of reproductive hormonal levels (50). AOAs may be detected in serum prior to the onset of clinical presentation of POF (51).

The antibodies binding to the various steroid hormone-producing cells (52-56), gonadotropins and their receptors (57-60), zona pellucida (61, 62), oocyte (63, 64), corpus luteum (23, 65, 66), and several other antibodies such as anticardiolipin and antinuclear antibodies (34, 43) have been reported as the markers of ovarian autoimmunity. The multiplicity of the suspected auto antigens and related antibodies illustrates the variety of pathologic process causing ovarian damage.

The antibodies directed against steroidproducing cells of various endocrine glands such as adrenal cortex cells, Leydig cells of the testis, syncytiotrophoblast of the placenta, and theca cells of the ovary named as steroid cell antibodies (StCAs). These antibodies are polyclonal immunoglobulins of the IgG class involved (67). Some enzymes in steroidogenesis pathways are the targets of these autoantibodies. The main antigenic targets of StCAs are P450-17α-hydroxylase (17α-OH). P450-side-chain cleavage (P450scc), and 21- hydroxylase (21-OH) (68). StCAs are present in some autoimmune diseases such as autoimmune polyendocrine syndromes (APS); type I, type II, Addison's disease (AD), and POF (35, 69). Their prevalence is 60% in APS-I patients, 25-40% in APS-II patients, 60-87% in POF associated AD, and 3-10% in isolated POF (35, 43, 54-56). Reato et al. suggested that primary amenorrhea in association with StCAs leads to autoimmune background of the ovarian failure (55). Some studies demonstrated that the presence of these autoantibodies is a predictive marker for developing POF in patients with autoimmune AD (23, 55). It is likely that 17α-OH and P450scc are the main molecular targets of StCAs in sera positive patients with POF associated AD (23, 35, 54, 55. 68). Nevertheless. in sera of approximately 10% of these patients, neither nor P450scc 17α-OH antibodies were detected (35). This observation illustrates the presence of some unidentified autoimmune targets for StCAs. Falroni and colleagues

found that the women with AD related POF are often (>91%) positive for one of three markers major immune of steroid-cell autoimmunity [17a-OH antibodies, P450scc antibodies. 3B-Hydroxysteroid and dehydrogenase (3β-HSD) antibodies]. In their study, only 3% of the patients with isolated POF are positive for these markers (35). These data are consistent with other studies that StCAS are not major antibodies in isolated POF or POF associated with nonautoimmune disease. adrenal These autoantibodies seem to be main serologic markers for ovarian failure in AD related POF patients (23, 54. 70-72). Falroni and colleagues also showed that 3B-HSD is not a major auto antigen in autoimmune POF. They concluded that autoantibodies against this enzyme have limited application in routine clinical practice. In fact, the presence of 3β-HSD autoantibodies could be the secondary consequence of activation of immune system (35). In contrast with this study, Arif et al. suggested that 3β-HSD autoantibodies have a higher diagnostic sensitivity than other StCAs for autoimmune POF (73). The discrepancy between these studies could be explained in differences part by the in laboratory techniques used to measure antibodies; radio binding assay with in vitro translated recombinant 35S-labelled auto antigens for Falroni and immunoblotting technique for Arif (35, 73).

In earlier published studies, some autoantibodies have been detected on the surface of granulose cells bv indirect immunofluoresence. The researchers have suggested that gonadotropin receptors are the targets for these antibodies (30, 63, 74). The inhibition of biological activity of gonadotropins and the decline in binding capacity of gonadotropins to their receptors are two main suggested mechanisms (75, 76). The findings of the earliest studies regarding the role of these autoantibodies in development of autoimmune POF were not supported by subsequent researches (58, 77). In other way, anti-gonadotropin receptor antibodies were also detected in iatrogenic ovarian failure (37). In fact, the real significance of these antibodies remains to be determined (78).

In order to detect autoantibodies directed against gonadotropins, some researchers investigated serum factors that might be able to inhibit FSH or Luteinizing hormone (LH) activity. Gobert and colleagues demonstrated the existence of anti- $\beta$ FSH antibodies in sera of POF patients (57), their clinical significance, and diagnostic relevance still needs to be investigated through well-designed diagnostic studies.

Zona pellucida (ZP) is an extracellular matrix surrounding the mammalian oocyte. Its molecular structure consists of glycoproteins with strong antigenic potency. Takamizawa et al. detected anti-ZP antibodies in sera from idiopathic POF patients. They introduced a new microdot assay with high specificity for detecting anti-ZP antibodies (79). Their results are in agreement with the various independent reports on the existence of anti-ZP antibodies (61, 62, 80). The suspected pathological effect is the impaired communication between oocyte and granulose cells (62). At present time, there are no well-designed diagnostic studies to demonstrate the prevalence of these antibodies in POF patients. Thus, the exact importance of these autoantibodies is still unclear.

Anti-oocyte cytoplasm antibodies have been detected in patients with POF (63, 64). Pires and his group demonstrated that the cytoplasm of oocyte probably contains the most autoimmune targets in POF patients (78, 81). Although, the exact nature of the antigenic targets are still unclear, MATER (Maternal Antigen That Embryo Require), a 125KDa protein may be a possible candidate (26, 82-84). Very little is known about the precise nature of this protein. We need further studies to provide information about MATER and aid in deciphering its role in ovarian biology. The other identified antigens are (ALDH1AI), Aldehyde dehydrogenase1A1 Selenium Binding Protein 1 (SBP1), αenolase, and Heat Shock Protein 90 (HSP90) (70, 81, 85, 86). According to Pires et al. study, HSP90ß is the most immunodominant antigen (78, 81). In contrast with these studies, the autoantibodies against oocyte were also found in sera of healthy individuals and a number of neoplastic, autoimmune, and inflammatory diseases (86-91).

In various studies, AOAs were detected by different methods (eg; enzyme-linked immunosorbent assay, immunohistochemistry, Western blotting, and indirect immunofluorescence). The most common used methods are indirect immunofluorescence (IIF) and enzyme-linked

immunosorbent assay (ELISA) (23). Few studies have considered the measuring of autoantibodies against 21-OH by immunoprecipitation assay equivalents to measurement of these autoantibodies by (IIF) (92, 93). In the other way, Novosad *et al.* showed that validity of immunofluorescence for detecting AOAs is questionable (29).

The prevalence of AOAs in POF women varies greatly, ranging from 3 to 66.6 percent (23, 28, 35, 94, 95). These conflicting results could be explained by the difference in design elements of studies such as criteria for study and comparison groups, sample size, method of antigen preparation and antibody detection (2, 10, 18, 23, 94, 96). We suppose that the assessment of only one of several antigenic targets in some of studies, the differences in antigen sources, heterogeneity of histological targets, selection of animal or human ovaries at different ages or at different periods of menstrual cycle, and transient appearance of some of antiovarian antibodies have also changed the outcome of above-mentioned experiences.

AOAs might be the main cause of pathogenesis of autoimmune POF, or might be the result of ability of antibodies production by this autoimmune disease. In the other hand, there is poor correlation between antibody levels and severity of disease (18). Some authors conclude that clinical diagnosis of autoimmune POF, exclusively based on the detection of presence or absence of AOAs will be a great mistake (2).

A major weakness in the assessment of the role of AOAs is the high rate of false positive results (poor specificity) (29). These autoantibodies could be found in significant numbers of control groups (13, 29). It might be due to the presence of naturally occurring antibodies (NAA) (97).

At present time, there is no valid serum marker to certainly prove the diagnosis of autoimmune POF.

## Lymphocytic oophoritis and POF

The histopathological evidences of autoimmune ovarian involvement have been illustrated in 9.1-11% of the samples of ovarian biopsies belong to normal karyotype women with hyper gonadotropic amenorrhea (37, 40). Autoimmune oophoritis is usually detected in AD associated POF patients. Rarely, this pathologic picture was seen in association with isolated POF (14). Cellular infiltration of follicles by macrophages, natural killer cells, T-lymphocytes, plasma cell, and Blymphocytes is the characteristic sign of an autoimmune oophoritis. The steroid-producing cells are the main target of autoimmune attack (18, 98). In some cases, abnormal activation of epithelial cells and subsequent follicular depletion and fibrosis were seen (98). Nevertheless, the specific histological picture is sparing of primordial and primary follicles and involvement of developing follicles by lymphocytic infiltration of theca cells (56, 99, 100). With respect to the multifollicular appearance of ovaries in this condition, the size of the involved ovaries could be normal or enlarged on sonographic view (56). Although, follicular depletion is the final stage of this autoimmune attack, the histological samples of ovarian tissues reveal the developing follicles in varying sizes in the majority of patients (101, 102). This observation supports the positive effect of immunosuppressive agents in restoring ovarian functions.

Massin and colleagues showed pelvic ultrasonography and hormonal profile (FSH, LH, estradiol, inhibin B serum levels) are insufficient to predict the presence or absence of follicular structures in POF patients with Thev recommended normal karyotype. ovarian biopsy as the reliable assessment for detecting follicle presence and activity in these patients (102). Ovarian biopsy is the gold standard for detecting autoimmune involvement of ovarian tissue and the presence of developing follicles in involved 37, 40). Nevertheless, ovary (28, this procedure is invasive and expensive. In other way, few studies demonstrated that the ability of ovarian biopsy in representing of follicular density of the whole ovary is questionable (10, 14, 103). We need a noninvasive screening test to avoid unnecessary ovarian biopsy and to select the women with possibility of autoimmune oophoritis. The antibodies to steroid-producing cells have been frequently found in the patients with histological evidences of lymphositic infiltration of the ovarian tissue (37, 54). Bakalov et al. showed a positive correlation between the presence of autoimmune oophoritis and serum adrenal cortex antibodies. In light of this observation, they recommended the measuring of adrenal cortex autoantibodies in sera of POF patients to select the patients with possibility of

autoimmune oophoritis (56). Unfortunately, we did not find the published placebo-controlled randomized clinical trials with appropriate powers to support this finding.

# POF in association with other autoimmune diseases

Ovary is a common target of autoimmune attack in organ-specific and svstemic autoimmune diseases (18, 23, 54, 89, 104, 105). Approximately 10%-55% of patients with POF have associated autoimmune disorders (18, 23). Hypothyroidism is the most common associated autoimmune disorder with POF (25-60%) (3, 106, 107). Coincidence with diabetes mellitus is 2.5% (18). Autoimmune adrenal insufficiency may present as an isolated disease or may be associated with autoimmune disorders other (68). Approximately 10-20% of Addison patients have POF, and 2.5-20% of women with POF showing some evidences of autoimmunity against the adrenal gland (18, 55, 108). Dal Pra et al. showed no difference in the age at onset of POF in patients with or without Addison's disease (54). Autoimmune POF usually occurs before the onset of adrenal involvement (23). In Reato et al. study, AD preceded POF. Nevertheless, the mean ages at onset of each disease were close (27 years for AD and 28.5 years for POF) (55). The association of POF with AD could be due to the presence of cross-reacting autoantibodies that react against auto antigens common to steroid producing cells with different origins (54, 55, 108). StCAs were detected in 60-87% of POF-associated adrenal involvement and 3-10% of patients with isolated form of this disorder (35, 37, 43, 52-56). The detection of antibodies against  $17\alpha$ -OH and P450scc in Addison patients is a predictor factor of for development of POF (68, 92). In the other hand, some authors suggested that the presence of 21-OH antibodies in sera from women with autoimmune POF might be important marker in identifying patients at risk developing of autoimmune adrenal insufficiency (35, 54, 68, 92). As already recognized by above mentioned reports, the POF patients with positive sera for StCAs are at risk for adrenal insufficiency, a potentially fatal condition, especially during pregnancy (10, 13, 35, 68, 108). Untreated adrenal insufficiency could be associated with serious fetal and maternal complications such as crisis postpartum adrenal (13, 109). Therefore, identification of this subgroup of patients POF with subclinical adrenal insufficiency is essential before decision making for egg or embryo donation. Bakalov et al. suggested that the morning serum cortisol level has low sensitivity and specificity, when used as a screening test in POF patients. Based on their findings, the measuring of adrenal cortex autoantibodies by indirect immunofluorescence technique or antibodies against 21-OH by immunoprecipitation assay are effective screening tests to detect asymptomatic adrenal insufficiency in these patients (108). It is in agreement with the previous reports on this subject (71, 92, 108). They also recommended that the presence of adrenal insufficiency could be confirmed by the standard ACTH stimulation test (108). We suggest referral of POF patients with positive adrenal cortex antibodies to endocrinologist for additional evaluations and long-term follow-up of adrenal functions. We also propose routine screening of thyroid function and glucose tolerance in POF patients. Due to the low frequency of the other associated wider autoimmune diseases. endocrine function screening is not recommended.

Autoimmune polyglandular syndromes (APS) are a series of disorders characterized by autoimmunity against two or more endocrine organs (10, 27). APS-I is a rare autosomal recessive disorder caused by mutation in the AIRE (autoimmune regular) gene. The role of this gene is regulation of immune tolerance (105). POF develops in 41-72% of patients with APS type I (13, 18, 55, 101, 104). Gonadal failure tends to appear at a younger age and in the highest prevalence compared with the other forms of APS (54, 55, 68). This event could be due to the mutations of AIRE gene in patients with APS-I (54, 55, 68). APS-II is an autosomal dominant disease. The prevalence of ovarian failure in APS-II is 10-25% (55, 105, 110). Autoantibodies directed against steroidogenic enzymes and ovarian steroid-producing cells mediate ovarian dysfunction (111). In general, AD precedes POF in patients with APS-I, and follows POF in those with APS-II (55). Reato and colleagues suggested that StCAs,  $17\alpha$ -OHAb, and P450sccAb are the immunological predictive markers of potential autoimmune POF in the patients with autoimmune Addison's disease and the different types of APS (55).

A review of literature regarding the relation between POF and autoimmunity indicates that autoimmune involvement is the main mechanism of POF in cases associated with other autoimmune diseases, but in isolated POF the evidences supporting this subject is weak (23, 43). However, the higher anti-Mullerian hormone and inhibin levels, a low incidence of histological evidences of oophoritis. and frequent reports of spontaneous pregnancy isolated in autoimmune POF could be the best evidences of transient ovarian damage and some degrees of function preservation (24, 112, 113). We need the placebo-controlled randomized clinical trials to determine the true roles of autoimmunity in pathogenesis of isolated POF.

### Infertility and autoimmune POF

The involvement of autoimmune mechanisms have been suggested in various ovarian pathology, such as polycystic ovarian syndrome, unexplained infertility, recurrent pregnancy loss, endometriosis, and other conditions with poor reproductive outcome (14, 23, 30, 43, 51, 114).

Numerous studies illustrated that standard treatment outcome for infertility is less effective the presence of ovarian in autoimmunity. The presence of anti-oocyte cytoplasm antibodies in follicular fluid was associated with unsuccessful oocyte retrieval and fertilization failure in IVF patients (115). studies demonstrated association Some between the presence of serum AOAs and lower pregnancy rate in women seeking infertility treatment (34, 51, 116). Furthermore, a high correlation was seen between poor response to IVF programs and the presence of AOAs in sera of infertile women (34, 117, 118). Monnier-Barbarino and colleagues suggested that hormonal stimulation and ovarian trauma during follicular puncture could be the causes of developing or enhancing of AOAs formation in patients with history of repeated IVF attempts (117). In another study, the immunization process against exogenous gonadotropins and the development of anti-FSH and anti-LH antibodies were considered as the cause of poor responding to IVF program (118). In the other hand, some authors suggested that anti-ZP antibodies are one of the causes of IVF failure by their blocking effects on sperm-ZP binding and sperm penetration (79, 119-122). In light of this observation. Mardesic et al. intracytoplasmic recommended sperm injection (ICSI) as the method of choice in anti-ZP antibody-positive infertile couples (123). Pires proposed the evaluation of AOAs as a prognostic factor in patients undergoing IVF-ET program (78). Later on, Khole recommended that serological assessment of AOAs before starting IVF protocols would be justified in poor responder patients (2).

With respect to the possibility of the presence of AOAs in sera of POF women vears before the onset of clinical presentation as well as detection of these autoantibodies in one third of patients with unexplained infertility (95), some authors suggested this form of infertility as an early stage of autoimmune POF. They also recommended the evaluation of ovarian autoimmunity as part of an infertility workup to predict the chance of development of gonadal failure in future and identify the best candidates for oocyte retrieval before follicular depletion (23, 70). In fact, we have little information about the predictive value of AOAs in ovarian reserve assessment. We need well-designed studies to clarify this subject.

# The effect of Immunosuppressive treatments

Lymphocytic infiltration of theca layer of developing follicles and sparing of primary and primordial follicles presents the possibility of resumption of ovarian function by immunosuppressive treatments (99, 100).

Although in most instances, treatment with immunosuppressive agents failed to reverse the course of the ovarian autoimmunity or enhance ovarian response the to gonadotropins (18, 124). Numerous studies have recommended cell-mediated and humoral immunity suppression by glucocorticoids or anti-B-cell therapies for reversal of infertility or resumption of ovarian function in the selected groups of patients with autoimmune POF (101, 111, 125-128). It should be in mind that none of these studies was randomized controlled trial with wellselected autoimmune POF patients (129). Indeed, these reports were mostly case series studies (127, 130, 131). On the other hand, some patients developed serious complications such as Cushing syndrome and after knee osteonecrosis receiving corticosteroids (132). Bats et al. recommended tissue diagnosis of local autoimmune reaction by ovarian biopsy before starting immunosuppressive agents (98). Although, the gold standard for detecting autoimmune POF is ovarian biopsy (28, 37, 40), this procedure is not recommended due to unknown clinical value, expense, and risks (10, 14, 103). With respect to the association between the presence of adrenal cortex antibodies and autoimmune oophoritis, the measuring of these autoantibodies in serum has been proposed as noninvasive screening test to avoid unnecessary ovarian biopsy and select the patients with possibility of autoimmune oophoritis (56).

Dehydroepiandrosterone (DHEA) is an endogenous steroid agent. Zona reticularis layer of the adrenal cortex and ovarian theca cells are sources of secretion in healthy women. This molecule is precursor of testosterone, androstenedione, and estradiol (133). DHEA promotes activation of the oocytes and inhibits atretic phenomena (134). Barad et al. noted higher pregnancy rates with DHEA supplementation in patients with diminished ovarian function (134, 135). Mamas and Mamas reported successful pregnancy following DHEA therapy in women with POF (130). Barad and colleagues believed that the patients in this study did not fulfill the criteria of POF. They recommend the termination of "premature ovarian aging" (POA) for definition of these patients with increased FSH levels and diminished ovarian reserve (135). We did not find a specific treatment modality for autoimmune oophoritis with proven efficacy and safety confirmed by prospective randomized placebo-controlled studies.

Other suggested fertility options in POF patients are IVM (in vitro maturation) of oocyte derived from primordial follicles or stem cells

and IVF (in vitro fertilization) using donor gametes or embryos (10, 136).

# Conclusion

In conclusion, an abundance of evidence shows that autoimmunity is responsible for development of POF especially in cases associated with other autoimmune diseases. Several potential immune targets in various parts of ovary have been proposed for antibody-mediated autoimmune attack. The oocyte seems to be the most often targeted cell. Despite the considerable scientific work, the precise nature of molecular targets responsible for this autoimmune process remained unclear. The studies regarding antiovarian antibodies have conflicting results, partly due to differences in the detection methods, heterogeneity of the patients and control groups, and the lack of high specificity and sensitivity tests. This fact illustrates the need for accurate diagnostic tools in order to identification the specific antigenic targets and related autoantibodies.

Numerous studies illustrated that standard treatment outcome for infertility is less effective in the presence of ovarian autoimmunity. Subsequently, some authors suggested the possible effects of immunomodulating therapy on the resumption of ovarian function and fertility in a selected group of POF patients. Nevertheless, in most instances, this treatment fails to reverse the course of the disease.

further information regarding The autoimmune actions of AOAs will lead to elucidate the underlying mechanisms of ovarian damage. It also will help the development of better approaches to diagnose, and effective therapeutic regiments. We also suppose that the specific noninvasive tests are essential to detect associated disorders, as well as to select the women with acceptable chance of restoring of ovarian function and fertility by immunosuppressive therapy.

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### **Conflict of interest**

The authors declare there are no conflicts of interest.

### References

- 1. Ebrahimi M, Akbari Asbagh F. Pathogenesis and causes of premature ovarian failure: An update. *Int J Fertil Steril* 2011; 5: 54-65.
- 2. Khole V. Does ovarian autoimmunity play a role in the pathophysiology of premature ovarian insufficiency? *J Midlife Health* 2010; 1: 9-13.
- Rees M, Purdie D. Premature menopause. In: Rees M, Purdie D, editors, Management of the menopause: the handbook. London: Royal Society of Medicine Press Ltd; 2006; 142-149.
- 4. Pal L, Santoro N. Premature ovarian failure (FOF): discordance between somatic and reproductive aging. *Ageing Res Rev* 2002; 1: 413-423.
- Nelson LM. Primary ovarian insufficiency. N Engl J Med 2009; 360: 606-614.
- 6. The Practice Committee of the American Society for Reproductive Medicine. Current evaluation of amenorrhea. *Fertil Steril* 2006; 86: 148-155.
- 7. Taylor AE . Systemic adversities of ovarian failure. *J* Soc Gynecol investing 2001;8: S7-S9.
- Akbari asbagh F, Ebrahimi M. A case report of spontaneous pregnancy during hormonal replacement therapy for premature ovarian failure. *Iran J Reprod Med* 2011; 9: 47-49.
- Nelson LM, Bakalov VK. Mechanisms of follicular dysfunction in 46,XX spontaneous premature ovarian failure. *Endocrinol Metab Clin North Am* 2003; 32: 613-637.
- Goswami D, Conway GS. Premature ovarian failure. *Hum Reprod update* 2005; 11: 391-410.
  Albright F, Smith PH, Fraser R. A syndrome
- Albright F, Smith PH, Fraser R. A syndrome characterized by primary ovarian insufficiency and decrease stature. *Am J Med Sci* 1942; 204: 625-648.
- 12. Alzubadi NH, Chapin HL, Vanderhoof VH, Calis KA, Nelson LM. Meeting the needs of young women with secondary amenorrhea and spontaneous ovarian failure. *Obstet Gynecol* 2002; 99: 720-725.
- Panay N, Kalu E. Management of premature ovarian failure. Best Prac Res Clin Obstet Gynecol 2009; 23: 129-140.
- Dragojevic-Dikic S, Marisavljevic D, Mitrovic A, Dikic S, Jovanovic T, Jankovic-Raznatovic S. An immunological insight into premature ovarian failure (POF). Autoimmune Rev 2010; 9: 771-774.
- Luborsky JL, Meyer P, Sowers MF, Gold EB, Santoro N. Premature menopause in a multi ethnic population study of the menopause transition. *Hum Reprod* 2003; 18: 199-206.
- 16. Beck-Peccoz P, Persani L. Premature ovarian failure. Orphanet J Rare Dis 2006; 6: 1-9.
- 17. Nolting M, Perez L. Belen M. Martinez Amuchastegui J. Straminsky V, Onetto Claudia C, *et al.* Importance

of detecting transition to the premature ovarian failure. *Gynecol Endocrinol* 2008; 24: 257-258.

- Lebovic D I, Naz R. Premature ovarian failure: Think "autoimmune disorder". Sex Reprod Menopause 2004; 2: 230-233.
- Aittomaki K, Lucena JL, Pakarinen P, Sistonen P, Tapanainen J, Gromoll J, *et al.* Mutation in the follicle-stimulating hormone receptor gene cause hereditary hypergonadotropic ovarian failure. *Cell* 1995; 82: 959-968.
- 20. Laml T, Preyer O, Umek W, Hengstschlager M, Hanzal H. Genetic disorders in premature ovarian failure. *Hum Reprod Update* 2002; 8: 483-491.
- 21. Schlessinger D, Herrera I, Crisponi I, Mumm S, Percesepe A, Pellegrini M *et al.* Genetic and translocations involved in POF. *Am J Med Genet* 2002; 111: 328-333.
- 22. Matikainen T, Perez GI, Jurisicova A, Pru JK, Schlezinger JJ, Ryu HY, *et al.* Aromatic hydrocarbon receptor-driven Bax gene expression is required for premature ovarian failure caused by biohazardous environment chemical. *Nat Genet* 2001; 28: 355-360.
- 23. Forges T, Monnier-Barbarino P, Faure GC, Bene M C. Autoimmunity and antigenic targets in ovarian pathology. *Hum Reprod Update* 2004; 10: 163-175.
- 24. Rebar WR. Premature ovarian failure. Obstet Gynecol 2009; 113: 1355-1363.
- 25. Meskhi A, Seif MW. Premature ovarian failure. *Curr Opin Obstet Gynecol* 2006; 18: 418-426.
- Tuohy VK, Altuntas CZ. Autoimmunity and premature ovarian failure. *Curr Opin Obstet Gynecol* 2007; 19: 366-369.
- Conway SG, Christin- Maitre S. Premature ovarian failure. In: Fauser CJB, editor. Reproductive medicine molecular, cellular, and genetic fundamentals, New York: Parthenon Publishing: 2003; 587-599.
- 28. Yan G, Schoenfeld D, Penney C, Hurxthal K, Taylor AE, Faustman D. Identification of premature ovarian failure patients with underlying autoimmunity. *J* womens health Gend Based med 2000; 9: 275-287.
- 29. Novosad JA, Kalantaridou SN, Tong ZB, Nelson LM. Ovarian antibodies as detected by indirect immunofluorescence are unreliable in the diagnosis of autoimmune premature ovarian failure. A controlled evaluation. *BMC Womens Health* 2003; 13: 2-9.
- 30. Luborsky JL. Ovarian autoimmune disease and ovarian autoantibodies. *J Women's Health Gend Based Med* 2002; 11: 585-599.
- Coulam CB, Adamson SC, Annegers JF. Incidence of premature ovarian failure. *Obstet Gynecol* 1986; 67: 604-606.
- 32. de Moraes Ruehsen M, Blizzard RM, Garcia-Bunuel R, Jones GS. Autoimmunity and ovarian failure. *Am J Obstet Gynecol* 1972; 112: 693-703.
- 33. Kamada M, Hasebe H, Irahara M, Kinoshita T, Naka O, Mori T. Detection of anti-zona pellucid activities in human era by the passive hemaglutination reaction. *Fertil Steril* 1984; 41: 901-906.
- 34. Pires ES, Meherji PK, Vaidya RR, Parikh FR, Ghosalkar MN, Khole VV. Specific and sensitive immunoassays detect multiple anti-ovarian antibodies in women with infertility. *J Histochem Cytochem* 2007; 55: 1181-1190.
- 35. Falorni A, Laureti S, Candeloro P, Perrino S, Coronella C, Bizzarro A, *et al.* Steroid-cell

autoantibodies are preferentially expressed in women with premature ovarian failure who have adrenal autoimmunity. *Fertil Steril* 2002; 78: 270-279.

- Falorni A, Laureti S, Santeusanio F. Autoantibodies in autoimmune polyendocrine syndrome type II. Endocrinol Metab North Am 2002; 31: 369-389.
- 37. Hoek A, Schoemaker J, Drexhage HA. Premature ovarian failure and ovarian autoimmunity. *Endocr Rev* 1997; 18: 107-134.
- Sedmak DD, Hart WR, Tubbs RR. Autoimmune oophoritis: a histopathologic study of involved ovaries with immunologic characterization of the mononuclear cell infiltrate. *Int J Gynecol Pathol* 1987; 6: 73-81.
- 39. Hill JA, Welch WR, Faris HM, Anderson DJ. Induction of class II major histocompatibility complex antigen expression in human granulose cells by interferon gamma: a potential mechanism contributing to autoimmune ovarian failure. *Am J Obstet Gynecol* 1990; 162: 534-540.
- 40. Spremovic-Radienovic S. Laparoscopic ovarian biopsy in women with premature ovarian failure. 8th European congress on Menopause (EMAS). *Maturitas* 2009; 63: S1-136.
- Poppe K, Velkeniers B. Female infertility and thyroid. Best Pract Res Clin Endocrinol Metab 2004; 18: 153-165.
- 42. Poppe K, Glinoer D, Toumaye H, Devroey P, Schiettecatte J, Haentijens P, *et al.* Thyroid autoimmunity and female infertility. *Verh K Acad Geneeskd Belg* 2006; 68: 357-377.
- 43. Cervera R, Balasch J. Bidirectional effects on autoimmunity and reproduction. *Hum Reprod Update* 2008; 14: 359-366.
- 44. Tung ZB, Gold L, Pfeifer KE, Dorward H, Lee E, Bondy CA, *et al.* Mater, a maternal effect gene required for early embryonic development in mice. *Nat Genet* 2000; 26: 267-268.
- 45. Monnier-Barbarino P, Forges T, Faure GC, Bene MC. Gonadal antibodies interfering with female reproduction. *Best Pract Res Endocrinol Metab* 2005; 19: 135-148.
- 46. Chernyshov VP, Radysh TV, Gura IV, Tatarshuk TP, Khominskaya ZB. Immune disorders in women with premature ovarian failure. *Am J Reprod Immunol* 2001: 46; 220-225.
- 47. Tung KSK, Garza KM, Lau Y, Bagavant H. Autoimmune ovarian disease:mechanism of induction and prevention. *J Soc Gynecol Invest* 2001; 8: 49-51.
- 48. Wu R, Van der Hook KH, Ryan NK, Norman RJ, Robker RL. Macrophage contributions to ovarian function. *Hum Reprod Update* 2004; 10: 119-133.
- 49. Vallotton MB, Forbes AP. Antibodies to cytoplasm of ova. *Lancet* 1996; 2: 264-265.
- 50. Luborsky J L, Roussev R, Coulam C. Ovarian antibodies, FSH and inhibin are markers associated with unexplained infertility. *Hum Reprod* 2000; 15: 1046-1051.
- 51. Luborsky J, Pong R. Pregnancy outcome and ovarian antibodies in infertility patients undergoing controlled ovarian hyperstimulation. *Am J Reprod Immunol* 2000; 44: 261-265.
- 52. Lawrence M. Primary ovarian insufficiency. N Eng J Med 2009; 360: 606-614.
- 53. Betterle C, Volpato M. Adrenal and ovarian autoimmunity. *Eur J Endocinol* 1998; 138: 16-25.

- 54. Dal Pra C, Chen S, Furmaniak J, Rees Smith B, Pedini B, Moscon A, *et al.* Autoantibodies to steroidogenic enzymes in patients with premature ovarian failure with and without Addison's disease. *Eur J Endocrinol* 2003; 148: 565-570.
- 55. Reato G, Morlin L, Chen S, Furmaniak J, Smith B Rees, Masiero S, *et al.* Premature ovarian failure in patients with autoimmune Addison, s disease: clinical, genetic, and immunological evaluation. *J Clin Endocrinol Metab* 2011; 96: 1255-1261.
- 56. Bakalov VK, Anasti JN, Calis KA, Vanderhoof VH, Premkumar A, Chen S, *et al.* Autoimmune oophoritis as a mechanism of follicular dysfunction in women with 46, XX spontaneous premature ovarian failure. *Fertil Steril* 2005; 84: 958-965.
- 57. Gobert B, Jolivet-Reynaud C, Dalbon P, Barbario-Monnier P, Faure G, Jolivet M, *et al.* An immunoreactive peptide of the FSH involved in autoimmune infertility. *Biochem Biophys Res Commun* 2001; 289: 819-824.
- 58. Anasti JN, Flack MR, Froethlich J, Nelson LM. The use of recombinant gonadotropin receptors to search for immunoglobulin G-mediated premature ovarian failure. *J Clin Endocrinol Metab* 1995; 80: 824-828.
- 59. Ryan MM, Jones HR Jr. Myasthenia gravis and premature ovarian failure. *Muscle Nerve* 2004; 30: 231-233.
- 60. Chiauzzi VA, Bussmann I, Calvo JC, Sundblad V, Charreau EH. Circulating immunoglobulins that inhibit the binding of follicle stimulating hormone receptor: A putative diagnostic role resistant ovarian syndrome. *Clin Endocrinol (Oxf)* 2004; 61: 46-54.
- Koyama K, Hasegawa A. Premature ovarian failure syndrome may be induced by autoimmune reactions to zona pellucid proteins. *J Reprod Endocrinol* 2006; 3: 94-97.
- 62. Kinoshita A, Tanaka H, Komori S, Hasegawa A, Koyoma K. Autoimmunity to zona pellucida possibly causes premature ovarian failure (POF). *J reprod Immunol* 2006; 71: 155.
- 63. Damewood MD, Zacur HA, Hoffman GJ, Rock JA. Circulating antiovarian antibodies in premature ovarian failure. *Obstet Gynecol* 1986; 68: 850-854.
- 64. Luborsky JL, Visintin I, Boyers S, Asari T, Caldwell B, DeCherney A. Ovarian autoantibodies detected by immobilized antigens immunoassay in patients with premature ovarian failure. *J Clin Endocrinol Metab* 1990; 70: 69-75.
- Paoto SG ,Viana VS, Mendonca BB, Yoshinari NH, Bonfa E. Anti-corpus luteum antibody: a novel serological marker for ovarian dysfunction in systemic lupus erythematosus? *J Rheumatol* 1999; 26: 1087-1093.
- 66. Sotsiou F, Bottazzo GF, Doniach D. Immunofluorescence studies on autoantibodies to steroid-producing cells, and to germline cells in endocrine disease and infertility. *Clin Exp Immunol* 1980; 39: 97-111.
- 67. Anderson JR, Goudie RB, Gray KG, Stuart Smith DA. Immuonological features of idiopathic Addison's disease: an antibody to cells producing steroid hormones. *Clin Exper Immunol* 1968; 3: 107-110.
- 68. Betterle C, Dal Pra C, Mantero F, Zanchetta R. Autoimmune adrenal insufficiency and autoimmune polyendocrine syndromes: autoantibodies, autoantigens and their applicability in diagnosis and disease prediction. *Endocr Rev* 2002; 23: 327-364.

- 69. Perniola R, Falorni A, Clemente MG, Forini F, Accogli E, Lobergelio G. Organ-specific and nonorgan-specific autoantibodies in children and young adult with autoimmune polyendocrinopathy candidiasis-ectodermal dystrophy (APECED). *Eur J Endocrinol* 2000; 43: 97-503.
- 70. Edassery SL, Shatavi SV, Kunkel JP, Hauer Ch, Brucker C, Penumatsa K, et al. Autoantigens in ovarian autoimmunity associated with unexplained infertility and premature ovarian failure. *Fertil Steril* 2010; 94: 2636-2641.
- 71. Chen S, Sawicka J, Betterle C, Powell M, Prentice L, Volpato M, *et al.* Autoantibodies to steroidogenic enzymes in autoimmune polyglandular syndrome, Addison's disease, and premature ovarian failure. *J Clin Endocrinol Metab* 1996; 81: 1871-1876.
- 72. Reimand K, Peterson P, Hyoty H, Uibo R, Cooke I, Weetman AP, *et al.* 3beta-hydroxysteroid dehydrogenase autoantibodies are rare in premature ovarian failure. *J Clin Endocrinol Metab* 2000; 85: 2324-2326.
- 73. Arif S, Vallian S, Farzaneh F, Zanone MM, James SL, Pietropaolo M, et al. Identification of 3β– hydroxysteroid dehydrogenase as a novel target of steroid cell antibodies association of autoantibodies with endocrine autoimmune disease. *J Clin Endocrinol Metab* 1996; 81: 4439-4445.
- 74. Scully RE, Mark EG, McNeely BU. Case records of the Massachusetts General Hospital. N Engl J Med. 1986; 315: 1336-1343.
- 75. Van Weissenbruch MM, Hoek A, Van Vliet-Bleeker I, Schoemaker J, Drexhage H. Evidence for existence of immunoglobulins that block ovarian granulose cell growth in vitro. A putative role in resistance ovary syndrome? J Clin Endocrinol Metab 1991; 73: 360-367.
- 76. Chiauzzi V, Cigorraga S, Escobar ME, Rivarola MA, Charreau EH. Inhibition of follicle-stimulating hormone receptor binding by circulating immunoglobulins. J Clin Endocrinol Metab 1982; 54: 1221-1228.
- 77. Wheatcroft NJ, Toogood AA, Li TC, Cooke D, Weetman AP. Detection of antibodies to ovarian antigens in women with premature ovarian failure. *Clin Exp Immunol* 1994; 96: 122-128.
- 78. Pires E S. Multiplicity of molecular and cellular targets in human ovarian autoimmunity: an update. *J Assist Reprod Genet* 2010; 27: 519-524.
- 79. Takamizawa S, Shibahara H, Shibayama T, Suzuki M. Detection of antizona pellucida antibodies in the sera from premature ovarian failure patients by a highly specific test. *Fertil Steril* 2007; 88: 925-932.
- 80. Smith S, Hosid S. Premature ovarian failure associated with autoantibodies to the zona pellucida. *Int J Fertil Menopausal Stud* 1994; 39: 316-319.
- 81. Pires ES, Khole VV. A block in road to fertility: Autoantibodies to heat–shock protein 90-beta in human ovarian autoimmunity. *Fertil Steril* 2009; 92: 1395-1409.
- Nelson LM. Autoimmune ovarian failure: comparing the mouse model and the human disease. J Soc Gynecol Invest 2001; 8: S55-S57.
- 83. Tong ZB, Nelson LM. Mater encodes a maternal protein in mice with a leucine–rich repeat domain homologous to porcine ribonuclease inhibitor. *Mamm Genome* 2000; 11: 281-287.

- 84. Tong ZB, Gold L, De Pol A, Vanevski A, Dorward H, Sena P, et al. Developmental expression and subcellular localization of mouse MATER, an oocytespecific protein essential for embryo development. *Endocrinol* 2004; 145: 1427-1434.
- 85. Sundblad V, Bussmann L, Chiauzzi VA, Pancholi V, Charreau EH. Alpha-enolase: a novel autoantigen in patients with premature ovarian failure. *Clin Endocrinol (Oxf)* 2006; 65: 745-751.
- 86. Vidal CI, Mintz PJ, Lu K, Ellis LM, Manenti L, Chiavazzi R, et al. An HSP90-mimic peptide revealed by fingerprinting the pool of antibodies from ovarian cancer patients. *Oncogene* 2004; 23: 8859-8867.
- Gitlits VM, Toh BH, Sentry JW. Disease association, origin, and clinical relevance of autoantibodies to the glycolytic enzyme enolase. *J Investig Med* 2001; 49: 138-145.
- 88. Pashov A, Kenderov A, Kyukchiev S, Kehayov I, Hristova S, Lacroix-Desmazes S, *et al.* autoantibodies to heat shock protein 90 in the human natural antibody repertoire .*Int Immunol* 2002; 14: 453-461.
- 89. Li WH, Zhao J, Li HY, Liu H, Li AL, Wang HX, *et al.* Proteomics-based identification of autoantibodies in the sera of healthy Chinese individuals from Beijing. *Proteomics* 2006; 6: 4781-4789.
- 90. Panchli V. Multifunctional alpha-enolase: its role in disease. *Cell Mol Life Sci* 2001; 58: 902-920.
- 91. Kim JW, Dang CV. Multifaceted roles of glycolytic enzymes. *Trends Biochem Sci* 2005; 30: 142-150.
- 92. Betterle C, Volpato M, Pedini B, Chen S, Smith BR, Furmaniak J. Adrenal cortex autoantibodies and steroid-producing cells autoantibodies in patients with Addison's disease: comparison of immunoflurescence and immunoprecipitation assays. *J Clin Endocrinol Metab* 1999; 84: 618-622.
- 93. Tanaka H, Perez MS, Powell M, Sanders JF, Sawicka J, Chen S, *et al.* Steroid 21-hydroxylase autoantibodies: measurement with a new immunoprecipitation assay. *J Clin Endocrinol Metab* 1997; 82: 1440-1446.
- 94. Kelkar RL, Meherji PK, Kadam SS, Gupta SK, Nandedkar TD. Circulating auto-antibodies against the zona pellucid and thyroid microsomal antigen in women with premature ovarian failure. *J Reprod Immunol* 2005; 66: 53-67.
- 95. Luborsky J, Llanes B, Davies S, Binor Z, Radwanska E, Pong R. Ovarian autoimmunity: greater frequency of autoantibodies in premature menopause and unexplained infertility than in general population. *Clin Immunol* 1999; 90: 368-374.
- 96. Luborsky J L. Ovarian autoimmune disease and ovarian autoantibodies. *J Women's Health Gend Based Med* 2002; 11: 585-599.
- 97. Pires ES, Parte PP, Meherji PK, Khan SA, Khole VV. Naturally occurring Anti-albumin antibodies are responsible for false positivity in diagnosis of autoimmune premature ovarian failure. *J Histochem Cytochem* 2006; 54: 397-405.
- 98. Bats AS, Barbarino P, Bene M C, Faure GC, Forges T. Local lymphocytic and epithelial activation in a case of autoimmune oophoritis. *Fertil Stertil* 2008; 90: 849.
- 99. Gloor E, Hurlimann J. Autoimmune oophoritis. *Am J Clin Pathol* 1984; 81: 105-109.
- 100. Bannatyne P, Russell P, Shearman RP. Autoimmune oophoritis: a clinical assessment of 12 cases. *In J Gynecol Pathol* 1990; 9: 191-207.

- 101. Fenichel P. Premature ovarian failure: anautoimmune disease? In: Genazzani RA, Petraglia F, Artini GP, editors. Advances in gynecological endocrinology. New York: Parthenon Publishing: 2002; 143-149.
- 102. Massin N, Gougeon A, Meduri G, Thibaud E, Laborde K, Matuchansky C, *et al.* Significance of ovarian histology in the management of patients presenting a premature ovarian failure. *Hum Reprod* 2004; 19: 2555-2560.
- 103. Lass A. Assessment of ovarian reserve- is there a role for ovarian biopsy? *Hum Reprod* 2001; 16: 1055-1057.
- Perheentupa J. APS-1/ APECED: the clinical disease and therapy. *Endocrinol Metab Clin N Am* 2002; 31: 295-320.
- 105. Schatz DA, Winter WE. Autoimmune polyglandular syndrome II: clinical syndrome and treatment. *Endocrinol Metab Clin N Am* 2002; 31: 339-352.
- 106. Poppe K, Glinoer D, Van Steirteghem A, Tournaye H, Devroey P, Schiettecatte J, *et al.* Thyroid dysfunction and autoimmunity in infertile women. *Thyroid* 2002; 12: 997-1001.
- 107. Abalovich M, Mitelberg L, Allami C, Gutierrez S, Alcaraz G, Otero P. Subclinical hypothyroidism and thyroid autoimmunity in women with infertility. *Gynecol Endocrinol* 2007; 23: 279-283.
- Bakalov VK, Vanderhoof VH, Bondy C A, Nelson L M. Adrenal antibodies detect asymptomatic autoimmune adrenal insufficiency in young women with spontaneous premature ovarian failure. *Hum Reprod* 2002; 17: 2096-2100.
- Ambrosi B, Barbetta L, Morricone L. Diagnosis and management of Addison's disease in pregnancy. J Endcrinol Invest 2003; 26: 698-702.
- 110. Maclaren N, Chen QY, Kukrejia A, Marker J, Zhang CH, Sun ZS. Autoimmune hypogonadism. *J Soc Gynecol Invest* 2001; 8: 52-54.
- 111. Kauffmann RP, Castracane VD. Premature ovarian failure associated with autoimmune polyglandular syndrome: pathophysiological mechanisms and future fertility. *J Womens Health (Larchmt)* 2003; 12: 513-520.
- 112. Gleicher N, Weghofer A, Barad HD. A pilot study of premature ovarian senescence: Different genotype and phenotype for genetic and autoimmune etiologies. *Fertil Steril* 2009; 91: 1707-1711.
- 113. Tsigkou A, Marzotti S, Borges L, Brozzetti A, Reis F, Candeloro P, *et al.* High serum inhibin concentration discriminates autoimmune oophoritis from other forms of primary ovarian insufficiency. *J Clin Endocrinol Metab* 2008; 93: 1263-1269.
- 114. Nothnick WB. Treating endometriosis as an autoimmune disease. *Fertil Steril* 2001; 76: 223-231.
- 115. Horejesi J, Martinek J, Novakova D, Madar J, Brandejska M. Autoimmune antiovarian antibodies and their impact on the success of an IVF/ET program. *Ann NY Acad Sci* 2000; 900: 351-356.
- 116. Forges T, Monnier-Barbarino P,Guillet-May F, Faure GC, Bene MC. Corticosteroids in patients with antiovarian antibodies undergoing in vitro fertilization: a prospective pilot study. *Eur J Clin Pharmacol* 2006; 62: 699-705.
- 117. Monnier-Barbarino P, Jouan C, Dubois M, Gober B, Faure GC, Bene MC. Antiovarian antibodies and in vitro fertilization: cause or consequence? *Gynecol Obstet Fertil* 2003; 31: 770-773.

- 118. Meyer WR, Lavy G, DeCherney AH, Vinstin I, Economy K, Lubersky JL. Evidence of gonadal and gonadotropin antibodies in women with a suboptimal ovarian response to exogenous gonadotropin. *Obstet Gynecol* 1990; 75: 795-799.
- 119. Koyama K, Hasegawa A, Inoue M, Isojima S. Blocking of human sperm-zona interaction by monoclonal antibodies to a glycoprotein family (ZP4) of porcine zona pellucid. *Biol Reprod* 1991; 45: 727-735.
- 120. Sivapurapu N, Upadhyay A, Hasegawa A, Koyama K, Gupta SK. Native zona pellucid reactivity and invitro effect on human sperm–egg binding with antisera against bonnet monkey ZP1 and ZP3 synthetic peptides. *J Reprod Immunol* 2002; 56: 77-91.
- 121. Mantzavinos T, Dalamanga N, Hassiakos D, Dimitriadou F, Konidaris S, Zourlas PA. Assessment of autoantibodies to the zona pellucida in serum and follicular fluid in–vitro fertilization patients. *Clin Exp Obstet Gynecol* 1993; 20: 111-115.
- 122. Papale ML, Grillo A, Leonardi E, Giuffrida G, Palumbo M, Palumbo G. Assessment of the relevance of zona pellucida antibodies in follicular fluid of in-vitro fertilization (IVF) patients. *Hum Reprod* 1994; 9: 1827-1831.
- 123. Mardesic T, Ulcova-Gallova Z, Huttelova R, Muller P, Voboril J, Mikova M, *et al.* The influence of different types of antibodies on in vitro fertilization results. *Am J Reprod Immunol* 2000; 43: 1-5.
- 124. Van Kasteren YM, Braat DDM, Hemrika DJ, Lambalk CB, Rekers–Mlombrag LTM, Von Blomberg BME, et al. Corticostroids do not influence ovarian responsiveness to gonadotropins in patients with premature ovarian failure: a randomized, placebocontrolled trial. *Fertil Steril* 1999; 71: 90-95.
- 125. Corenblum B, Rowe T, Taylor PJ. High dose, shortterm glucocorticoids for the treatment of infertility resulting from premature ovarian failure. *Fertil Steril* 1993; 59: 988-991.
- 126. Wang L, Huang P, Huang X, [Analysis of the treatment of 1,020 patients with immunologic infertility] *Zhonghua Fu Chan Ke Za Zhi* 1999; 34: 234-236. (in Chinese).
- 127. Barbarino-Monnier P, Gobert B, Guillet-May F, Bene MC, Barbarino A, Foliguet B, *et al.* Ovarian autoimmunity and corticotherapy in an in-vitrofertilization attempt. *Hum Reprod* 1995; 10: 2006-2007.
- 128. Levesque MC. Translational mini-review series B cell-directed therapies: recent advances in B cell-directed biological therapies for autoimmune disorders. *Clin Exp Immunol* 2009; 157: 198-208.
- Corenblum B, Rowe T, Taylor PJ. High-dose, shortterm glucocorticoids for the treatment of infertility resulting from premature ovarian failure. *Fertil Steril* 1993; 59: 988-991.
- 130. Mamas L, Mamas E. Premature ovarian failure and dehydroepiandrosterone. *Fertil Steril* 2009; 91: 644-646.
- 131. Barad D, Gleicher N. Increased oocyte production after treatment with dehydroepiandrosterone. *Fertil Steril* 2005; 84: 756.
- 132. Kalantaridou SN, Braddock DT, Patronas NJ, Nelson LM. Treatment of autoimmune premature ovarian failure. *Hum Reprod* 1999; 14: 1777-1782.

- 133. Burger HG. Androgen production in women. *Fertil Steril* 2002; 77: S3-5.
- 134. Barad D, Gleicher N. Effect of dehydroepiandrosterone on oocyte and embryo yields, embryo grade and cell number in IVF. *Hum Reprod* 2006; 21: 2845-2849.
- 135. Barad D, Brill H, Gleicher N. Update on the use of dehydroepiandrosterone supplementation among women with diminished ovarian function. *J Assist Reprod Genet* 2007; 24: 629-634.
- 136. Kalu E, Panay N. Spontaneous premature ovarian failure: management challenge. *Gynecol Endocrinol* 2008; 24: 273-279.