

# Oral administration of titanium dioxide nanoparticle through ovarian tissue alterations impairs mice embryonic development

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## Abstract

**Background:** Titanium dioxide nanoparticle (TiO<sub>2</sub>NP) is commonly used in industrial products including food colorant, cosmetics, and drugs. Previous studies have shown that oral administration of TiO<sub>2</sub>NP can be toxic to the reproductive system, but little is known if TiO<sub>2</sub>NP could be able to affect the functions of the female reproductive system, in particular fertility.

**Objective:** The objective was to evaluate the effects of oral administration of TiO<sub>2</sub>NP on histological changes in ovaries, pregnancy rate and in vitro fertility in mice.

**Materials and Methods:** In this experimental study, 54 adult female NMRI mice were randomly assigned to two groups: control group (received vehicle orally) and TiO<sub>2</sub>NP group (received 100 mg/kg/daily TiO<sub>2</sub>NP solution orally). After 5 wk, pregnancy and in vitro fertilization rates, histological changes in ovaries, malondyaldehyde and estrogen hormone levels in the blood serum were investigated and compared between groups.

**Results:** Our results revealed that TiO<sub>2</sub>NP administration induced histological alterations in ovary including, degenerating and reduction of ovarian follicles, ovarian cyst formation and disturbance of follicular development. Compared to control, animals in TiO<sub>2</sub>NP group have shown significant reduction of pregnancy rates and number of giving birth ( $p=0.04$ ). TiO<sub>2</sub>NP caused significant reduction in oocyte number, fertilization rate, and pre-implantation embryo development ( $p<0.001$ ). Furthermore, malondyaldehyde and estrogen hormone levels were significantly ( $p<0.01$ ) increased in mice received TiO<sub>2</sub>NP.

**Conclusion:** Our findings suggest that TiO<sub>2</sub>NP exposure induces alterations on mice ovary resulting in a decrease in the rate of embryo development and fertility.

**Key words:** Nanoparticle, Titanium dioxide, Ovary, In vitro fertilization, Mice.

## Introduction

Nanoparticles (NPs) are materials which at least one of their three dimensions is in the range of 1-100 nm and have unique physiochemical properties and functions which have obtained well interest in many areas in applications such as, food, industry, medical sciences, agriculture and military fields (1). Nanoparticles are being increasingly used and worries on their safety have been increased in scientific society and the general public, especially oral-use food related NPs (2, 3). NPs compared with same large chemical composites have low size and great surface to volume ratio which promote entrance to cells and also increase catalytic and biological activities (4).

Titanium dioxide nanoparticle (TiO<sub>2</sub>NP) or titana is one of the most common materials used in various products and can be found in

three forms, rutile, anatase and brookite (5, 6). Among them, anatase crystal is the most effective form. TiO<sub>2</sub>NP is used in products such as, paints, plastic, papers, cosmetics, clothings, electronics, toothpastes, and especially in sunscreens which due to their ability to block ultraviolet light protect the skin (7, 8). It also extensively used in food industry as a food additive. Thus, oral intaking is a major route from food products (9, 10), as a 75 kg adult human receive 15-37.5 mg/kg/day from food (7).

It has been proven that TiO<sub>2</sub>NP can be toxic to human and animal. Oral administration of this nanoparticle to the mice increases inflammation and disrupts the function of the liver, kidney and reproductive system (11-13) and also increases plasma glucose level (14, 15). Previous studies also have indicated that TiO<sub>2</sub>NP administration leads to aggregation in the vital organs such as, liver, brain, lung, spleen, and kidney (11,

16). Long term  $\text{TiO}_2\text{NP}$  administration (90 days) to mice can be accumulated within ovarian cells, resulting in ovarian dysfunction, mating and pregnancy rate reduction, ovarian inflammation and follicular atresia (13, 17).

It also has been shown that  $\text{TiO}_2\text{NP}$  was transferred to the brain of mice which can affect hormonal release from pituitary gland and female reproduction that leads to imbalance of sex hormone (11, 13). In addition, recent studies have shown that this nanoparticle is able to decrease sperm counts and motility and increases the number of abnormal sperms in epididymis (18, 19). Other studies indicated that  $\text{TiO}_2\text{NP}$  inhibits follicular growth and oocyte maturation of rat (20), and also induces genotoxicity in Chinese hamster ovary cells in vitro (21). Hong and colleague demonstrated that oral administration of 100 mg/kg  $\text{TiO}_2\text{NP}$  to pregnant mice can cross the blood fetal barrier and placental barrier and suppresses embryonic development and also induces fetal skeletal malformation (22).

A considerable body of evidences in laboratory animal models have shown that  $\text{TiO}_2\text{NP}$  administration can cause toxicity in the male reproductive system and induced impairment in testicular morphology and function but, to the best of our knowledge, there is no detailed study on toxic effects of subchronic oral administration of  $\text{TiO}_2\text{NP}$  in the female reproductive system function, particularly in vitro fertilization (IVF). A few previous studies (13, 17) investigated the long term exposure (90 days) of  $\text{TiO}_2\text{NP}$  on female reproductive system and IVF potential has not been evaluated. However, the effects of  $\text{TiO}_2\text{NP}$ -induced in vitro fertility impairment have yet to be studied.

Thus, the current study was conducted to evaluate the adverse effects of oral administration of female mice to  $\text{TiO}_2\text{NP}$  on the histological alterations of ovary, estrogen ( $\text{E}_2$ ) hormone levels, malondialdehyde (MDA) concentration, pregnancy and IVF rates.

## Materials and methods

### Animals and treatments

In this experimental study, 54 adult female mice 10 wk ( $25 \pm 2$  gr) were used. In order to evaluate IVF assay, sperms obtained from a

normal male NMRI mouse was used. The mice were obtained from the Animal Center of Medical School. Animals were kept in stainless steel cages in a ventilated animal house. Room temperature of the animal house was maintained at  $24 \pm 2^\circ\text{C}$  with a 12 hr light/dark cycle. Food and water were given ad libitum. Prior the study, the mice were adapted to this environment for one wk. The mice were randomly categorized into 2 groups ( $n=27/\text{each}$ ): control group received vehicle and test group was supplemented with  $\text{TiO}_2\text{NP}$  solution orally at a dose of 100 mg/kg of body weight for 5 wk (Figure 1).

### Preparation of $\text{TiO}_2\text{NP}$ solution

The size of the  $\text{TiO}_2\text{NP}$  (99% anatase) was 10-25 nm and obtained from US Research Nanomaterials, Inc. It was prepared every wk in phosphate buffered saline ( $\text{pH} = 7.4$ ) with 0.5% Tween 80 and immediately before using was dispersed and sonicated for 10 min.

### Mating of animals

We used 10 mice from each group to evaluate the effect of  $\text{TiO}_2\text{NP}$  on the fertility and pregnancy potential rate. 24 hr after last day of administration, 3 control or test female mice were put in a common cage with 3 males for 11 days and percentage of pregnancy and numbers of newborns were evaluated (23).

### Histological assessments of ovary

All histological assessments were done using standard laboratory methods. After 5 wk, mice were sacrificed and their ovaries were collected. The left ovary from each mouse was fixed with 10% formaldehyde and embedded in paraffin blocks and then sectioned at 5  $\mu\text{m}$  thickness. The stained sections with hematoxylin-eosin (H&E) (Merck, Germany) were evaluated by at least two investigators unaware of the treatments under a light microscope. Seven mice per group used for this part of the study.

### Serum $\text{E}_2$ and MDA levels

At the end of 5 wk, the blood serum of the animals used for histopathological examination were obtained for evaluating the

levels of MDA (Zell Bio, GmbH, Germany) and E<sub>2</sub> hormone (Monobind Inc., Lake Forest, CA, USA) using commercial ELISA kits according to the manufacturer's protocols.

### IVF assays

We used 10 mice from each group for IVF assay. To collect mature oocytes from oviducts, each mouse was intraperitoneally injected with 10 IU pregnant mare's serum gonadotropin (PMSG, Folligon, Netherland) and 10 IU human chorionic gonadotropin (hCG, Folligon, Netherland) after 48 hr. The mice were sacrificed 13-14 hr after human chorionic gonadotropin injection, and then the cumulus-oocyte complexes were obtained from both ampulla of oviducts and transferred to a petri dish containing human tubular fluid (HTF) medium with 4 mg/ml bovine serum albumin (BSA), (Sigma, St. Louis, USA).

The epididymal sperm were obtained from the caudal epididymis of a male adult mouse. Sperm suspensions were placed in HTF-BSA medium and capacitated by incubation at 37°C and 5% CO<sub>2</sub> for at least 1 hr. Then 1×10<sup>6</sup> sperms/ml were added to 500 µL fertilization droplets of HTF-BSA medium containing oocytes. Mineral oil was used to cover droplets. Under inverted microscope, fertilized oocytes were assessed by presence of male and female pronuclei and polar body. After 24 hr of zygotes culture, the percentage of two-cell embryo was assessed and embryonic growth was evaluated after 120 hr (24). In this study the rate of the arrested embryos was categorized into three types:

Type I: embryos with full fragmented blastomeres.

Type II: embryos with partially lysed or fragmented blastomeres.

Type III: embryos with some fragmented blastomeres.

### Ethical consideration

The study was approved and performed according to Ethical Committee Guidance for Research at Laboratory Animals of Urmia University of Medical Sciences (1394-0-32-1791).

### Statistical analysis

The results of IVF were analyzed by 2 proportional test using Minitab software version 15.1 (Minitab Inc., PA, USA). Other results were examined by independent Student's t-test using SPSS software (Statistical Package for the Social Sciences, version 16.0, SPSS Inc, Chicago, Illinois, USA). All results were shown as means±standard deviation (SD) and a p<0.05 was determined as statistically significant.

## Results

### Pregnancy rate and number of newborns

As shown in table I, pregnancy rate and number of newborns were significantly decreased in TiO<sub>2</sub>NP exposed group compared with control group (p=0.04).

### MDA concentration

To determine whether orally administrated TiO<sub>2</sub>NP affect lipid peroxidation in mice, serum MDA levels were measured. As shown in table I, the level of MDA in TiO<sub>2</sub>NP group was significantly (p<0.01) higher than control group.

### E<sub>2</sub> hormone levels

The levels of E<sub>2</sub> hormone in the blood serum of female mice is presented in table I. Orally TiO<sub>2</sub>NP exposure caused significant increase in E<sub>2</sub> levels compared to control (p<0.01).

### Histological observations

Histological examination of the ovaries in the control group showed normal architecture of ovarian tissue with normal follicles at different stages of development. Several pathological changes were observed in the TiO<sub>2</sub>NP group, including degeneration and reduction of ovarian follicles, ovarian cyst formation, and follicular development impairment suggesting that ovaries have been damaged by TiO<sub>2</sub>NP exposure.

### Fertilization rate and embryonic development

Data of oocytes number, IVF rate, and pre-implantation embryonic development in

studying groups are presented in table II. The results indicated that number of oocytes, percentages of fertilization, two-cell embryos (as an indicator of cleavage initiation), and embryos in blastocyst stage in the TiO<sub>2</sub>NP

group were significantly lower than control group (p<0.001). Compared with the control group, in TiO<sub>2</sub>NP group the total percentages of arrested embryos were significantly higher (p<0.001).

**Table I.** The effects of TiO<sub>2</sub>NP on pregnancy rate, number of newborns, E2 and MDA levels of mice after oral administration with 100 mg/kg body weight for 5 wk

Index	Control	TiO <sub>2</sub> NP (100mg/kg)
Pregnancy rate (%)	100	70
Number of newborns	8.9 ± 1.19	7.2 ± 0.92 <sup>a</sup>
MDA (nmol/ml)	1.43 ± 0.07	1.8 ± 0.24 <sup>b</sup>
E <sub>2</sub> (pmol/l)	61.56 ± 5.71	73.74 ± 7.56 <sup>b</sup>

Data of pregnancy rate presented as% and the rest presented as means ± SD.  
independent-samples t-test (n=10)

<sup>b</sup> Significantly different from the control (p<0.01)  
MDA: Malondyaldehyde

<sup>a</sup> Significantly different from the control (p<0.05)  
TiO<sub>2</sub>NP: Titanium dioxide nanoparticle  
E<sub>2</sub>: Estrogen

**Table II.** The number of oocytes and percentages of fertilized oocytes and embryos (two- cell and blastocyst) of mice after oral exposure with 100 mg/kg TiO<sub>2</sub>NP for 5 wk

Groups	Oocytes	Fertilized oocytes	Two cells	Blastocysts	Arrest	Arrest type I	Arrest type II	Arrest type III
Control	271 <sup>a</sup>	258 (95.2) <sup>b</sup>	234 (90.7) <sup>a</sup>	155 (60.08) <sup>a</sup>	103 (39.9) <sup>a</sup>	3 (0.78) <sup>a</sup>	7 (2.71) <sup>a</sup>	94 (36.43) <sup>b</sup>
TiO <sub>2</sub> NPS	194	173 (89.17)	102 (52.95)	37 (21.39)	136 (78.6)	34 (19.65)	26 (15.03)	76 (43.93)

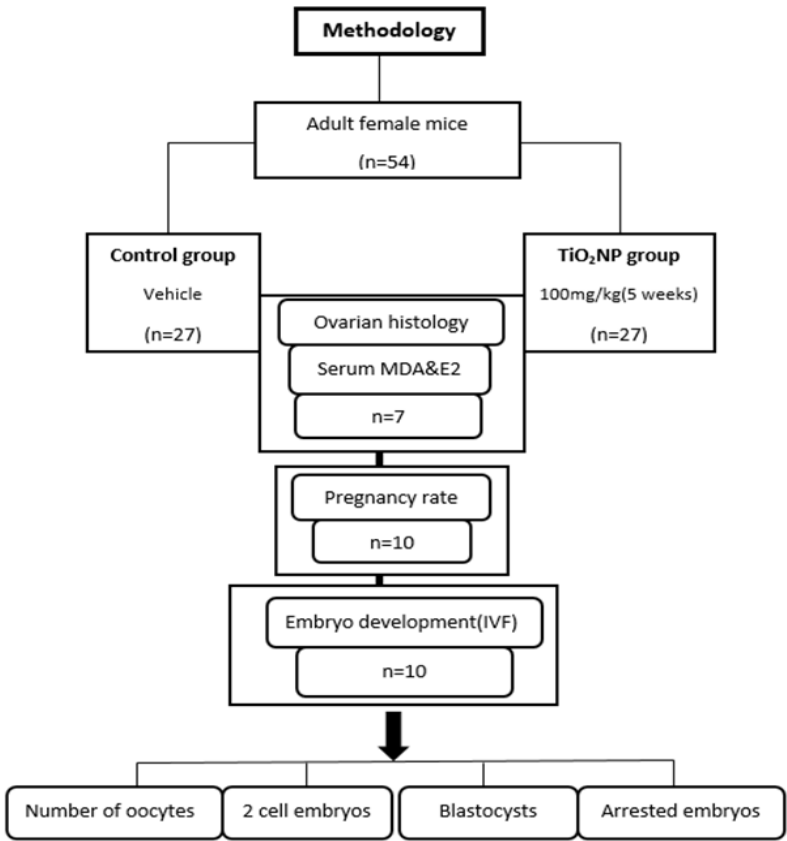
Data of oocytes presented as n and the rest presented as n (%)

2 proportional test (n=10)

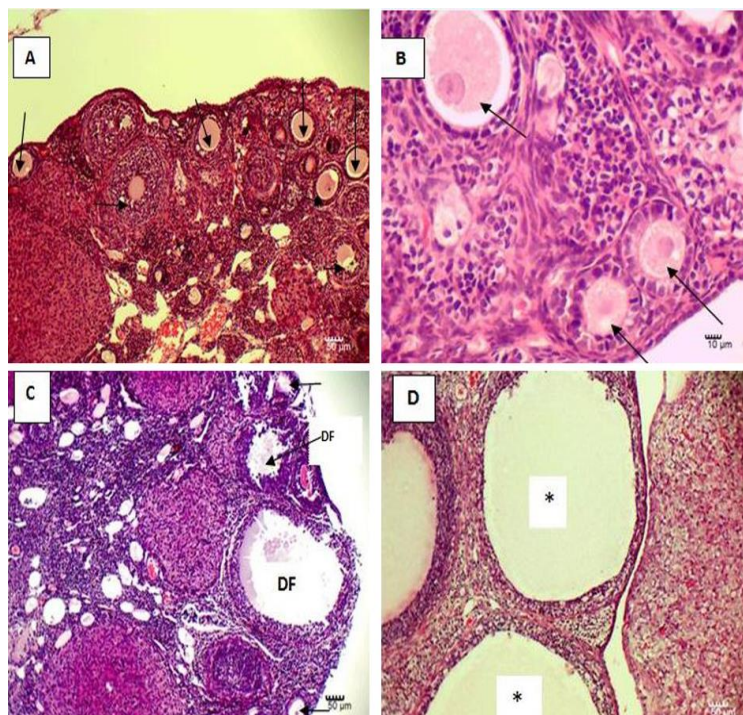
<sup>b</sup> Significantly different from the control (p<0.05)

<sup>a</sup> Significantly different from the control (p<0.001)

TiO<sub>2</sub>NP: Titanium dioxide nanoparticle



**Figure 1.** Flowchart for the experiment design.



**Figure 2.** Effects of oral administration of  $\text{TiO}_2\text{NP}$  on mice ovary: A and B, control group showing normal structure with normal follicles; C and D, ovarian tissue in  $\text{TiO}_2\text{NP}$  exposed group showing reduction and degenerating of follicles and cyst formation. Arrows indicate follicle, DF; degenerating follicle, \*; cyst in ovary. (A and D  $\times 100$ ; B and C  $\times 400$  magnification).

## Discussion

The results of the present study showed that oral administration of  $\text{TiO}_2\text{NP}$  with 100 mg/kg for 5 wk caused reproductive system toxicity. The findings indicated that  $\text{TiO}_2\text{NP}$  administration led to folliculogenesis impairment and pregnancy and in vitro fertilization rates reduction.

It has been proven that orally administration with  $\text{TiO}_2\text{NP}$  accumulated in mouse ovary and lead to ovarian dysfunction such as atresia of primary and secondary follicles and also ovarian apoptosis induction (17). In this study, adult female mice were treated for 5 wk. Considering the time of the mouse estrous cycle is 4-5 days, in this study we exposed the mice for about 5 cycles to investigate the toxic effects of  $\text{TiO}_2\text{NP}$ .

Our findings indicated that subchronic  $\text{TiO}_2\text{NP}$  exposure resulted in an increase in the serum  $\text{E}_2$  concentration that was agreed with previous studies which also showed significant reduction of levels of luteinizing hormone (LH), testosterone, progesterone, and follicle-stimulating hormone (FSH) (13, 17). Decreased pregnancy rate in the present study may be related to imbalance of sex hormone concentrations. It is well known that there is a direct relationship between normal

fertility and ovarian follicular development with the levels of sex hormones (17). In females, progesterone plays a key role in ovulation, implantation, and pregnancy (25) and also has been proven to be important for ovulation through increasing of proteolytic enzymes production (26). Furthermore, LH and FSH are the most important hormones of hypothalamic-pituitary-gonadal axis, which regulates the production of gametes and fertility.  $\text{E}_2$  hormone also facilitates induction of receptor systems for FSH and LH in the granulosa cells (27). Thus, reduction of folliculogenesis and fertility by  $\text{TiO}_2\text{NP}$  administration may be due to impairments of sex hormones release such as  $\text{E}_2$ , FSH, LH, and progesterone (17). Increased  $\text{E}_2$  levels in the current study and in previous studies may be associated with the activation of cytochrome p450 aromatase which is responsible for converting testosterone to  $\text{E}_2$ , but proving this needs further investigation (28).

Our findings also indicated that the in vitro fertility potential and embryonic development in mice administrated with  $\text{TiO}_2\text{NP}$  were lower than those of the control mice and also the percentage of arrested embryos was significantly higher after  $\text{TiO}_2\text{NP}$  administration. It is clear that embryonic



development in both in vitro and in vivo is associated to quality of both gametes (oocyte and sperm) (29). The previous studies indicated that chronic oral exposure of TiO<sub>2</sub>NP for 90 consecutive days led to significant accumulation in the mouse ovaries and altered the ovarian stromal ultrastructure such as, mitochondrial alterations including swelling and crista break, nucleus chromatin condensing, and nuclear membrane deformity. These alterations suggest that TiO<sub>2</sub>NP administration caused ovarian stromal cells apoptosis (17). Extracellular matrix of ovary and also the stromal cells play an important role in tissue integrity, normal function of the ovary, and follicular development. Therefore, the reduction in mice fertility potential following TiO<sub>2</sub>NP administration may be due to disturbances in the organization of the ovarian cells ultrastructure and also hormonal imbalance.

In the current study, we observed histological alterations in ovarian tissue caused by TiO<sub>2</sub>NP administration including impairment in folliculogenesis and degenerating of follicles resulted in decrease of mature oocytes. These findings were confirmed by IVF assay results which indicated that the number of oocytes collected from these mice were significantly lower than those of the unexposed mice. Moreover, the quality of these retrieved oocytes were lower in treatment group. Thus, decreased the number of two cell embryos and mating capacity of female mice following TiO<sub>2</sub>NP administration were closely associated with these findings. To our knowledge, no published data was available about the toxic effects of TiO<sub>2</sub>NP on in vitro development of female mice embryos. Thus, we are not able to compare the findings obtained from the present study with other studies.

In this study, the TiO<sub>2</sub>NP exposed mice showed a statistically significant increase in MDA levels. Previous studies showed that TiO<sub>2</sub>NP increase reactive oxidative species (ROS) levels in mice (30, 31). MDA produces following reaction between ROS and lipids, and its level is increased after ROS elevation (32). Thus, MDA is commonly used as ROS-related marker (30). In the present study, the analysis of serum MDA levels after 5 wk showed that its level in the TiO<sub>2</sub>NP supplemented group was higher than control

group which was consistent with previous report (30).

The toxic effects of TiO<sub>2</sub>NP exposure in vivo model may be caused through both direct and indirect pathways (33). In direct pathway, TiO<sub>2</sub>NP is able to enter the blood circulation after oral exposure and then be distributed to different tissues such as liver, spleen, kidney, lung (11) and also ovary (13). Thus, in current study, it seems that the accumulation of this nanoparticle in ovaries caused toxicity which was demonstrated by biochemical and histological alterations.

In indirect pathway, oral administration of TiO<sub>2</sub>NP affects ovary through increasing ROS levels and also through triggering inflammatory responses. According to previous studies, increased levels of inflammatory cytokines including tumor necrosis factor alpha, interleukin-6, and interleukin-8 in the blood serum of mice and rats after oral administration of TiO<sub>2</sub>NP suggests significant inflammatory responses (30, 34). As mentioned earlier, we found a significant elevation in the level of MDA which was consistent with previous studies. Thus, stress oxidative induced by TiO<sub>2</sub>NP exposure probably is responsible for female reproductive system damage.

Our study is not without limitation. We did not measure levels of other sex hormones (LH and FSH) and, we used only one dose of TiO<sub>2</sub>NP for 5 wk. Therefore, we propose to apply lower doses of exposure in future studies.

## Conclusion

The results of the current study indicate that subchronic oral administration of TiO<sub>2</sub>NP may affect pregnancy rate, in vitro fertility and embryo development and also induces ovarian tissue damage and ultimately leads to mice ovarian dysfunction which may be associated with lipid peroxidation and imbalance of sex hormones. Therefore, these data can provide useful information on the risk of application of TiO<sub>2</sub>NP in young females, especially during childbearing years.

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## Conflict of interest

The authors declare that there are no conflicts of interest.

## References

- Win-Shwe TT, Fujimaki H. Nanoparticles and neurotoxicity. *Int J Mol Sci* 2011; 12: 6267-6280.
- Vandebriel RJ, Vermeulen JP, van Engelen LB, de Jong B, Verhagen LM, de la Fonteyne-Blankestijn LJ, et al. The crystal structure of titanium dioxide nanoparticles influences immune activity in vitro and in vivo. *Part Fibre Toxicol* 2018; 15: 1-12.
- Bouwmeester H, Dekkers S, Noordam MY, Hagens WI, Bulder AS, de Heer C, et al. Review of health safety aspects of nanotechnologies in food production. *Regul Toxicol Pharmacol* 2009; 53: 52-62.
- Oberdörster G. Safety assessment for nanotechnology and nanomedicine: concepts of nanotoxicology. *J Int Med* 2010; 267: 89-105.
- Brun E, Barreau F, Veronesi G, Fayard B, Sorieul S, Chanéac C, et al. Titanium dioxide nanoparticle impact and translocation through ex vivo, in vivo and in vitro gut epithelia. *Part Fib Toxicol* 2014; 11: 1-16.
- Zhang L, Xie X, Zhou Y, Yu D, Deng Y, Ouyang J, et al. Gestational exposure to titanium dioxide nanoparticles impairs the placentation through dysregulation of vascularization, proliferation and apoptosis in mice. *Int J Nanomedicine* 2018; 13: 777-789.
- Weir A, Westerhoff P, Fabricius L, Hristovski K, von Goetz N. Titanium dioxide nanoparticles in food and personal care products. *Environ Sci Technol* 2012; 46: 2242-2250.
- Kaida T, Kobayashi K, Adachi M, Suzuki F. Optical characteristics of titanium oxide interference film and the film laminated with oxides and their applications for cosmetics. *J Cosmet Sci* 2004; 55: 219-220.
- Dorier M, Brun E, Veronesi G, Barreau F, Pernet-Gallay K, Desvergne C, et al. Impact of anatase and rutile titanium dioxide nanoparticles on uptake carriers and efflux pumps in Caco-2 gut epithelial cells. *Nanoscale* 2015; 7: 7352-7360.
- Tassinari R, Cubadda F, Moracci G, Aureli F, D'Amato M, Valeri M, et al. Oral, short-term exposure to titanium dioxide nanoparticles in Sprague-Dawley rat: focus on reproductive and endocrine systems and spleen. *Nanotoxicology* 2014; 8: 654-662.
- Jia X, Wang S, Zhou L, Sun L. The potential liver, brain, and embryo toxicity of titanium dioxide nanoparticles on mice. *Nanoscale Res Lett* 2017; 12: 478.
- Jafari A, Rasmi Y, Hajaghadzadeh M, Karimipour M. Hepatoprotective effect of thymol against subchronic toxicity of titanium dioxide nanoparticles: Biochemical and histological evidences. *Environ Toxicol Pharmacol* 2018; 58: 29-36.
- Zhao X, Ze Y, Gao G, Sang X, Li B, Gui S, et al. Nanosized TiO<sub>2</sub>-induced reproductive system dysfunction and its mechanism in female mice. *PLoS One* 2013; 8: e59378.
- Hu H, Guo Q, Wang C, Ma X, He H, Oh Y, et al. Titanium dioxide nanoparticles increase plasma glucose via reactive oxygen species-induced insulin resistance in mice. *J Appl Toxicol* 2015; 35: 1122-1132.
- Hu H, Li L, Guo Q, Zong H, Yan Y, Yin Y, et al. RNA sequencing analysis shows that titanium dioxide nanoparticles induce endoplasmic reticulum stress, which has a central role in mediating plasma glucose in mice. *Nanotoxicology* 2018; 12: 341-356.
- Gui S, Zhang Z, Zheng L, Cui Y, Liu X, Li N, et al. Molecular mechanism of kidney injury of mice caused by exposure to titanium dioxide nanoparticles. *J Hazard Mater* 2011; 195: 365-370.
- Gao G, Ze Y, Li B, Zhao X, Zhang T, Sheng L, et al. Ovarian dysfunction and gene-expressed characteristics of female mice caused by long-term exposure to titanium dioxide nanoparticles. *J Hazard Mater* 2012; 243:19-27.
- Khorsandi L, Orazizadeh M, Moradi-Gharibvand N, Hemadi M, Mansouri E. Beneficial effects of quercetin on titanium dioxide nanoparticles induced spermatogenesis defects in mice. *Environ Sci Pollut Res Int* 2017; 24: 5595-5606.
- Hong F, Zhao X, Si W, Ze Y, Wang L, Zhou Y, et al. Decreased spermatogenesis led to alterations of testis-specific gene expression in male mice following nano-TiO<sub>2</sub> exposure. *J Hazard Mater* 2015; 300: 718-728.
- Juan H, XuYing W, Fei W, GuiFeng X, Zhen L, TianBao Z. Effects of titanium dioxide nanoparticles on development and maturation of rat preantral follicle in vitro. *Acad J Sec Mil Med Univ* 2009; 30: 869-873.
- Di Virgilio AL, Reigosa M, Arnal PM, Fernández Lorenzo de Mele M. Comparative study of the cytotoxic and genotoxic effects of titanium oxide and aluminium oxide nanoparticles in Chinese hamster ovary (CHO-K1) cells. *J Hazard Mater* 2010; 177: 711-718.
- Hong F, Zhou Y, Zhao X, Sheng L, Wang L. Maternal exposure to nanosized titanium dioxide suppresses embryonic development in mice. *Int J Nanomedicine* 2017; 12: 6197-6204.
- Thum C, McNabb WC, Young W, Cookson AL, Roy NC. Prenatal caprine milk oligosaccharide consumption affects the development of mice offspring. *Mol Nutr Food Res* 2016; 60: 2076-2085.
- Ahmadi A, Bamohabat Chafjiri S, Sadrkhanlou RA. Effect of satreja khuzestanica essential oil against fertility disorders induced by busulfan in female mice. *Vet Res Forum* 2017; 8: 281-286.
- Graham JD, Clarke CL. Physiological action of progesterone in target tissues. *Endocr Rev* 1997; 18: 502-519.
- Iwamasa J, Shibata S, Tanaka N, Matsuura K, Okamura H. The relationship between ovarian progesterone and proteolytic enzyme activity during ovulation in the gonadotropin-treated immature rat. *Biol Reprod* 1992; 46: 309-313.
- Lyon MF, Glenister PH. Reduced reproductive performance in androgen-resistant Tfm/Tfm female mice. *Proc R Soc Lond B Biol Sci* 1980; 208:1-12.
- Auger AP, Tetel MJ, McCarthy MM. Steroid receptor coactivator-1 (SRC-1) mediates the development of sex-specific brain morphology and behavior. *Proc Natl Acad Sci U.S.A.* 2000; 97: 7551-7555.

29. Shoukir Y, Chardonens D, Campana A, Sakkas D. Blastocyst development from supernumerary embryos after intracytoplasmic sperm injection: a paternal influence? *Hum Reprod* 1998; 13: 1632-1637.
30. Gu N, Hu H, Guo Q, Jin S, Wang C, Oh Y, et al. Effects of oral administration of titanium dioxide fine-sized particles on plasma glucose in mice. *Food Chem Toxicol* 2015; 86: 124-131.
31. Hu H, Guo Q, Wang C, Ma X, He H, Oh Y, et al. Titanium dioxide nanoparticles increase plasma glucose via reactive oxygen species-induced insulin resistance in mice. *J Appl Toxicol* 2015; 35: 1122-1132.
32. Niska K, Pyszka K, Tukaj C, Wozniak M, Radomski MW, Inkielewicz-Stepniak I. Titanium dioxide nanoparticles enhance production of superoxide anion and alter the antioxidant system in human osteoblast cells. *Int J Nanomedicine* 2015; 10: 1095-1107.
33. Nelin TD, Joseph AM, Gorr MW, Wold LE. Direct and indirect effects of particulate matter on the cardiovascular system. *Toxicol Lett* 2012; 208: 293-299.
34. Chen Z, Wang Y, Zhuo L, Chen S, Zhao L, Luan X, et al. Effect of titanium dioxide nanoparticles on the cardiovascular system after oral administration. *Toxicol Lett* 2015; 239: 123-130.