

# Relationship between *MspI* polymorphism of *CYP1A1* gene and the risk of endometriosis in an Iranian population: A case-control study

Azam Babaki<sup>1</sup> M.Sc., Ehsan Zare Mehrjardi<sup>1,2</sup> M.Sc., Razieh Dehghani Firouzabadi<sup>3</sup> M.D., Abbas Aflatoonian<sup>3</sup> M.D., Seyed Morteza Seifati<sup>1</sup> Ph.D.

1. Medical Biotechnology Research Center, Ashkezar Branch, Islamic Azad University, Ashkezar, Yazd, Iran.
2. Young Researchers and Elite Club, Ashkezar Branch, Islamic Azad University, Ashkezar, Yazd, Iran.
3. Research and Clinical Center for Infertility, Yazd Reproductive Sciences Institute, Shahid Sadoughi University of Medical Sciences, Yazd, Iran.

## Corresponding Author:

Seyed Morteza Seifati, Medical Biotechnology Research Center, Ashkezar Branch, Islamic Azad University, Daneshgah Blvd. Ashkezar, Yazd, Iran. Postal Code: 8941698480  
**Email:** seifati@gmail.com  
**Tel:** (+98) 9131546072

Received: 7 February 2018

Revised: 25 June 2018

Accepted: 21 July 2018

## Abstract

**Background:** Endometriosis is a disease that affects women of reproductive age. This disease is characterized by the presence of endometrial-like tissues (endometrial or stromal glands) outside the uterus and shows significantly elevated prevalence in industrial regions. Additionally, an interaction between genetics and environmental factors is assumed for the disease. Enzymes belonging to the cytochrome P450 (CYP) family are participated in detoxification process of a wide range of environmental toxins and carcinogens. Thereby, they are good link for the interaction. *CYP1A1* which belong to cytochrome P450 (CYPs) superfamily, is a very important gene for the metabolism of carcinogens.

**Objective:** The aim of this study was to analyze the frequency of the *MspI* polymorphism of *CYP1A1* gene and its relation to endometriosis.

**Materials and Methods:** Genomic DNA was isolated from 93 endometriosis women and 139 healthy controls. Genotyping was performed using polymerase chain reaction followed by restriction fragment length polymorphism analysis.

**Results:** Frequencies of the TT, TC, and CC genotype of *CYP1A1* gene polymorphism in patients were 73.1%, 22.6%, and 4.3%, while frequencies in controls were 74.1%, 22.3%, and 3.6%, respectively. So there was no significant differences between the genotypes in two groups ( $p=0.961$ ).

**Conclusion:** According to our study, *MspI* polymorphism of *CYP1A1* gene appears to be not associated with the risk of endometriosis in the studied population. However, additional studies, especially with larger sample size are needed to validate these findings.

**Key words:** Endometriosis, Cytochrome P-450 *CYP1A1*, *MspI* endonuclease.

*This article extracted from M.Sc thesis. (Azam Babaki)*

## Introduction

Endometriosis is defined as the presence of endometrial glands and stroma outside their normal locations (1, 2). This disease is one of the main causes of infertility and chronic disorders among women, affecting approximately 7-10% of women in their fertility age and 5-50% of infertile women (3-6). It has a wide range of symptoms of which pelvic pain, dysmenorrhea, dyspareunia, and abnormal uterus bleeding are the most important ones (7). Endometriosis is a complex trait. Some of studies have suggested the role of immunological, endocrine, environmental and genetics factors in the pathogenesis of this disease. Moreover, it has been revealed that endometriosis is more common in first degree relatives of endometriosis patients compared to first degree relatives of unaffected

individuals; reflecting the involvement of genetics factors (3-5). Increasing evidences have shown that multiple gene loci interact with each other and with the environment to produce endometriosis (6, 7). So far, the relationship between endometriosis and different genes has been investigated. Among them, genes involved in detoxification systems such as Cytochrome (CYP) P450 superfamily are the most important ones and possibly a good link for interactions of genetic and environmental factors (8, 9). Enzymes of this family are participated in detoxification process of a wide range of environmental pollutants and carcinogens (10).

*CYP1A1*, a member of P450 cytochrome superfamily, is located on chromosome 15 (15q 24.1) (11). *CYP1A1* rs4646903 (T>C) polymorphism, downstream from the polyadenylation site at the 3' end of exon 7, is one of the most important polymorphisms of

the gene. Substitution of T to C at this position formed a target site for *MspI* enzyme, so this polymorphism is also named *MspI* or M1 (11, 12). It has been reported that this polymorphism is correlated with CYP1A1 enzyme inducibility phenotype. There are some studies reported the relationship between *MspI* polymorphism and the risk of endometriosis (13, 14).

The goal of this study was investigation of the association between *MspI* polymorphism of CYP1A1 gene and endometriosis in women referred to two infertility centers in Yazd, Iran who were from central and southern parts of Iran.

## Materials and methods

In this case-control study, 93 women with endometriosis (age range 20-42 yr) referred to two infertility centers in Yazd, Iran: Madar Hospital and Yazd Research and Clinical Center for Infertility were enrolled in this study as the case group. Endometriosis was diagnosed by an experienced gynecologist through laparoscopy. Guidelines of American Society for Reproductive Medicine (ASRM) was used to classify patients (ASRM) into 4 stages; minimal (stage I), mild (stage II), moderate (stage III), and severe (stage IV) (15).

The control group consisted of 139 unrelated healthy premenopausal women (age range: 18-50 yr) undergoing Cesarean section or hysterectomy at the same centers with no history of endometriosis and without any lesion suggesting endometriosis during Cesarean section or hysterectomy. Women with a history of autoimmune disease and cancer were excluded from the research. Clinical characteristics of all the participants were collected personally using a designed questionnaire.

## Genotype determination

Genomic DNA was extracted from 5 mL of EDTA anticoagulated whole blood using AccuPrep® Genomic DNA Extraction Kit (Bioneer, South Korea) based on manufacturer's manual. To study *MspI* polymorphism of CYP1A1 gene, PCR-based restriction fragment length polymorphism (RFLP) method was used. The specific primers for the target region (forward primer of 5'-TAGGAGTCTTGTCTCATGCC-3' and reverse primer of 5'-CAGTGAAGAGGTGTAGCCGC-3') was designed by Gene Runner (version 5.1) software and synthesized by Macrogen (South Korea). PCR reaction mixture was contained 1X PCR buffer, 2 mM

MgCl<sub>2</sub>, 2.5 unit of Taq DNA Polymerase (Cinnagen, Iran), 0.5 mM dNTP, 5 pM of each primer, and 30 ng of pattern DNA up to the volume of 25 µl of sterile distilled water. Amplification was performed as follows: initial denaturation at 95°C for 5 min, then 30 cycles of denaturation at 94°C for 30 s, annealing at 60°C for 45 s, extension at 72°C for 60 s and final extension at 72°C for 10 min.

The amplification products (343 bp) were digested with *MspI* (Fermentas, Germany) for 16 hr at 37°C. Then *MspI* restriction digestion products were loaded on 2% agarose gel. The genotypes were characterized as TT when the PCR products remained uncut (343 bp), as CC when the restriction site was present, and as TC when the digestion produced a pattern of three DNA fragments sized 343 bp, 136 bp, and 207 bp.

## Ethical consideration

The Islamic Azad University, Yazd Branch Ethics Committee approved this study (IR.IAU.YAZD.REC.1396,23). Written informed consent was obtained from all participants.

## Statistical analysis

The Pearson  $\chi^2$  or Fisher's exact test was used to compare genotype distributions between patients and controls. The statistical modeling using logistic regression was used to calculate the relative risk (odds ratio; OR) of genotypes for case-control study. Odds ratios were expressed together with the 95% confidence interval (16).  $p \leq 0.05$  was considered statistically significant. The Hardy-Weinberg equilibrium analysis was conducted by comparing observed versus expected genotype frequencies using a Chi-square test.

## Results

The mean age of the case group was  $28.91 \pm 5.18$  yr and  $29.85 \pm 8.77$  yr in control group ( $p=0.356$ ). The mean body mass index (BMI) was  $24.30 \pm 3.69$  in the case group and  $26.14 \pm 4.82$  in controls. Statistical analysis showed that BMI of the cases is significantly less than control group ( $p=0.003$ ) (Table I). Stage II of the disease has the highest frequency (40.20%). Totally, 51 patients (55.40%) had primary stages of disease (I, II) and 41 of them (44.60%) were in the advanced stages of endometriosis (III, IV). Regarding both controls and patients; the distribution of *MspI* genotypes was in Hardy-Weinberg equilibrium.

The most frequent genotype in two groups was TT. The frequencies of this genotype in

patients and control group were 73.1% and 74.1%, respectively. Data analysis by  $\chi^2$  test revealed that there is no significant difference between the two groups ( $p=0.961$ ). Using the TT homozygote as a reference group, the OR of TC heterozygotes (OR, 1.016; 95% CI, 0.54 to 1.93) and CC homozygotes was (OR, 1.204; 95% CI, 0.31-4.6). Allele frequencies of *MspI* polymorphism in the case and control groups are shown in table II. Comparison between the alleles by  $\chi^2$  test indicates no significant difference between two groups ( $p=0.804$ ). In all stages of endometriosis, CC genotype has the lowest frequency of 7.2%, 5.4%, 0.0%, and 4.5% for stages I to IV, respectively (Table III). Statistical analysis of the frequencies of *MspI* genotypes by Fisher's

exact test indicated that there is no significant difference between the genotypes and different stages of endometriosis ( $p=0.491$ ). In addition, the patients with advanced stages were compared with the primary stages and control group; there was no significant difference between two groups ( $p=0.427$  and  $p=0.507$ , respectively).

**Table I.** General characteristics of the case and control groups

| Characteristic            | Case             | Control          | p-value            |
|---------------------------|------------------|------------------|--------------------|
| Age (yr)*                 | 28.91 $\pm$ 5.18 | 29.85 $\pm$ 8.77 | 0.356 <sup>a</sup> |
| BMI (kg/m <sup>2</sup> )* | 24.09 $\pm$ 3.70 | 26.14 $\pm$ 4.82 | 0.003 <sup>a</sup> |
| Primary infertility**     | 64 (76.2)        | 7 (63.6)         | 0.373 <sup>b</sup> |
| Secondary infertility**   | 20 (23.8)        | 4 (36.4)         |                    |

\* Data presented as mean $\pm$ SD. <sup>a</sup> Students' *t* test

\*\*Data presented as n (%). <sup>b</sup> Fisher's exact test

$p>0.05$  was considered statistically significant.

**Table II.** Genotype and allele frequencies of *MspI* polymorphism in the case and control groups

| Genotypes and allele | Case group | Control group | Odds ratio             | p-value            |
|----------------------|------------|---------------|------------------------|--------------------|
| CC                   | 4 (4.3)    | 5 (3.6)       | 1.204 (0.31-4.6)       | 0.961 <sup>a</sup> |
| TC                   | 21 (22.6)  | 31 (22.3)     | 1.016 (0.54-1.93)      |                    |
| TT                   | 68 (73.1)  | 103 (74.1)    | Ref.                   | 0.804 <sup>b</sup> |
| T                    | 157 (84.4) | 237 (85.3)    | Ref.                   |                    |
| C                    | 29 (15.6)  | 41 (14.7)     | 0.9366 (0.5587-1.5699) |                    |

Data presented as n (%).  $p>0.05$  was considered statistically significant. <sup>a</sup> Fisher's exact test. <sup>b</sup>  $\chi^2$  test

**Table III.** Genotype frequencies of *MspI* polymorphism in different stages of endometriosis

| Severity | Minimal   | Mild      | Moderate  | Severe    | Total     |
|----------|-----------|-----------|-----------|-----------|-----------|
| Genotype |           |           |           |           |           |
| TT       | 10 (71.1) | 23 (62.2) | 17 (89.5) | 17 (77.3) | 67 (72.8) |
| TC       | 3 (21.4)  | 12 (32.4) | 2 (10.5)  | 4 (18.2)  | 21 (22.8) |
| CC       | 1 (7.2)   | 2 (5.4)   | 0 (0)     | 1 (4.5)   | 4 (4.4)   |
| Total    | 14 (100)  | 37 (100)  | 19 (100)  | 22 (100)  | 92 (100)  |

Data presented as n (%).  $p>0.05$  was considered statistically significant. Fisher's exact test ( $p=0.491$ )

## Discussion

The role of products of *CYP1A1* gene in metabolism of estrogen and PAH has been well-defined (9, 17). It was observed that *CYP1A1* was implicated in metabolism of dioxin, which may increase the severity of endometriosis in rhesus monkeys (18). Therefore, considering that endometriosis is an estrogen-dependent disease (19), *CYP1A1* may be associated with this disease. The level of gene expression or mRNA stability can be altered by the *CYP1A1 MspI* polymorphism which resulting in a highly inducible activity of the enzyme (20).

In the present study, results showed that there is no statistical significant difference between the frequencies of the genotypes in two groups ( $p=0.961$ ). Previous reports on association of *MspI* polymorphism of *CYP1A1* gene with endometriosis are controversial. In consistency with our findings, there are some studies in different populations such as in UK ( $p=0.11$ ) (15), China ( $p>0.05$ ) (21), India ( $p>0.05$ ) (22, 23), Austria ( $p=0.6$ ) (24), Taiwan ( $p>0.05$ ) (1), USA ( $p=0.65$ ) (14) and Brazil ( $p=0.087$ ) (12), which did not found any

correlation between the polymorphism and endometriosis. In contrast, Arvanitis and colleagues examined Greek population and noticed that TT genotype will result in 38% decrease in endometriosis risk (OR, 0.62; 95% CI, 0.440 to 0.883) (13). In our study, there was no significant difference between the allele frequencies of two groups ( $p=0.804$ ). The same results were reported by some other studies (1, 14, 15, 21-24). In contrast, Barbosa and colleagues suggested that the frequencies of C allele was higher in Brazilian women with endometriosis as compared to the control group ( $p=0.006$ ). Diversity in genetics and environmental background, sample size and selection of control group might be reason for differences among results. In our research, no significant difference was observed between age and cause of infertility of patient and control groups. However, mean BMI of the patients was less than the control group and this difference was statistically significant ( $p=0.003$ ). These findings confirmed the results of other studies which were reported decrease of BMI is associated with endometriosis (1, 14, 25, 26).

## Conclusion

In conclusion, this study showed that CYP1A1 MspI polymorphism was not associated with endometriosis risk in our population. More studies with larger sample size and especially, in other populations with different ethnicity can provide a better understanding of the relationship between population-specific environmental variables and genetic backgrounds.

## Acknowledgments

This article was financially and technically supported by the Islamic Azad University, Ashkezar Branch. The authors are grateful to the Madar Hospital and Yazd Research and Clinical Center for Infertility for helping us.

## Conflict of interest

The authors report no conflicts of interest.

## References

- Juo SH, Wang TN, Lee JN, Wu MT, Long CY, Tsai EM. CYP17, CYP1A1 and COMT polymorphisms and the risk of adenomyosis and endometriosis in Taiwanese women. *Hum Reprod* 2006; 21: 1498-1502.
- Missmer SA, Cramer DW. The epidemiology of endometriosis. *Obstet Gynecol Clin North Am* 2003; 30: 1-19.
- Seifati SM, Parivar K, Aflatoonian A, Dehghani Firouzabadi R, Sheikhha MH. No association of GSTM1 null polymorphism with endometriosis in women from central and southern Iran. *Iran J Reprod Med* 2012; 10: 23-28.
- Simpson JL, Bischoff FZ. Heritability and molecular genetic studies of endometriosis. *Ann N Y Acad Sci* 2002; 955: 239-251.
- Stefansson H, Geirsson RT, Steinthorsdottir V, Jonsson H, Manolescu A, Kong A, et al. Genetic factors contribute to the risk of developing endometriosis. *Hum Reprod* 2002; 17: 555-559.
- Hur SE, Lee JY, Moon HS, Chung HW. Polymorphisms of the genes encoding the GSTM1, GSTT1, and GSTP1 in Korean women: no association with endometriosis. *Mol Hum Reprod* 2005; 11: 15-19.
- Dun EC, Taylor RN, Wieser F. Advances in the genetics of endometriosis. *Genome Med* 2010; 2: 75.
- Cascorbi I. Genetic basis of toxic reactions to drugs and chemicals. *Toxicol Lett* 2006; 162: 16-28.
- Fan W, Huang Z, Xiao Z, Li S, Ma Q. The cytochrome P4501A1 gene polymorphisms and endometriosis: a meta-analysis. *J Assist Reprod Genet* 2016; 33: 1373-1383.
- Parveen F, Faridi RM, Das V, Tripathi G, Agrawal S. Genetic association of phase I and phase II detoxification genes with recurrent miscarriages among North Indian women. *Mol Hum Reprod* 2010; 16: 207-214.
- Sachan Sh, Nair RR, Khanna A, Singh K. CYP1A1, and GSTM1 genes polymorphism and its association with endometriosis: A pilot study. *Asian Pacific Journal of Reproduction* 2013; 2: 297-300.
- Barbosa AM, de Souza SR, Frare AB, Costa E Silva RC, da Costa IR, Freitas E Silva KS, et al. Association of CYP1A1 (cytochrome P450) MspI polymorphism in women with endometriosis. *Genet Mol Res* 2016; 15. doi: 10.4238/gmr.15038389.
- Arvanitis DA, Koumantakis GE, Goumenou AG, Matalliotakis IM, Koumantakis EE, Spandidos DA. CYP1A1, CYP19, and GSTM1 polymorphisms increase the risk of endometriosis. *Fertil Steril* 2003; 79 (Suppl.): 702-709.
- Trabert B, Schwartz SM, Peters U, De Roos AJ, Chen C, Scholes D, et al. Genetic variation in the sex hormone metabolic pathway and endometriosis risk: an evaluation of candidate genes. *Fertil Steril* 2011; 96: 1401-1406.
- Hadfield RM, Manek S, Weeks DE, Mardon HJ, Barlow DH, Kennedy SH, et al. Linkage and association studies of the relationship between endometriosis and genes encoding the detoxification enzymes GSTM1, GSTT1, and CYP1A1. *Mol Hum Reprod* 2001; 7: 1073-1078.
- Ozkan S, Murk W, Arici A. Endometriosis and infertility. epidemiology and evidence-based treatments. *Ann N Y Acad Sci* 2008; 1127: 92-100.
- Tsuchiya Y, Nakajima M, Yokoi T. Cytochrome P450-mediated metabolism of estrogens and its regulation in human. *Cancer Lett* 2005; 227: 115-124.
- Matsuzaka Y, Kikuti YY, Goya K, Suzuki T, Cai LY, Oka A, et al. Lack of an association human dioxin detoxification gene polymorphisms with endometriosis in Japanese women: results of a pilot study. *Environ Health Prev Med* 2012; 17: 512-517.
- André GM, Martins Trevisan C, Pedrucci IN, Fernandes RFM, Oliveira R, Christofolini DM, et al. The impact of FSHR gene polymorphisms Ala307Thr and Asn680Ser in the endometriosis development. *DNA Cell Biol* 2018; 37: 584-591.
- Shah PP, Saurabh K, Pant MC, Mathur N, Parmar D. Evidence for increased cytochrome P450 1A1 expression in blood lymphocytes of lung cancer patients. *Mutat Res* 2009; 670: 74-78.
- Peng DX, He YL, Qiu LW, Yang F, Lin JM. Susceptibility to endometriosis in women of Han Nationality in Guangdong Province associated with Msp I polymorphisms of cytochrome P450 1A1 gene. *Di Yi Jun Yi Da Xue Xue Bao* 2002; 22: 814-816.
- Babu KA, Reddy NP, Deendayal M, Kennedy S, Shivaji S. GSTM1, GSTT1, and CYP1A1 detoxification gene polymorphisms and their relationship with advanced stages of endometriosis in South Indian women. *Pharmacogenet Genomics* 2005; 15: 167-172.
- Rozati R, Reddy S, Giragalla SB, Bakshi H, Doddmaneni S, Khaja N, et al. The CYP1A1 and GSTM1 Genetic polymorphisms and susceptibility to Endometriosis in women from South India. *Int J Fertil Steril* 2008; 2: 105-112.
- Huber A, Keck CC, Hefler LA, Schneeberger C, Huber JC, Bentz EK, et al. Ten estrogen-related polymorphisms and endometriosis: a study of multiple gene-gene interactions. *Obstet Gynecol* 2005; 106: 1025-1031.
- Treloar SA, Wicks J, Nyholt DR, Montgomery GW, Bahlo M, Smith V, et al. Genomewide linkage study in 1,176 affected sister pair families identifies a significant susceptibility locus for endometriosis on chromosome 10q26. *Am J Hum Genet* 2005; 77: 365-376.
- Hediger ML, Hartnett HJ, Louis GM. Association of endometriosis with body size and figure. *Fertil Steril* 2005; 84: 1366-1374.