Iran J Reprod Med Vol. 13. No. 10. pp: 615-622, October 2015

Original article

In vitro developmental competence of bovine oocytes: Effect of corpus luteum and follicle size

Hamed Karami Shabankareh[1] D.V.M., Ph.D, Mohammad Hamed Shahsavari[1, 2], M.Sc., Hadi Hajarian[1] Ph.D., Gholamali Moghaddam[2], D.V.M., Ph.D.

1. *IVF and ET Laboratory, Department of Animal Sciences, Razi University, Kermanshah, Iran.*
2. *Department of Animal Sciences, Faculty of Agriculture, University of Tabriz, Tabriz, Iran.*

Corresponding Author:
Hamed Karami Shabankareh, Department of Animal Sciences, Razi University, Imam Khomeini Highway, Kermanshah, Iran.
Postal code: 6714644397
Email:
hkaramishabankareh@yahoo.com
Tel: (+98) 9188332484

Received: 11 January 2015
Revised: 21 June 2015
Accepted: 29 June 2015

Abstract

Background: Previous studies reported many discrepancies about the effects of corpus luteum (CL) and ovarian follicle size on the developmental competence of oocytes.

Objective: The aim of this study was to investigate the effects of CL and different size of follicle on the developmental potential of bovine oocytes.

Materials and Methods: After ovarian classification based on presence or absence of CL, sample follicles were placed in three groups according to their diameter; small (S; 3–6 mm), medium (M; 6–9 mm), and large (L; 10–20 mm). Collected oocytes in each group were subjected to the in vitro embryo production processes.

Results: Results showed that, the percentages of blastocyst obtained from oocytes originating from small and medium follicles of ovaries bearing a CL (CL+S-oocytes and CL+M-oocytes, respectively) were lower ($p<0.001$) than those of small and medium follicles of ovaries not bearing a CL (CL-S-oocytes and CL-M-oocytes, respectively) (30.8% and 33.6% vs. 36.9% and 38.7% respectively). Although, the percentages of blastocyst obtained from CL-M-oocytes and CL-L-oocytes were greater ($p< 0.001$) than those of CL+S-oocytes and CL+M-oocytes. There were no significant differences in the percentages of blastocyst formation between controls (C-oocytes), CL-S-oocytes and CL+L-oocytes.

Conclusion: According to the results of this study, the negative effect of CL on the developmental competence of bovine oocyte depends on the follicle size. Therefore, oocytes originating from large grown follicles were not influenced by negative effects of CL as much as those originating from small and medium follicles did.

Key words: *Bovine, Development, Oocyte.*

This article is extracted from Master thesis (Mohammad Hamed Shahsavari).

Introduction

There are distinct problems associated with in vitro embryo production (IVEP) of bovine embryos (1). It has been clearly established that oocyte quality (intrinsic quality of primary oocyte coupled to maturation conditions used) determines the rate of blastocyst production (2, 3). These problems include or are derived from the fact that the origin of the oocytes recovered from ovaries obtained from slaughtered animals (stage of estrous cycle, stage of follicular wave, origin of ovaries, etc.) is unknown or heterogeneous and therefore oocyte quality is very variable (4, 5). Previous studies confirmed that there was a relationship between the development of corpus luteum (CL) and the development of follicles which may cause asymmetry in the function of the reproductive organs in dairy cows (6, 7).

The evolution of the CL is associated with viviparity in mammals and is necessary for the production of progesterone [throughout the luteal phase of the estrous cycle to maintain pregnancy (if a conceptus is present) and during pregnancy, to decrease gonadotrophin secretion and prevent behavioral estrous occurring (8, 9). In addition, this small, transient endocrine gland secrets small

[Downloaded from ijrm.ir on 2026-11-16]

quantities of oestradiol-17ß, prostaglandins and a number of peptide hormones such as relaxin, oxytocin, oxytocin-related neurophysin-I, vasopressin and inhibin (10). A relationship between the development of CLs and the development of follicles, offers new evidence to support the existence of factors associated with heterogeneity in the developmental competence of oocytes (6). Boediono *et al* and Sugulle *et al* reported some discrepancies about positive and negative effects of CL on oocyte quality too (11, 12).

It appears that there are other factors associated with heterogeneity in the developmental competence of oocytes such as different stages of growth, atresia and different follicle sizes (5, 13, 14). Therefore, another source of heterogeneity in the developmental competence of oocytes is the size of the follicle from which the oocyte is obtained. In the pig as in other species, acquiring meiotic potential and subsequent developmental competence of the oocyte has been shown to be directly correlated to the follicle size (15, 16). In cows, it was reported that the size and the quality of the follicle of origin influence the developmental capacity of bovine oocytes(17, 18). Therefore, interaction between follicle size and the phase of follicular wave affected the efficiency of embryo production (19).

In the present study the effects of CL, different size of follicle and CL by follicle size interactions on the developmental potential of bovine oocytes were investigated.

Materials and methods

This experimental study was approved and performed under the guidelines of ethics committee for Animal use of Razi University. Unless otherwise stated, all chemicals were purchased from Sigma Chemical Company (St. Louis, MO, USA). This study was performed at Razi University's IVF & ET Laboratory, located in Kermanshah province; Iran (34°18′ N and 47°3′ E) from January 2013 to March 2013.

Oocyte collection

Bovine ovaries were recovered from female adult cows (Holstein friesian) 4-7 years of age with clinically normal reproductive tract after slaughtering. Static ovaries, those associated with pregnant cattle and those that had any pathological lesions such as cystic follicles (>20 mm in diameter) were not included in the study. Collected ovaries were transported to laboratory (within 2 h after slaughter) in a thermos flask in sterile normal salin containing penicillin (100 IU/ml) and streptomycin (50 mg/ml) at 30–35°C. To evaluate the developmental potential of oocytes originating from ovaries bearing a CL (CL+-oocytes) or not bearing a CL (CL-oocytes) ovaries were divided into three groups: CL+, CL-, and control ovaries. After ovaries classification based on presence or absence of CL, sample follicles were placed in three groups according to their diameters; small (S; 3–6 mm), medium (M; 6–9 mm) and large (L; 10–20 mm) (20). Cumulus-oocyte complexes (COCs) from these ovarian follicles were aspirated using 18-gauge needles attached to a 10 ml syringe. Therefore, 1330 oocytes were allocated into 7 experimental groups and 6 replicates based on their origin (CL+S-, CL+M-, CL+L-oocytes and CL-S-, CL-M-, CL-L-oocytes and C-oocytes) and their developmental potential were assessed following IVEP. In the control (C-oocytes, without selection based on presence or absence of a CL and follicle size) treatment: oocytes were cultured directly after recovery in the IVP process.

In vitro maturation (IVM)

After classification, the COCs were washed three times in the medium in which they were to be cultured. Oocytes were transferred in groups (8-10/group) into 50 ml droplets of IVM culture medium, consisting of Tissue culture medium-199 (TCM 199) supplemented with

[Downloaded from ijrm.ir on 2026-11-16]

0.23 mmol/L sodium pyruvate, 0.02 IU/ml pure human follicle-stimulating hormone (pFSH), 1 µg/ml 17ß estradiol, 50 ng/ml epidermal growth factor (EGF, NO. E-1257, Sigma), 10% (v/v) Fetal calf serum (FCS) and 50 µg/ml gentamicin.

The droplets containing oocytes were covered with pre-warmed (38.5°C) mineral oil and incubated for 24 hours (h) at 38.5°C in a CO2 incubator (5% CO2 in air, 90–95% relative humidity). After IVM, expansion rate was recorded.

In vitro fertilization (IVF)

Frozen bull semen was thawed and prepared by a swim-up procedure (21). After IVM, the oocytes were washed in washing solution and IVF solution twice each. Oocytes were then placed (6-7 COCs/50µl medium) in IVF- Tyrodes albumin lactate pyruvate (TALP) droplets (21).

The samples of capacitated spermatozoa were added to the oocytes in the droplets for a final concentration of 1×10^6 spermatozoa/ml. Subsequently, 2 µl of PHE mixture (penicillamine 20 µM, hypotaurine 10, and epinephrine 1 µM) was added to the suspension. Oocytes and spermatozoa were co-incubated for 18 h at 38.5°C under 5% CO2 in air with maximum humidity.

In vitro culture (IVC)

Approximately 18h post insemination (pi), presumptive zygotes were denuded by gentle vortexing and washed twice in HEPES-TCM 199 containing 4% BSA and once in potassium simplex optimization medium (KSOM) (22).

Groups of 25 to 30 zygotes were cultured in 400 µL of KSOM medium supplemented with 1 mg/mL BSA (KSOM1) overlaid with mineral oil for the first 48 h. Culture droplets were replenished at intervals of 48 h using KSOM medium containing 5% FCS (KSOM2) for the remaining days of culture. Embryonic cleavage was recorded on day 2 (the day of IVF considered as day 0) and blastocyst development was recorded on days 6, 7, 8 and 9 pi (Day 0 - day of IVF) and was expressed as blastocyst per total oocytes and per cleaved embryos.

Statistical analysis

Data of this study were collected and the percentage values were arcsine transformed. Analysis of variance was performed by using aov function of Stats package in R software (software statistical package R version 2.15.2). Means were compared using the Tukey HSD test and Student's t-test was used to compare two groups. Differences with a probability value of $p \leq 0.05$ were considered to be significant.

Results

Table I shows the developmental potential of oocytes originating from different follicle size of ovaries bearing a CL (CL+-oocytes) or not bearing a CL (CL--oocytes). In this experiment, follicle size had an effect on percentage of expanded oocytes and S-oocytes were considered to have the lowest percentage of expansion among groups. Presence or absence of CL by follicle size interactions had an effect on the percentage of blastocyst per oocytes and blastocyst per cleaved embryos ($p<0.001$). The percentage of blastocyst per oocytes of CL+.S-oocytes was the lowest (30.8%) among groups. There was no significant difference ($p=0.75$) in the percentage of blastocyst between oocytes originating from different follicle size of ovaries not bearing a CL (CL-S, CL-M and CL-L). However, there was heterogeneity in the developmental competence of oocytes originating from different follicle size of ovaries bearing a CL (CL+S, CL+M and CL+L). The percentage of blastocyst obtained from CL-.M-oocytes and CL-.L-oocytes was greater ($p< 0.001$) than that of CL+.S-oocytes and CL+.M-oocytes. However, there were no significant differences in the percentages of blastocyst formation between C-oocytes, CL-.S-oocytes and CL+L-oocytes. In this experiment, presence or absence of CL by follicle size interactions did not have any effect on expansion and cleavage rates among groups.

[Downloaded from ijrm.ir on 2026-11-16]

Table I. Developmental competence (mean±SE) of bovine oocytes originating from different follicle size of ovaries bearing a corpus luteum (CL) or not bearing a CL (replicates = 6)

Groups		Oocytes (n)	Expansion (%)	Cleavage (%)	Blastocysts per total oocytes (%)	Blastocysts per cleavage (%)
CL effect (main effect) [1]	C	251	89.2 ± 6.1	84.3 ± 3.0	35.0 ± 0.8	40.0 ± 0.7
	CL-	531	89.8± 3.1	84.2 ± 4.7	40.2 ± 3.6	45.7 ± 4.9
	CL+	548	85.1 ± 3.2	78.3 ± 2.8	31.4 ± 3.4	38.6 ± 4.2
Size effect (main effect) [2]	C	251	89.2 ± 6.1 [a]	84.3 ± 3.0	35.0 ± 0.8	40.0 ± 0.7
	S	435	72.1± 3.6[b]	79.1 ± 2.7	30.8 ± 3.8	36.2 ± 4.9
	M	494	81.3 ± 2.4[a]	84.4± 2.3	38.3 ± 3.2	41.0 ± 3.9
	L	150	90 ± 3.2[a]	80.5 ± 6.5	41.3 ± 5.4	46.8 ± 7.6
CL and size effect (Interactions) [3]	C	251	89.2 ± 6.1	84.3 ± 3.0	35.0± 0.8[b]	40.0± 0.7[ab]
	CL-.S	217	84.8 ± 5.1	81.9 ± 4.9	36.9 ± 4.0[ab]	39.9 ± 5.4[ab]
	CL-.M	236	88.0 ± 2.1	86.5± 2.6	38.7 ± 5.3[a]	43.6 ± 6.4[a]
	CL-.L	78	90.4 ± 4.8	84.1 ± 12.5	40.4 ± 8.9[a]	46.1 ± 13.1[a]
	CL+.S	218	82.4 ± 2.8	76.1 ± 2.4	30.8 ± 6.7[c]	36.8 ± 8.6[b]
	CL+.M	258	83.2 ± 3.5	82.0± 3.1	33.6 ± 3.7[bc]	38.8 ± 4.7[b]
	CL+.L	72	86.1 ± 6.9	76.6 ± 5.5	35.1 ± 6.7[b]	40.7 ± 8.9[ab]

CL: Corpus luteum Abc; Values in the column with different superscripts differ significantly ($p < 0.001$). (Student's *t*-test).
1; CL+, CL- and C: Ovaries bearing a CL, not bearing a CL and control ovaries, respectively.
2; S, M, and L: Oocytes originating from small, medium and large follicles, respectively.
3; CL-.S, CL-.M and CL-.L: Oocytes originating from small, medium and large follicles of ovaries not bearing a CL, respectively.
CL+.S, CL+.M and CL+.L: Oocytes originating from small, medium and large follicles of ovaries bearing a CL, respectively.

Discussion

There are many discrepancies about positive or negative effects of CL on reproductive parameters and the developmental potential of oocytes (23, 24). In addition, another source of heterogeneity in the developmental competence of oocytes is the size of the follicle from which the oocyte is obtained (17). Therefore, the present study was conducted to evaluate the developmental potential of oocytes originating from different follicle size of ovaries bearing a CL (CL+-oocytes) or not bearing a CL (CL-oocytes). The results of the present study showed that presence or absence of CL by follicle size interactions had an effect on the percentage of blastocyst per oocytes and blastocyst per cleaved embryos.

The percentage of blastocyst formation of oocytes originating from medium and large follicle of ovaries not bearing a CL (CL-M- and CL-L-oocytes) was greater than that of oocytes originating from small and medium follicle of ovaries bearing a CL (CL+S- and CL+M-oocytes) and this pattern was the same in blastocyst per oocytes and blastocyst per cleaved embryos. Furthermore, our results showed that there was a significant difference in the percentage of blastocyst formation between oocytes originating from the same follicle size of ovaries not bearing a CL and ovaries bearing a CL. It was shown that the percentages of blastocyst formation of oocytes originating from small and medium follicle of ovaries bearing a CL (CL+S- and CL+M-oocytes) were lower than that of ovaries not bearing a CL (CL-S- and CL-M-oocytes).

These observations are in agreement with the studies in humans, in which dominant follicles that appeared contralateral to the previous site from where ovulation occurred were healthier than ipsilateral follicles, leading to an enhanced pre-embryo quality in estrous cycles from which oocytes were obtained for IVF (23). Inhibitions of folliculogenesis and ovulation by presence of CL are more frequent on the ovary ipsilateral to the side of the previous ovulation (25, 26). In addition, the results of previous studies in ruminants revealed that the average number of good quality oocytes recovered from ovaries without a CL was comparably higher to the ovaries

[Downloaded from ijrm.ir on 2026-11-16]

with a CL, which can be effectively used for IVF (27, 28).

Local paracrine and autocrine influences exerted by the follicles and CL have been reported in many in vitro studies (20, 29, 30). Furthermore, receptors for progesterone and estrogen have been detected in the bovine ovary (31). Indeed, absence of dominance effects during the active phase of the CL, offers new evidence to support the hypothesis of Adams about the existence of suppressive effects of progesterone from the CL on lifespan of dominant follicles in hair breed sheep (32, 6). The mechanism by which progesterone inhibits follicular growth is through suppression of LH pulse frequency, which is critical for continued growth of large follicles (33). It is very well understood that progesterone functions on follicle development by a systemic pathway is through diminishing the frequency of LH pulses. Progesterone may also exert local effects on the growth of large antral follicles, in both luteal and non-luteal ovaries, independent of changes in gonadotrophin secretion (34).

Contreras-Solis *et al* reported the existence of not only a systemic, but also a possible intraovarian effect from the CL on ovine follicular dynamics, with a greater decrease in the number of follicles growing to large sizes in the ovary ipsilateral to the CL (6, 34). The same authors results support the existence of local inhibitory factors released from the CL. Inhibin, which is secreted by the CL of goats and cattle and secreted into ovarian venous blood of ewes, is widely known to affect ovarian follicular growth (6, 35, 36).

As stated earlier, previous studies reported many discrepancies about positive or negative effects of CL on reproductive parameters. For example, in an earlier study in cattle, however, the cleavage rates were found to be higher, and blastocyst production lower in oocytes collected from ovaries without a CL compared to those collected from ovaries with a CL (11). Similarly, in cattle, Boediono *et al* found no difference in the mean number of oocytes per ovary between CL-bearing and non-bearing ovaries and in other studies, the presence or absence of CL did not significantly influence the cleavage rate and blastocyst development (11, 12, 37).

Data from the present study contrasts with findings in cow that reported higher blastocyst yields have been obtained in vitro from oocytes collected from luteal phase ovaries and from ovaries bearing a CL compared to follicular phase ovaries and those without a CL, probably due to higher progesterone level in the circulation and constant follicular turnover (11, 24, 38). Furthermore, Penitente-Filho *et al* reported that ovaries with CL showed greater numbers of good quality oocytes than ovaries without CL (39).

In the pig as in other species, acquiring meiotic potential and subsequent developmental competence of the oocyte has been shown to be directly correlated to the follicle size (15, 16). In the present study, there was homogeneity in the developmental competence of oocytes originating from different follicle size of ovaries not bearing a CL (CL-S, CL-M and CL-L). However, follicle size had an effect on the developmental competence of oocytes originating from ovaries bearing a CL, where the percentage of blastocyst obtained from CL+L-oocytes was greater than that of CL+S-oocytes. These data contrasts with findings in a study performed in Japan on Black cow that reported the presence of CL within the ovaries does not affect the ovarian follicular dynamics after follicular aspiration and the developmental competence of collected COCs (40). In cows, it was reported that the size and the quality of the follicle of origin influence the developmental capacity of bovine oocytes. In cattle, goats and sheep oocytes originating from large follicles are more competent in terms of in vitro embryo development than oocytes from small follicles (17, 18, 41, 42). Our results confirmed these observations where S-oocytes were considered to have the lowest percentage of expansion among groups.

Furthermore, in the present study, it appears that all of the CL+-oocytes were not influenced by negative effects of CL and there was heterogeneity in the developmental

[Downloaded from ijrm.ir on 2026-11-16]

competence of CL+-oocytes due to the effect of follicle size. It seems that CL+.B-oocytes were not influenced by negative effects of CL as much as CL+.S-oocytes did. A possible reason is that as the follicular diameter increases to approximately 2mm (small follicles) and the oocytes increase in diameter from 110 to 120µm, developmental competency is acquired and the majority of oocytes become capable of supporting fertilization and embryonic development (43). Therefore, it can be hypothesized that CL mostly exerts negative effects on the growth of small growing follicles than large grown follicles.

On the other hand, the concentration of various biochemical constituents of follicular fluid may be changed with advancing the follicular growth and the presence or absence of a CL (44). Shabankareh *et al* suggested that follicular fluid from follicles in the bovine ovaries without a CL is more appropriate to use in oocyte maturation than those from ovaries with a CL because their data showed that the metabolites content of follicular fluid in the ovaries not bearing a CL were higher than those of the ovaries bearing a CL at the time of aspiration (20). These various biochemical constituents of follicular fluid may account for greater percentages of cleavage and blastocyst rates of CL-oocytes than that of CL+-oocytes in the present study. Furthermore, Shabankareh *et al* reported that glucose in the follicular fluid from large follicles being significantly higher compared with that from small and medium follicles in the CL—ovaries (20). Therefore, it appears that presence or absence of CL by follicle size interactions have an effect on oocyte environment and its developmental competence as observed in our study. After formation of the CL, new blood vessels were formed for the development of CL. The CL will receive the greatest rate of blood flow compared with other tissues in the ovary (21). Indeed, the increase in blood flow to the follicular cells in the ovaries without a CL results in an increased supply of gonadotrophins and other systemic biochemicals and hormonal factors necessary for follicular development (45).

In conclusion, the results of this in vitro study clearly demonstrated that CL exerts negative effects on the developmental competence of bovine oocytes. However, oocytes originating from large grown follicles were not influenced by negative effects of CL as much as those originating from small and medium follicles did. In fact, collection of ovaries categorized based on presence or absence of CL and different follicle sizes would be helpful in commercial production of cattle laboratory embryos.

Acknowledgments

The authors are grateful to Mr. Mehrdad Sadeghi for supplying the bovine ovaries from local slaughterhouse. Thanks to Ms. Golshan Azimi, Bahare Akhtarshenas and Mehrangiz Piri for helping in the IVF and ET laboratory. In addition, the authors would like to thank the Razi University for financial support.

Conflict of interest

None of the authors have any conflict of interest to declare.

References

1. Mermillod P, Marchal R. Oocyte of domestic mammals: a model for the study of in vitro maturation. *Contracep Fertil Sex* 1999; 27: 440-448.
2. Lonergan P, Gutiérrez-Adán A, Rizos D, Pintado B, De La Fuente J, Boland MP. Relative messenger RNA abundance in bovine oocytes collected in vitro or in vivo before and 20 hr after the preovulatory luteinizing hormone surge. *Mol Reprod Dev* 2003; 66: 297-305.
3. Alm H, Torner H, Löhrke B, Viergutz T, Ghoneim I, Kanitz W. Bovine blastocyst development rate in vitro is influenced by selection of oocytes by brillant cresyl blue staining before IVM as indicator for glucose-6-phosphate dehydrogenase activity. *Theriogenology* 2005; 63: 2194-2205.
4. Lonergan P, Fair T, Corcoran D, Evans A. Effect of culture environment on gene expression and developmental characteristics in IVF-derived embryos. *Theriogenology* 2006; 65: 137-152.
5. Karami-Shabankareh H, Mirshamsi SM. Selection of developmentally competent sheep oocytes using the brilliant cresyl blue test and the relationship to follicle

[Downloaded from ijrm.ir on 2026-11-16]

size and oocyte diameter. *Small Ruminant Research* 2012; 105: 244-249.
6. Contreras-Solis I, Diaz T, Lopez G, Caigua A, Lopez-Sebastian A, Gonzalez-Bulnes A. Systemic and intraovarian effects of corpus luteum on follicular dynamics during estrous cycle in hair breed sheep. *Anim Reprod Sci* 2008; 104: 47-55.
7. Penitente-Filho JM, Carrascal E, Oliveira FA, Zolini AM, Oliveira CT, Costa Soares ÍA, et al. Influence of dominant follicle and corpus luteum on recovery of good quality oocytes for in vitro embryo production in Cattle. *Brit Biotech J* 2014; 4: 1305-1312.
8. Mann GE, Lamming GE, Robinson RS, Wathes DC. The regulation of interferon-tau production and uterine hormone receptors during early pregnancy. *J Reprod Fertil Suppl* 1998; 54: 317-328.
9. Powell ML, Kavanaugh S, Sower SA. Identification of a functional corpus luteum in the Atlantic hagfish, Myxine glutinosa. *Gen Comp Endocrinol* 2006; 148: 95-101.
10. Field PA. Relaxin and other luteal secretory peptides: cell localization and function in the ovary. In: Familiari G, Makabe S, Motta PM (eds.). Ultrastructure of the Ovary. Kluwer Academic Publishers; 1991: 177-198.
11. Boediono A, Rajamahendran R, Saha S, Sumantri C, Suzuki T. Effect of the presence of a CL in the ovary on oocyte number, cleavage rate and blastocyst production in vitro in cattle. *Theriogenology* 1993; 43: 169.
12. Sugulle AH, Dochi O, Koyama H. Developmental competence of bovine oocytes selected by Brilliant Cresyl Blue Staining: Effect of the presence of corpus luteum on embryo development. *J Mammalian Ova Research* 2008; 25: 50-55.
13. Mirshamsi SM, Shabankareh HK. Selection of developmentally competent sheep zygotes using the Brilliant Cresyl Blue (BCB) test, after IVF. *Small Ruminant Research* 2012; 105: 250-254.
14. Mirshamsi SM, KaramiShabankareh H, Ahmadi-Hamedani M, Soltani L, Hajarian H, Abdolmohammadi A. Combination of oocyte and zygote selection bybrilliant cresyl blue (BCB) test enhanced prediction of developmental potential to the blastocyst in cattle. *Anim Reprod Sci* 2013; 136: 245–251.
15. Motlik J, Fulka J. Factors affecting meiotic competence in pig oocytes. *Theriogenology* 1986; 25: 87-96.
16. Yoon K-W, Shin T-Y, Park J-I, Roh S, Lim JM, Lee B-C, et al. Development of porcine oocytes from preovulatory follicles of different sizes after maturation in media supplemented with follicular fluids. *Reprod Fertility Dev* 2001; 12: 133-139.
17. Beker-van Woudenberg AR, Zeinstra EC, Roelen BA, Colenbrander B, Bevers MM. Developmental competence of bovine oocytes after specific inhibition of MPF kinase activity: effect of estradiol supplementation and follicle size. *Anim Reprod Sci* 2006; 92: 231-240.
18. Blondin P, Sirard M-A. Oocyte and follicular morphology as determining characteristics for developmental competence in bovine oocytes. *Mol Reprod Dev* 1995; 41: 54-62.
19. Machatkova M, Krausova K, Jokesova E, Tomanek M. Developmental competence of bovine oocytes: effects of follicle size and the phase of follicular wave on in vitro embryo production. *Theriogenology* 2004; 61: 329-335.
20. Shabankareh HK, Kor NM, Hajarian H. The influence of the corpus luteum on metabolites composition of follicular fluid from different sized follicles and their relationship to serum concentrations in dairy cows. *Anim Reprod Sci* 2013; 140: 109-114.
21. Parrish J, Susko-Parrish J, Leibfried-Rutledge M, Critser E, Eyestone W, First N. Bovine in vitro fertilization with frozen-thawed semen. *Theriogenology* 1986; 25: 591-600.
22. Lawitts JA, Biggers JD. Culture of preimplantation embryos. *Methods Enzymol* 1993; 225: 153-164.
23. Fukuda M, Fukuda K, Andersen CY, Byskov AG. Contralateral selection of dominant follicle favours pre-embryo development. *Hum Reprod* 1996; 11: 1958-1962.
24. Pirestani A, Hosseini SM, Hajian M, Forouzanfar M, Moulavi F, Abedi P, et al. Effect of ovarian cyclic status on in vitro embryo production in cattle. *Int J Fertil Steril* 2010; 4: 172-175
25. Filho FT, Santos MHB, Carrazzoni PG, Paula-Lopes FF, Neves JP, Bartolomeu CC, et al. Follicular dynamics in Anglo-Nubian goats using transrectal and transvaginal ultrasound. *Small Ruminant Research* 2007; 72: 51-56.
26. Islam MR, Khandoker MA, Afroz S, Rahman MG, Khan RI. Qualitative and quantitative analysis of goat ovaries, follicles and oocytes in view of in vitro production of embryos. *J Zhejiang Univ Sci B* 2007; 8: 465-469.
27. Kumar N, Paramasivan S, Sood P, Singh M. Micrometry of different category oocytes recovered from goat ovaries. *Indian J Anim Sci* 2004; 74.
28. Jamil H, Samad HA, Qureshi ZI, Rehman NU, Lodhi LA. Harvesting and evaluation of riverine buffalo follicular oocytes. *Turk J Vet Anim Sci* 2008; 32: 25-30.
29. Findlay J, Drummond A, Dyson M, Baillie A, Robertson D, Ethier J-F. Production and actions of inhibin and activin during folliculogenesis in the rat. *Mol Cell Endocrinol* 2001; 180: 139-144.
30. Shabankareh HK, Habibizad J, Torki M. Corpus luteum function following single and double ovulation during estrous cycle in Sanjabi ewes. *Anim Reprod Sci* 2009; 114: 362-369.
31. Berisha B, Pfaffl MW, Schams D. Expression of estrogen and progesterone receptors in the bovine ovary during estrous cycle and pregnancy. *Endocrine* 2002; 17: 207-214.
32. Adams G. Comparative patterns of follicle development and selection in ruminants. *J Reprod Fertil Suppl* 1999; 54: 17-32.
33. Effects of a 6-Day treatment with medroxyprogesterone acetate after prostaglandin F2α-induced luteolysis at midcycle on antral follicular development and ovulation rate in nonprolific

[Downloaded from ijrm.ir on 2026-11-16]

western white-Faced Ewes. *Biol Reprod* 2003; 68, 1403–1412.
34. Bartlewski PM, Beard AP, Rawlings NC. Ultrasonographic study of the effects of the corpus luteum on antral follicular development in unilaterally ovulating western white-faced ewes. *Anim Reprod Sci* 2001; 65: 231-244.
35. Sangha G, Sharma R, Guraya S. Biology of corpus luteum in small ruminants. *Small Rumin Res* 2002; 43: 53-64.
36. Campbell BK, Picton HM, Mann GE, McNeilly AS, Baird DT. Effect of steroid-and inhibin-free ovine follicular fluid on ovarian follicles and ovarian hormone secretion. *J Reprod Fertil* 1991; 93: 81-96.
37. Vassena R, Mapletoft RJ, Allodi S, Singh J, Adams GP. Morphology and developmental competence of bovine oocytes relative to follicular status. *Theriogenology* 2003; 60: 923-932.
38. Taylor C, Rajamahendran R. Effect of mid-luteal phase progesterone levels on the first wave dominant follicle in cattle. *Can J Anim Sci* 1994; 74: 281-285.
39. Penitente-Filho JM, Jimenez CR, Zolini AM, Carrascal E, Azevedo JL, Silveira CO, et al. Influence of corpus luteum and ovarian volume on the number and quality of bovine oocytes. *Anim Sci J* 2015; 86: 148-152.
40. Takuma T, Otsubo T, Kurokawa Y, Otoi T. 416 Effects of the corpus luteum within the ovary on the follicular dynamics after follicular aspiration and on the developmental competance of aspirated oocytes. *Reprod Fertil Deve* 2006; 19: 324.
41. Lequarre AS, Vigneron C, Ribaucour F, Holm P, Donnay I, Dalbiès-Tran R, et al. Influence of antral follicle size on oocyte characteristics and embryo development in the bovine. *Theriogenology* 2005; 63: 841-859.
42. Crozet N, Ahmed-Ali M, Dubos M. Developmental competence of goat oocytes from follicles of different size categories following maturation, fertilization and culture in vitro. *J Reprod Fertil* 1995; 103: 293-298.
43. Hyttel P, Fair T, Callesen H, Greve T. Oocyte growth, capacitation and final maturation in cattle. *Theriogenology* 1997; 47: 23-32.
44. Shabankareh HK, Habibizad J, Sarsaifi K, Cheghamirza K, Jasemi VK. The effect of the absence or presence of a corpus luteum on the ovarian follicular population and serum oestradiol concentrations during the estrous cycle in Sanjabi ewes. *Small Rumin Res* 2010; 93: 180-185.
45. Acosta TJ, Miyamoto A. Vascular control of ovarian function: ovulation, corpus luteum formation and regression. *Anim Reprod Sci* 2004; 82: 127-140.

[Downloaded from ijrm.ir on 2026-11-16]