

## Pregnancy outcome of “delayed start” GnRH antagonist protocol versus GnRH antagonist protocol in poor responders: A clinical trial study

Abbas Aflatoonian<sup>1</sup> M.D., Robabe Hosseiniadat<sup>2</sup> M.D., Ramesh Baradaran<sup>1</sup> M.D., Maryam Farid Mojtahedi<sup>3</sup> M.D.

1. Research and Clinical Center for Infertility, Reproductive Sciences Institute, Shahid Sadoughi University of Medical Sciences, Yazd, Iran.

2. Department of Obstetrics and Gynecology, School of Medicine, Kerman University of Medical Sciences, Kerman, Iran.

3. Department of Obstetrics and Gynecology, Endocrinology and Female Infertility Unit, Roozhan Arash Women's Health Research and Educational Hospital, Tehran University of Medical Sciences, Tehran, Iran.

### Corresponding Author:

Robabe Hosseiniadat,  
Department of Obstetrics and Gynecology, School of Medicine, Kerman University of Medical Sciences, Kerman, Iran. Post code: 7616913911  
Email: robabehosseiniadat@yahoo.com  
Tel: (+98) (35 38247085)

Received: 16 October 2016

Revised: 16 January 2017

Accepted: 16 February 2017

Updated: 4 September 2021

### Abstract

**Background:** Management of poor-responding patients is still major challenge in assisted reproductive techniques (ART). Delayed-start GnRH antagonist protocol is recommended to these patients, but little is known in this regards.

**Objective:** The goal of this study was assessment of delayed-start GnRH antagonist protocol in poor responders, and in vitro fertilization (IVF) outcomes.

**Materials and Methods:** This randomized clinical trial included sixty infertile women with Bologna criteria for ovarian poor responders who were candidate for IVF. In case group (n=30), delayed-start GnRH antagonist protocol administered estrogen priming followed by early follicular-phase GnRH antagonist treatment for 7 days before ovarian stimulation with gonadotropin. Control group (n=30) treated with estrogen priming antagonist protocol. Finally, endometrial thickness, the rates of oocytes maturation, embryo formation, and pregnancy were compared between two groups.

**Results:** Rates of implantation, chemical, clinical, and ongoing pregnancy in delayed-start cycles were higher although was not statistically significant. Endometrial thickness was significantly higher in case group. There were no statistically significant differences in the rates of oocyte maturation, embryo formation, and IVF outcomes between two groups.

**Conclusion:** There is no significant difference between delayed-start GnRH antagonist protocol versus GnRH antagonist protocol.

**Keywords:** Pregnancy outcome, Poor responder, In vitro fertilization, GnRH antagonist protocol.

Registration ID in IRCT: IRCT2015100424335N1

### Introduction

Some women undergoing infertility treatments are poor responders to the routine controlled ovarian hyper-stimulation (COH) protocols (1, 2). The “poor responder” was initially defined by Garcia *et al* in 1981 (3). Despite great advances in assisted reproductive technologies (ART) has been established, but the management of poor-responder patients is still a major challenge (4). In IVF programs, the incidence of poor ovarian response (POR) after ovarian stimulation is variable from 9-25% (5). Cause of poor response can be related to age, endometriosis, genetics factors, obesity, or may be iatrogenic such as surgery, radio, and chemotherapy (6, 7). Recently, due to changing social structure and the worldwide trend of delaying marriage and childbirth, there has been increased interest in

improving the reproductive ability of older women (6). There are no certain definitions of poor response. Recently, the European Society for Human Reproduction and Embryology (ESHRE) has established a new standardized definition for poor responders that called “Bologna criteria” to help identify of these challenging patients for clinical trials and optimal treatment management (8). According to the ESHRE agreement, at least two of the following three features indication must be present:

1. Advanced maternal age ( $\geq 40$ ) or any other risk factor for poor ovarian response,
2. Previous poor response (cycles cancelled or  $\leq 3$  oocytes with a conventional stimulation protocol),
3. Abnormal ovarian reserve test (ORT) (AMH  $< 0.5$ - $1.1$  ng/mL or AFC  $< 5$ - $7$  follicles).

In the absence of above criteria, two previous incidents of poor ovarian response after maximal stimulation are sufficient to define a poor responder (8).

Various factors have been associated with a poor response. Alterations in intra ovarian factors or gonadotropin receptor regulation could contribute to suboptimal response (9, 10). Poor responses may result from a shortened follicular phase with limited ability to recruit a follicular cohort or from different sensitivity of early antral follicles to FSH due to follicular different developmental stages with various FSH receptor levels leading to heterogeneity of antral follicle (11, 12). During the last days of the menstrual cycle, FSH increases to preserve antral follicles from atresia and ensure their next growth step (13). Depending on antral follicles inherent sensitivity to FSH, some of them, specially larger follicles are able to respond to the lower levels of FSH better than others, and to start their maturity during the late luteal phase, and leading to asynchronous growth during the first days of the subsequent cycle with COH (14, 15, 16). This lack of coordination in size causes fewer follicles respond to COH (17).

COH protocols for poor responders are designed to limited early follicle selection in the luteal phase and optimize the follicular hormonal environment and antral follicle responsiveness (14, 16). Oral contraception pills (OCPs) or gonadotropin-releasing hormone agonist (GnRH agonist) long protocol in the late luteal phase can suppress FSH and premature dominant follicle selection. For poor responders, GnRH agonist long protocol or OCPs before GnRH antagonist may cause over suppression of ovarian function and desensitization of the ovary, leading to reduction in the number of mature oocytes and increase the dose of gonadotropins (18-20).

Administration of luteal estradiol ( $E_2$ ) to GnRH antagonist protocols resulted in a reduction of both antral follicular sizes and heterogeneity in the early follicular phase, and increases the number of follicles due to FSH suppression (21-23). Another treatment for these patients is late luteal or early follicular GnRH antagonist administration that

suppresses FSH levels and reduces baseline antral follicular size and heterogeneity (24). Recently pretreatment  $E_2$  and start of antagonist in early follicular phase from day 2-8 before gonadotropin therapy (double suppression) appears to improve ovarian response during COH and may result in more uniform follicular development. This protocol named delayed-start protocol (25). There is no sufficient research from efficiency of current protocol (delayed-start protocol), thus we planned a study about the effect of delayed-start protocol with GnRH antagonist in outcome of ART cycle in poor responders.

The objective of this study was to assess the effect of delayed-start GnRH antagonist protocol versus GnRH antagonist protocol in ovarian poor responders.

## Materials and methods

This randomized clinical trial was recruited in Yazd Research and Clinical Center for Infertility between March and September 2015. Totally, 60 infertile women between 18-45 yr old with Bologna criteria for ovarian poor responders were allocated in this study (8). Women with history of endocrine disorders, severe endometriosis, and azoospermia in their husband were excluded. Women were allocated randomly in two groups (delayed and control) according to random number table method.

Control group ( $n=30$ ) treated with estrogen priming antagonist protocol and case group ( $n=30$ ) with delayed-start GnRH antagonist protocol. In both group 4mg estradiol valerate tablet ( $E_2$ ) (Aburaihan Co., Tehran, Iran) was administered from day 21 in previous cycle and continues for 10 days. In delayed group immediately after administration of  $E_2$ , patients received GnRH antagonist cetrotide (0.25 mg cetrorelix acetate; Merck Serono, Germany) subcutaneously for 7 days, and then we initiated ovarian stimulation with 375 IU FSH (Gonal-f; Merck Serono, Germany).

In control group immediately after administration of  $E_2$ , ovarian stimulation with 375 IU FSH (Gonal-f; Merck Serono, Germany) was performed. In both groups when follicle size was 12 mm, cetrotide added

again to prevent premature ovulation and continued until the hCG trigger. When at least two follicles achieved 17 mm in diameter, Human chorionic gonadotropin (hCG) (Choriomon 10000 IU, IBSA Institute, Switzerland) was administered for final oocyte maturation. Oocyte retrieval performed under transvaginal ultrasound guidance 34-36 hr after hCG triggering. Intra-cytoplasmic sperm injection performed with mature oocytes (metaphase II [MII]) in all cycles.

Day 2 after oocyte retrieval embryos were categorized in four grades from A (high quality) to D (low quality) depending on the number of blastomeres, fragmentation, multinucleation and symmetry; and were transferred with COOK catheter (COOK catheter, USA) (26).

The main primary outcomes measured were total and mature (MII) oocytes number collected after E<sub>2</sub> priming antagonist protocol versus delayed-start ovarian stimulation protocol. Secondary outcomes were oocyte maturity rate (MII number /total oocytes number), oocyte yield (total oocytes number /antral follicle count [AFC]), mature oocyte yield (MII number/AFC), total dosage of gonadotropin, ovarian stimulation days, and fertilization rate (two-pronuclear [2PN]/ MII, 16 hr after Intra-cytoplasmic sperm injection treatment) and embryo number. Other secondary outcomes were assessed based on positive serum  $\beta$ hCG test (chemical pregnancy), 14 days after embryo transfer and observation of gestational sac on transvaginal ultrasound examination (clinical pregnancy), 3 wk after positive serum  $\beta$ hCG.

Implantation rate was defined by the number of gestational sacs divided on the number of transferred embryos in each group. The Ongoing pregnancy rate was assessed as the presence of fetal heart activity by ultrasound after 12 wk. The miscarriage rate was the number of miscarriages before 20 weeks gestation per number of women with a positive clinical pregnancy.

### **Ethical consideration**

Our study proposal was approved by Ethics Committee Shahid Sadoughi University of

Medical Sciences, Yazd, Iran. Informed written consent was obtained from all couples.

### **Statistical analysis**

Data was analyzed using Statistical Package for the Social Sciences 20.0 (SPSS, SPSS Inc, Chicago, Illinois). Continuous data were presented as mean $\pm$ SD and assessed by Mann-Whitney test and independent Student's *t*-test. Enumeration data were compared by chi-square or Fisher exact test. A P-value<0.05 was considered statistically significant.

### **Results**

Totally, 72 poor responder women entered to study. 12 women were excluded and finally data of 60 women were analyzed (Figure 1). Baseline characteristics of the patients are presented in table I. The mean age of participants was 38.76 $\pm$ 3.46 in cases and 40.30 $\pm$ 3.01 in controls; however this difference was not statistically significant. There was no significant difference in Infertility duration, type of infertility, basal FSH level, anti-Mullerian hormone (AMH), antral follicle count (AFC), and previous retrieval cycles between two studied groups (Table I).

HCG day estradiol and progesterone, gonadotropins dose, and days of ovarian stimulation were similar between two groups (Table II). Endometrial thickness in triggering day was significantly higher in delayed group compared to those of control group ( $p=0.04$ ) (Table II). There was no significant difference in the number of total and mature (MII) oocyte, obtained and transferred embryos between two studied groups, although there was lower mean in case group versus control group. There were no significant differences in the maturation rate (MII/total oocytes), oocyte yield (Oocytes/AFC), mature oocyte yield (MII/AFC), fertilization rate (2PN/MII) between two groups (Table III).

Although it was not statistically significant difference between two groups in ART outcomes but in delayed-start protocol cycles, chemical (13.30% vs. 3.30%), clinical (13.30% vs. 3.30%) and ongoing (6.66% vs. 3.33%) pregnancy rate and implantation rate (11.4% vs. 3.8%) was higher than other group (Table IV).

**Table I.** Baseline characteristics of study participants in both groups

Variable	Case group (n=30)	Control group (n=30)	p-value
Age (yr)	38.76 ± 3.46	40.30 ± 3.01	0.07
Infertility duration (yr)	6.15 ± 4.77	6.70 ± 6.60	0.60*
Infertility type			0.19
Primary	19 (63.3%)	13 (43.3%)	
Secondary	11 (36.7%)	17 (56.7%)	
Baseline FSH (IU/L)	8.05 ± 2.17	7.76 ± 2.12	0.59
AMH (ng/ml)	0.78 ± 0.49	0.92 ± 0.57	0.34*
AFC	4.86 ± 2.09	4.76 ± 2.43	0.86
Previous COH cycle	0.83 ± 0.87	0.63 ± 0.88	0.38*

FSH=follicle-stimulating hormone; AMH: Anti-Mullerian hormone; AFC: Antral Follicle Count; COH: Controlled Ovarian Hyper stimulation. Continuous data presented as mean ± SD with p-values obtained from Independent- Samples *t* test; Enumeration data presented as n (%) with p-value obtained from Chi-Square or fisher exact tests. \* Mann-Whitney test

**Table II:** Cycle characteristics in case and control groups

Variable	Case group (n=30)	Control group (n=30)	p-value
hCG day Estradiol (pg/ml)	1152.26 ± 667.55	1233.80 ± 780.64	0.66
hCG day Progesterone (pg/ml)	0.81 ± 0.61	0.58 ± 0.43	0.11
hCG day endometrial thickness (mm)	10.18 ± 2.08	9.13 ± 1.84	0.04
Days of ovarian stimulation	11.60 ± 2.5	12.76 ± 1.50	0.87
Gonadotropin dose (IU)	3372.50 ± 1055.24	3617.50 ± 759.42	0.30*

hCG: human Chorionic Gonadotropin. Continuous data presented as mean ± SD with p-values obtained from Independent- Samples *t* test; \* Mann-Whitney test

**Table III:** Cycle outcomes of study participants in both groups

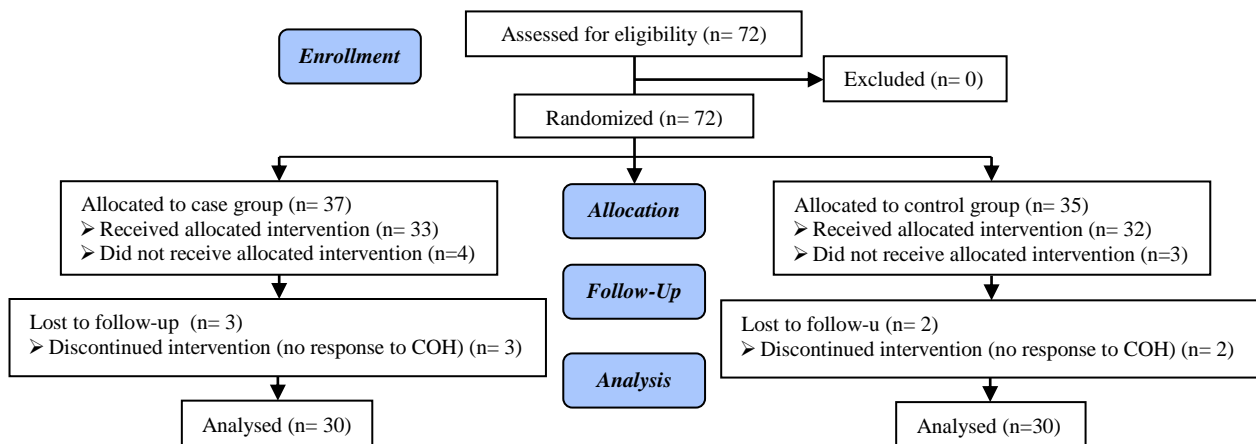
Variable	Delayed group (n=30)	Control group (n=30)	p-value
Total oocytes number	3.63 ± 3.02	5.06 ± 4.37	0.14*
MII Oocytes number	2.86 ± 2.50	4.33 ± 3.72	0.07*
Maturation rate (%)	77	85	0.29*
Oocyte yield	0.86 ± 0.81	1.11 ± 0.87	0.27*
MII oocyte yield	0.66 ± 0.60	0.94 ± 0.78	0.13*
2PN umber	1.63 ± 1.67	2.66 ± 2.61	0.10**
Fertilization rate (%)	55	62%	0.48*
Embryos umber	1.40 ± 1.56	2.13 ± 1.92	0.07**
Transferred embryos number	1.13 ± 1.04	1.56 ± 0.89	0.09***
Transferred Embryos quality			
A	2 (5.88%)	5 (10.63%)	
B	19 (56.0%)	28 (59.57%)	
C	11 (32.35%)	12 (25.53%)	0.91*
D	2 (5.88%)	2 (4.25%)	

MII oocyte: metaphase II oocyte; 2PN = Tow pronuclei; AFC: Antral Follicle Count. Continuous data presented as mean ± SD with \* Chi-Square, \*\* Mann-Whitney test, \*\*\* Independent- Samples *t* test

**Table IV.** IVF outcomes in case and control groups

Variable	Case group (n=30)	Control group (n=30)	p-value*
Chemical pregnancy	4 (13.30%)	1 (3.30%)	0.35
Clinical pregnancy	4 (13.30%)	1 (3.30%)	0.35
Ongoing pregnancy	2 (6.66%)	1 (3.33%)	1
Implantation rate (%)	11.4%	3.8%	0.27
Miscarriage rate	2 (50%)	0 (0.0%)	0.49

Enumeration data presented as N (%) with p-value obtained from Chi-Square or Fisher exact tests. \*Chi-Square or fisher exact test.

**Figure1.** Consort flow diagram.

## Discussion

Despite frequent developments in assisted reproduction, there is no agreement on the effective stimulation protocol for poor ovarian responder patients. In the present study, we compared ART outcomes in poor responders with early follicular GnRH antagonists pretreatment for 7 days after preceding late luteal estrogen priming and before the beginning of ovarian stimulation (delayed start protocol) with GnRH antagonists with estrogen priming without GnRH antagonists pretreatment. Our results showed delayed protocol in poor responders can improve pregnancy and implantation rate although number of oocyte and embryo was lower in this group. Endometrial thickness was significantly higher in study group. It was showed no significant differences in other parameters and ART outcomes.

The definition and treatment of patients with poor response to controlled ovarian hyper stimulation remains controversial. The heterogeneity in patients and inclusion criteria has increased the difficulty in comparing outcomes between the various treatment approaches that have been suggested by different investigators (26). One alternative approach introduced in the 1980, was oocyte donation (27). While oocyte donation has become a highly successful option with greater than 50% live birth rate for poor responders most patients are anxious to achieve a pregnancy with others oocytes. Therefore some protocols suggested improving ART outcomes in poor responders. Managing poor response cycles, however, continues major challenges for the reproductive medicines. For more than one decade, GnRH antagonists have been available in IVF preparation. GnRH antagonists prevent premature LH surge without early suppression of follicular development (28).

Pu and colleagues in a meta-analysis compared the use of GnRH agonist protocols with GnRH antagonists of 14 prospective randomized controlled trials. Their result showed no significant difference in IVF outcomes (29). Several approaches have been proposed and investigated to improve poor responder's treatment outcomes with GnRH antagonists. During the late luteal phase, FSH levels increase progressively to

antral follicles ensures growth. Larger follicles are more sensitive to rising levels of FSH and therefore begin to develop during the late luteal phase (30, 31). This discrepancy is detrimental in COH and confused synchronized maturation of the follicular cohort. Coordination of the early antral follicles has been improved by two methods, late luteal estradiol and late luteal or early follicular administration of a GnRH antagonist (26).

Fanchin *et al* in 2003 defined the use of luteal E<sub>2</sub> to decrease the premature gradual exposure of follicles to FSH in the late luteal phase. By using the late luteal E<sub>2</sub>, there was a significant reduction of mean follicular size at baseline and improvement in overall follicular size coordination (22, 23). In addition, Fanchin *et al* in another study used one dosage (3mg) of GnRH antagonist in the late luteal phase (on day 25) in normal responders and described that it decreased the exposure of early antral follicles to gradient levels of FSH and synchronized follicular size on day 2 of the cycle pretreated with GnRH antagonist (24).

In another study, Dragisic *et al* demonstrated that the further suppression with either luteal E<sub>2</sub> patch and 3 days luteal-phase GnRH antagonist appears to be a new option in the treatment of poor responders and yielded superior results compared to patients' prior IVF cycles (32). However, studies of Fanchin *et al* and Dragisic *et al* were not randomized controls trial. Weitzman *et al* in 2009 demonstrates that the use of E<sub>2</sub> patch and 3 days GnRH antagonist during the preceding luteal phase in patients with poor history can provide IVF outcomes similar to the microdose GnRH agonist protocol (26).

Ata *et al* in 2011 found similar results in IVF outcomes between luteal E<sub>2</sub>/GnRH antagonists until starting menstruation, and microdose GnRH agonist flare protocol (33). In our study, suppression with GnRH antagonist in early follicular phase was greater than two previous above studies (7 days compare with 3 days) also poor responders inclusion criteria were different. In above studies, E<sub>2</sub> and GnRH antagonists administered together but in our study GnRH antagonists started after E<sub>2</sub> priming. Shastri *et al* described that in young poor responders who treated with a luteal E<sub>2</sub>/3 days GnRH antagonist (E<sub>2</sub>/ANT) protocol, IVF outcomes improved versus an OCP microdose

leuprolide protocol (OCP-MDL) (34). In this study mean age of patients was 32 and E<sub>2</sub> and GnRH antagonist administered together in late luteal phase but in our study mean age were 38.

Mashayekhi *et al* in 2013 compared the mild antagonist and microdose GnRH agonist flare protocols on IVF outcome in poor responders. they administrated clomiphene citrate before gonadotropin in mild protocol. Endometrial thickness, number of retrieved oocytes, mature oocytes and implantation rate were significantly higher in mild antagonist protocol. Clomiphene citrate improved outcome in antagonist protocol. But in our study suppression before gonadotropin administration improved outcome (35).

Cakmak *et al* demonstrated that the delayed-start protocol (10 days estrogen in late luteal phase then early follicular-phase GnRH antagonist for 7 days before COH) improves ovarian response and IVF outcomes in poor responders compared with E<sub>2</sub> pretreatment protocol. They showed that double suppression was more effectively from E<sub>2</sub> suppression alone (25). Cakmak *et al* study showed in E<sub>2</sub> pretreatment group, ovarian stimulation with gonadotropins was started on cycle day 2 of menstruation after administration E<sub>2</sub>. In case group GnRH antagonists also started on cycle day 2 of menstrual cycle after E<sub>2</sub>, but in our study COH in control group and GnRH antagonists in study group started immediately after completion of the E<sub>2</sub>. Therefore, suppression was shorter in our study.

### Conclusion

In summary based on this study, we concluded that delayed-start protocol in poor responders slightly but no significantly improves pregnancy and implantation rate. Moreover, delayed-start protocol should be investigated in larger prospective randomized studies. Also evaluation of delayed-start protocol without E<sub>2</sub> priming can compare with other poor protocols such as microdose GnRH agonist flare protocol. According to previous studies (that administrated 3 dose GnRH antagonists before COH) and for reduce costs and treatment duration, it is suggested to design a study for evaluation of lower administration days GnRH antagonist before COH.

### Acknowledgments

The authors thank staff from the Research and Clinical Center for Infertility, Shahid Sadoughi University of Medical Sciences, Yazd, Iran, for their skillful technical assistance during the course of this study. The study was supported financially by Research Deputy of Shahid Sadoughi University of Medical Sciences, Yazd, Iran. Special thanks to M. Gandom, A. Kermani, and Z. Sheibani.

### Conflict of interest

The authors declare they have no conflict of interest.

### References

1. Davar R, Rahsepar M, Rahmani E. A comparative study of luteal estradiol pre-treatment in GnRH antagonist protocols and in micro dose flare protocols for poor-responding patients. *Arch Gynecol Obstet* 2013; 287: 149-153.
2. Tarlatzis BC, Zepiridis L, Grimbizis G, Bontis J. Clinical management of low ovarian response to stimulation for IVF: a systematic review. *Hum Reprod Update* 2003; 9: 61-76.
3. Garcia J, Jones GS, Acosta AA, Wright Jr GL. Corpus luteum function after follicle aspiration for oocyte retrieval. *Fertil Steril* 1981; 36: 565-572.
4. Kahraman K, Berker B, Atabekoglu CS, Sonmezer M, Cetinkaya E, Aytac R, Satiroglu H. Microdose gonadotropin-releasing hormone agonist flare-up protocol versus multiple dose gonadotropin-releasing hormone antagonist protocol in poor responders undergoing intracytoplasmic sperm injection-embryo transfer cycle. *Fertil Steril* 2009; 91: 2437-2444.
5. Keay SD. Poor ovarian response to gonadotrophin stimulation-The role of adjuvant treatments. *Hum Fertil* 2002; 5 (Suppl.): S46-52.
6. Polyzos NP, Tournaye H. Poor ovarian responders: to meta-analyses or not, that is the question. *Hum Reprod* 2014; det426.
7. Akande VA, Fleming CF, Hunt LP, Keay SD, Jenkins JM. Biological versus chronological ageing of oocytes, distinguishable by raised FSH levels in relation to the success of IVF treatment. *Hum Reprod* 2002; 17: 2003-2008.
8. Ferraretti A, La Marca A, Fauser BC, Tarlatzis B, Nargund G, Gianaroli L, ESHRE working group on Poor Ovarian Response Definition. ESHRE consensus on the definition of 'poor response' to ovarian stimulation for in vitro fertilization: the Bologna criteria. *Hum Reprod* 2011; 26: 1616-1624.
9. De Sutter P, Dhont M. Poor response after hormonal stimulation for in vitro fertilization is not related to ovarian aging. *Fertil Steril* 2003; 79: 1294-1298.
10. Oudendijk JF, Yarde F, Eijkemans MJ, Broekmans FJ, Broer SL. The poor responder in IVF: is the

- prognosis always poor? A systematic review. *Hum Reprod Update* 2012; 18: 1-1.
11. Fauser BC, van Heusden AM. Manipulation of Human Ovarian Function: Physiological Concepts and Clinical Consequences 1. *Endocr Rev* 1997; 18: 71-106.
  12. Surrey ES, Schoolcraft WB. Evaluating strategies for improving ovarian response of the poor responder undergoing assisted reproductive techniques. *Fertil Steril* 2000; 73: 667-676.
  13. Chun SY, Eisenhauer KM, Minami SA, Billig H, Perlas EM, Hsueh AJ. Hormonal regulation of apoptosis in early antral follicles: follicle-stimulating hormone as a major survival factor. *Endocrinology* 1996; 137: 1447-1456.
  14. Klein NA, Battaglia DE, Fujimoto VY, Davis GS, Bremner WJ, Soules MR. Reproductive aging: accelerated ovarian follicular development associated with a monotropic follicle-stimulating hormone rise in normal older women. *J Clin Endocrinol Metab* 1996; 81:1038-1045.
  15. Roseff SJ, Bangah ML, Kettel LM, Vale W, Rivier J, Burger HG, Yen SS. Dynamic Changes in Circulating Inhibin Levels during the Luteal-Follicular Transition of the Human Menstrual Cycle. *J Clin Endocrinol Metab* 1989; 69: 1033-1039.
  16. Mais V, Cetel NS, Muse KN, Quigley ME, Reid RL, Yen SS. Hormonal dynamics during luteal-follicular transition. *J Clin Endocrinol Metab* 1987; 64: 1109-1114.
  17. Hill MJ, McWilliams GD, Miller KA, Scott RT, Frattarelli JL. A luteal estradiol protocol for anticipated poor-responder patients may improve delivery rates. *Fertil Steril* 2009; 91: 739-743.
  18. Schachter M, Friedler S, Raziel A, Strassburger D, Bern O, Ron-El R. Clinical Assisted Reproduction: Improvement of IVF Outcome in Poor Responders by Discontinuation of GnRH Analogue During the Gonadotropin Stimulation Phase-A Function of Improved Embryo Quality. *J Assist Reprod Genet* 2001; 18: 199-206.
  19. Feldberg D, Farhi J, Ashkenazi J, Dicker D, Shalev J, Ben-Rafael Z. Minidose gonadotropin-releasing hormone agonist is the treatment of choice in poor responders with high follicle-stimulating hormone levels. *Fertil Steril* 1994; 62: 343-346.
  20. Olivennes F, Righini C, Fanchin R, Torrisi C, Hazout A, Glissant M, Fernandez H, Frydman R. A protocol using a low dose of gonadotrophin-releasing hormone agonist might be the best protocol for patients with high follicle stimulating hormone concentrations on day 3. *Hum Reprod* 1996; 11: 1169-1172.
  21. Fanchin R, Cunha-Filho JS, Schonäuer LM, Righini C, de Ziegler D, Frydman R. Luteal estradiol administration strengthens the relationship between day 3 follicle-stimulating hormone and inhibin B levels and ovarian follicular status. *Fertil Steril* 2003; 79: 585-589.
  22. Fanchin R, Cunha-Filho JS, Schonauer LM, Kadoch IJ, Cohen-Bacri P, Frydman R. Coordination of early antral follicles by luteal estradiol administration provides a basis for alternative controlled ovarian hyperstimulation regimens. *Fertil Steril* 2003; 79: 316-321.
  23. Fanchin R, Salomon L, Castelo-Branco A, Olivennes F, Frydman N, Frydman R. Luteal estradiol pre-treatment coordinates follicular growth during controlled ovarian hyperstimulation with GnRH antagonists. *Hum Reprod* 2003; 18: 2698-2703.
  24. Fanchin R, Branco AC, Kadoch IJ, Hosny G, Bagirova M, Frydman R. Premenstrual administration of gonadotropin-releasing hormone antagonist coordinates early antralfollicle sizes and sets up the basis for an innovative concept of controlled ovarian hyperstimulation. *Fertil Steril* 2004; 81: 1554-1559.
  25. Cakmak H, Tran ND, Zamah AM, Cedars MI, Rosen MP. A novel "delayed start" protocol with gonadotropin-releasing hormone antagonist improves outcomes in poor responders. *Fertil Steril* 2014; 101: 1308-1314.
  26. Cruz M, Gadea B, Garrido N, Pedersen KS, Martínez M, Pérez-Cano I, Muñoz M, Meseguer M. Embryo quality, blastocyst and ongoing pregnancy rates in oocyte donation patients whose embryos were monitored by time-lapse imaging. *J Assist Reprod Genet* 2011; 28: 569-573.
  27. Weitzman VN, Engmann L, DiLuigi A, Maier D, Nulsen J, Benadiva C. Comparison of luteal estradiol patch and gonadotropin-releasing hormone antagonist suppression protocol before gonadotropin stimulation versus microdose gonadotropin-releasing hormone agonist protocol for patients with a history of poor in vitro fertilization outcomes. *Fertil Steril* 2009; 92: 226-230.
  28. Buster J, Bustillo M, Thorneycroft I, Simon J, Boyers S, Marshall J, Louw J, Seed R, Seed R. Non-surgical transfer of in vivo fertilised donated ova to five infertile women: report of two pregnancies. *Lancet* 1983; 322: 223-224.
  29. Keltz M, Sauerbrun-Cutler MT, Breborowicz A. Managing poor responders in IVF. *Exp Rev Obstet Gynecol* 2014; 8: 121-134.
  30. Pu D, Wu J, Liu J. Comparisons of GnRH antagonist versus GnRH agonist protocol in poor ovarian responders undergoing IVF. *Hum Reprod* 2011; 26: 2742-2749.
  31. Akman MA, Erden HF, Tosun SB, Bayazit N, Aksoy E, Bahceci M. Comparison of agonistic flare-up-protocol and antagonistic multiple dose protocol in ovarian stimulation of poor responders: results of a prospective randomized trial. *Hum Reprod* 2001; 16: 868-870.
  32. Kahraman K, Berker B, Atabekoglu CS, Sonmezer M, Cetinkaya E, Aytac R, Satioglu H. Microdose gonadotropin-releasing hormone agonist flare-up protocol versus multiple dose gonadotropin-releasing hormone antagonist protocol in poor responders undergoing intracytoplasmic sperm injection-embryo transfer cycle. *Fertil Steril* 2009; 91: 2437-2444.
  33. Dragisic KG, Davis OK, Fasouliotis SJ, Rosenwaks Z. Use of a luteal estradiol patch and a gonadotropin-releasing hormone antagonist suppression protocol before gonadotropin stimulation for in vitro fertilization in poor responders. *Fertil Steril* 2005; 84: 1023-1026.
  34. Ata B, Zeng X, Son WY, Holzer H, Tan SL. Follicular synchronization using transdermal estradiol patch and GnRH antagonists in the luteal phase; does it increase oocyte yield in poor responders to gonadotropin stimulation for in vitro fertilization (IVF)? A comparative study with microdose flare-up protocol. *Gynecol Endocrinol* 2011; 27: 876-879.

35. Shastri SM, Barbieri E, Kligman I, Schoyer KD, Davis OK, Rosenwaks Z. Stimulation of the young poor responder: comparison of the luteal estradiol/gonadotropin-releasing hormone antagonist priming protocol versus oral contraceptive microdose leuprolide. *Fertil Steril* 2011; 95: 592-595.
36. Mashayekhi M, Karimzadeh MA. Comparison of mild and microdose GnRH agonist flare protocols on IVF outcome in poor responders. *Arch Gynecol Obstet* 2011; 283: 1159-1164