

The effect of 24 hours delay in oocyte maturation triggering in IVF/ICSI cycles with antagonist protocol and not-elevated progesterone: A randomized control trial

Robab Davar¹ M.D., Elham Naghshineh^{1,2} M.D., Nosrat Neghab¹ M.D.

1. Research and Clinical Center for Infertility, Yazd Reproductive Sciences Institute, Shahid Sadoughi University of Medical Sciences, Yazd, Iran.
2. Department of Obstetrics and Gynecology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran.

Corresponding Author:
Elham Naghshineh, Department of Obstetrics and Gynecology, School of Medicine, Isfahan University of Medical Sciences, Hezar Jerib Ave., Isfahan, Iran. Postal code: 8174675731
Email: naghshineh@med.mui.ac.ir
Tel: (+98) 3136201993

Received: 11 March 2017
Revised: 15 May 2017
Accepted: 6 June 2017

Abstract

Background: The best time of final oocyte maturation triggering in assisted reproduction technology protocols is unknown. This time always estimated by combined follicular size and blood progesterone level.

Objective: The aim of this study was evaluation of the effect of delaying oocyte maturation triggering by 24 hr on the number of mature oocytes (MII) and other in vitro fertilization cycle characteristics in antagonist protocols with not-elevated progesterone ($p \leq 1$ ng/ml).

Materials and Methods: All patients' candidate for assisted reproduction technology underwent controlled ovarian hyperstimulation by antagonist protocol. When at least 3 follicles with ≥ 18 mm diameters were seen by vaginal ultrasonography; blood progesterone level was measured. The patients who had progesterone level ≤ 1 ng/dl entered the study. The participants' randomizations were done and patients were divided into two groups. In the first group, final oocyte maturation was done by human chorionic gonadotropin at the same day, but in the second group, this was performed 24 hr later. Oocytes retrieval was done 36 hr after human chorionic gonadotropin trigger by transvaginal ultrasound guide.

Results: Number of retrieved oocytes, mature oocytes (MII), fertilized oocytes (2PN), embryos formation, number of transferred embryos and embryos quality has not significant differences between two groups. Also, fertilization and implantation rate, chemical and clinical pregnancy did not differ between groups.

Conclusion: Delaying of triggering oocyte maturation by 24 hr in antagonist protocol with not-elevated progesterone (progesterone ≤ 1 ng/ml) have not beneficial nor harmful effect on the number of mature oocytes (MII) and other in vitro fertilization cycle characteristics.

Keywords: Assisted reproductive technologies, In vitro fertilization, Randomized controlled trial.

Registration ID in IRCT: IRCT2016071628756N3

Introduction

There are limited information for appropriate time of final oocyte maturation triggering in assisted reproductive technology (ART) cycles (1). The best time for triggering final oocyte maturation is dependent on several factors, such as diameter of largest follicle, blood estradiol and progesterone level on trigger day, peak estradiol to the number of follicles larger than 14 mm ratio, and patient prior protocols used for controlled ovarian hyperstimulation. Oocyte quality and endometrium receptivity are depending on time of luteinization before

oocyte retrieval (2). As mentioned by Hu *et al* that there is a positive relationship between follicular size and the level of cytoplasmic maturation. Postponing the oocyte maturation triggering through HCG administration delay, and so prolongation of follicular phase (which result in more mature oocytes) may have positive effects on clinical outcomes (2).

Rise in blood progesterone level can be a predictor of follicles numbers and serum estradiol up rise (3). The negative impact of progesterone rise on the endometrium is more than the oocyte/embryo quality (1). High serum progesterone level could result in endometrial maturation progression and

disable endometrial receptivity (4, 5). Van Vaerenbergh *et al*, Bosch *et al* and Labarta *et al* discovered that serum progesterone levels more than 1.5 ng/ml lead to transformation of gene expression in the endometrium, and cause reduced endometrial receptivity (6-8). Elevated serum progesterone levels on the day of HCG administration ($p > 1.5$ ng/ml) decreased ongoing pregnancy and delivery rates (7, 9). Vandekerckhove *et al* showed, if the progesterone level is higher than 1 ng/ml, delaying the administration of HCG by 24 hr has no effect on the number of mature oocytes (10).

Although, Tremellen and Lane demonstrated that 24 hr delay in oocyte retrieval from the best predicted time can cause small but significant increase in collected oocytes numbers and embryos formation, but had no meaningful effect on live birth rates (11). Kolibianakis *et al* also represent that follicular phase lengthening by HCG delaying administration cause more advancement in endometrium on oocyte retrieval day (12).

Vandekerckhove *et al* showed that in patients with low serum progesterone level (≤ 1 ng/ml) 24 hr delaying in the oocyte maturation trigger could cause to more total oocytes and mainly more mature oocytes (MII) in contrast with those triggered on the same day. Although, no meaningful differences were seen in the pregnancy rates and pregnancy outcomes between two groups (10). But in the patients with higher serum progesterone level (> 1 ng/ml) delaying the oocyte maturation trigger by 24 hr accompanied with no difference in IVF cycle characteristics (10). Venetis *et al*, Mio *et al*, Bustillo *et al* and Ventis *et al* demonstrated in cycles with higher progesterone levels, more oocytes were retrieved, but a cutoff point for progesterone levels did not achieved (13-16). The best time for oocyte maturation triggering by use of the ultrasound criteria is still challengeable (17). Commonly, it is accepted that follicles must achieve to at least 17 mm in diameter before triggering for final oocyte maturation (18).

The aim of this study was evaluation of the effect of 24 hr delaying of oocyte maturation triggering on the number of mature oocytes

(MII) and other IVF cycle characteristics in antagonist cycles with not-elevated progesterone ($p \leq 1$ ng/ml).

Materials and methods

This randomized controlled trial was performed at Research and Clinical Center for Infertility, Shahid Sadoughi University of Medical Sciences, Yazd, Iran, between August and October 2016. Based on Vandekerckhove *et al* study with Statistics Consultant, at least sample size of 20 patients in each group was needed (10). Infertile women, who their controlled ovarian hyperstimulation was done with GnRH antagonist protocol with fresh embryo transfer in the same cycle, were included. Participants with premature elevation of progesterone (> 1 ng/ml) and non-fresh embryo transfer and sever male factor infertility were excluded. Eighty five women were included in the study (42 in the delay triggering group, and 43 in the same day triggering group). The patients were randomized by random allocation software.

Stimulation protocols

All patients underwent GnRH antagonist controlled ovarian hyperstimulation protocols in ART cycles. The gonadotropins (Gonal-F, Merck-Serono, Spain) were started on day 2 of the menstrual cycle. The initial dose of gonadotropin was individualized for each infertile patient according to the age, body mass index, antral follicle count, anti-mulerian hormone (AMH) level, and previous responsiveness to ovarian stimulation. Then gonadotropins dose adjustments were done based on ovarian response by follicular diameter measurement with transvaginal ultrasound, obtained every 2 or 3 days from 7th day of stimulation.

The GnRH antagonist (cetorelix, MERC-SERONO, France) was administered when the mean diameter of dominant follicles reached to 13-14 mm, and continues until the day of triggering with HCG. When at least 3 follicles with ≥ 18 mm diameter were seen by vaginal ultrasonography, then blood estradiol and progesterone level was measured. The patients who had progesterone level ≤ 1 ng/dl

entered the study. The participants' randomizations were done using random numbers table, and patients were divided into two groups.

In the first group, final oocyte maturation was done by HCG (Pregnyl, Organon, Netherlands) at the same day, and 24 hr later in the second group. Oocytes retrieval was performed 36 hr after HCG trigger through transvaginal ultrasound guided. Routine IVF/ICSI was performed as appropriate. The luteal phase was supported with vaginal progesterone (Cyclogest, Actavis, UK), 400 mg, twice a day. One or two embryos have transferred 2 days later. Fertilization rate was defined as the percentage of fertilized oocytes (2PN) to all mature oocytes (MII). Chemical pregnancies were confirmed 2 wk after embryo transfer, by positive serum HCG measurement. Clinical pregnancy was defined by the presence of gestational sac in the uterus, 4 wk after embryo transfer. The implantation rate was the proportion of embryos transferred resulting in an intrauterine gestational sac.

Clinical data collection

During treatment, all patients' information including age, FSH level in 3rd day of the menstrual cycle before the treatment, AMH, duration of gonadotropin administration, total gonadotropin dose, estradiol and progesterone levels on the HCG day, and number of total oocytes retrieved, number of mature oocytes (MII), and number of fertilized oocytes (2PN), numbers of embryo formation and transferred embryos were documented. Chemical and clinical pregnancy was identified, and fertilization rate and implantation rate were also calculated.

Ethical consideration

This study was approved by the Ethics Committee of the Shahid Sadoughi University of Medical Sciences (IR.SSU.RSI.REC.1395.4). Informed written consent was provided from all participants.

Statistical analysis

The statistical analysis were performed using statistical package for the social sciences, version 17.0, (SPSS Inc, Chicago, Illinois, USA). Categorical data were expressed as number and percentage. For qualitative data, Chi-square test was used. For quantitative data, the normality status was checked at first with Kolmogorov Smirnov test. In normal data distribution, student-t, and in abnormal data distribution, Mann whitney-U test was used.

Results

Totally 85 infertile women (42 in the delay triggering group, and 43 in the same day triggering group) participated in this study (Figure 1). The mean age of participants was 32.2±6.7 yr old (32.5±4.6 in the delay triggering group, and 31.9±4.7 in the same day triggering group). Other demographic and ART cycle characteristics are shown in table I and II. There were no significant differences in age, third day FSH, basal AMH, and other characteristics between two groups. Number of retrieved oocytes, mature oocytes (MII), fertilized oocytes (2PN), embryos formation, transferred embryos and embryos quality has not significant differences between two groups. Also, fertilization and implantation rate, chemical and clinical pregnancy did not differ between groups.

Table I. Demographic and basal characteristics of the participants

Variable	Delay triggering group (n=42)	Same day triggering group (n=43)	p-value
Age (years) [#]	32.5 ± 4.6	31.9 ± 4.7	0.51
Infertility duration (years) [§]	6.26 ± 0.59	6.98 ± 0.63	0.26
Infertility type ^{**&}			0.54
Primary	33 (78.6%)	36 (83.7%)	
Secondary	9 (21.4%)	7 (16.3%)	
No. of previous failed IVF [§]	0.51 ± 0.19	0.50 ± 0.21	0.62
Third day FSH [#]	6.1 ± 2.4	6.5 ± 2.7	0.66
AMH [#]	3.5 ± 3.7	3.4 ± 2.5	0.90

*Data are presented as mean±SD

[#]Comparison was done by independent t-test

[&] Comparison was done by chi-square test

FSH= Follicle-stimulating hormone

** Data are presented as n (%)

[§]Comparison was done by Mann Whitney- U test

AMH= Anti-Müllerian hormone

IVF: In vitro fertilization

Table II. Cycle characteristics of the participants

Variable	Delay triggering group (n=42)	Same day triggering group (n=43)	p-value
Estradiol level on trigger day (pg/ml) ^{*#}	1547 ± 688	1841 ± 960	0.11
Progesterone level on trigger day (ng/ml) ^{*#}	0.72 ± 0.21	0.76 ± 0.16	0.28
Gonadotropin dose (IU) ^{*#}	2053 ± 666	2075 ± 821	0.89
Cycle duration (day) ^{*§}	14.2 ± 0.24	13 ± 0.30	0.03
Number of retrieved oocytes ^{*§}	9.5 ± 4.9	8.3 ± 4.7	0.22
Number of mature oocytes (MII) ^{*§}	8.2 ± 0.66	7.5 ± 0.69	0.24
Number of 2PN ^{*§}	4.6 ± 0.51	4.6 ± 0.56	0.73
Number of embryos ^{*§}	4.1 ± 0.48	4.3 ± 0.57	0.82
Number of transferred embryos ^{*§}	1.7 ± 0.13	1.9 ± 0.09	0.65
Total transferred embryo [§]	75	81	0.80
Embryo quality A ^{**&}	9 (12%)	8 (9.8%)	0.89
Embryo quality B ^{**&}	41 (54.6%)	44 (54.3%)	
Embryo quality C ^{**&}	25 (33.3%)	29 (35.8%)	
ET in trigger day (mm) ^{*§}	8.6 ± 0.22	9.0 ± 0.33	0.79
Fertilization rate	55%	59%	0.42
Chemical pregnancy ^{**&}	11 (30.6%)	9 (23.7%)	0.34
Clinical pregnancy ^{**&}	7 (19.4%)	8 (21.0%)	0.78
Implantation rate	10%	9.8%	0.91

*Data are presented as mean±SD

§Comparison was done by Mann Whitney- U test

pg/ml= Picogram/milliliter

IU= International Unit

2PN= Two-pronuclear zygote

** Data are presented as n (%)

& Comparison was done by chi-square test

ng/ml= Nanogram/milliliter

MII= Meiosis II (mature oocyte)

ET= Endometrial thickness

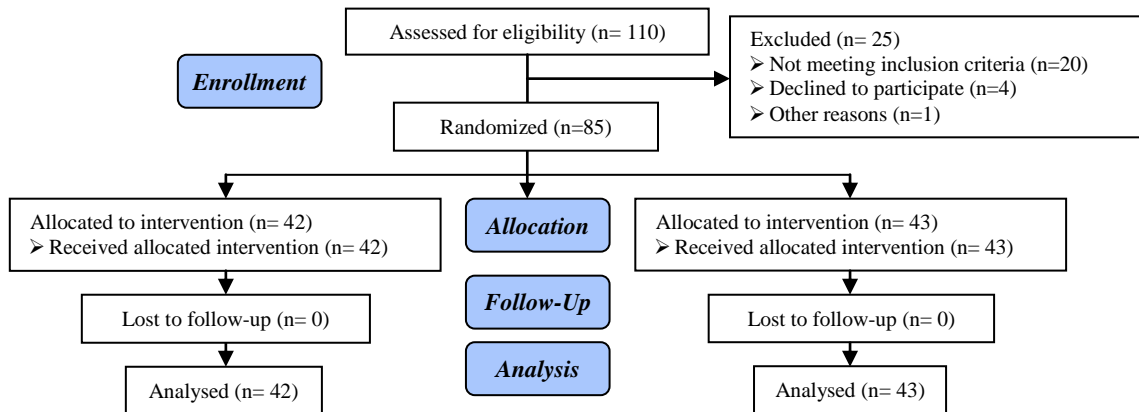


Figure 1. Consort flow chart

Discussion

The best time for oocyte maturation triggering by the ultrasound criteria is challengeable. In older cycles without use of GnRH agonist/antagonist, when maximum follicle diameter reached to 16 mm or more, and a serum estradiol was at least 600 pg/ml, triggering with hCG was considered (17). In antagonist cycles, the scientists agree with the leading follicle diameters of 16-17 mm (12, 19-21). In antagonist cycles, triggering is done earlier than in agonist cycles (21). In our study, we used follicle diameters (at least 3 follicles ≥ 18 mm) in selected patients with low progesterone levels (≤1 ng/ml). Numbers of retrieved oocytes, mature oocytes (MII), fertilized oocytes (2PN), embryos formation, transferred embryos and embryos quality

have not significant differences between two groups (Table II).

Vandekerckhove *et al* showed that in patients with low serum progesterone level (≤1 ng/ml), 24 hr delaying in the oocyte maturation trigger can cause more total oocytes and mainly more mature oocytes (MII), in contrast with those triggered on the same day (10). Tremellen and Lane demonstrated that 24 hr delay in oocyte retrieval can cause small but significant increase in collected oocytes numbers and embryos formation, but have no meaningful effect on live birth rates. They found that, advancing or delaying hCG administration by 1 day from ‘ideal’ had no adverse effect on IVF outcomes in GnRH antagonist cycles (11).

Also, our study results demonstrated that 24 hr delay in triggering has no effect on

fertilization and implantation rate, chemical and clinical pregnancy between two groups. Vandekerckhove *et al* advised that as soon as three follicles reached to a diameter ≥ 18 mm, decisions for best time of triggering must be made depend on the progesterone level. They recommended that if the progesterone level were more than 1 ng/ml, delaying in triggering has no effect on the mature oocytes. But if the progesterone level is ≤ 1 ng/ml, delaying oocyte maturation by 24 hr is advisable (9). Tremmelen and Lane found that advancing or delaying hCG administration by 1 day from 'ideal' had no adverse effect on IVF outcomes in GnRH antagonist cycles (10, 11).

Conclusion

Our findings in this study support these recommendations. Our study data showed that although delay in final oocyte triggering may have not beneficial effect on ART cycle's results, but it has not adverse effects, too.

Acknowledgments

This study was supported by Yazd Research and Clinical Center for Infertility, Shahid Sadoughi University of Medical Sciences. We want to thank all of study participants and infertility clinic staff specially Mrs. Behnaz Gandom.

Conflict of interest

All of the authors indicate they have not any financial support. All of the authors state that they have had full control of all primary data and that they agree to allow the Journal to review their data if requested.

References

1. Al-Azemi M, Kyrou D, Kolibianakis E, Humaidan P, Van Vaerenbergh I, Devroey P, et al. Elevated progesterone during ovarian stimulation for IVF. *Reprod Biomed Online* 2012; 24: 381-388.
2. Hu X, Luo Y, Huang K, Li Y, Xu Y, Zhou C, et al. New Perspectives on Criteria for the Determination of HCG Trigger Timing in GnRH Antagonist Cycles. *Medicine* 2016; 95: e3691.
3. Kyrou D, Al-Azemi M, Papanikolaou E, Donoso P, Tziomalos K, Devroey P, et al. The relationship of premature progesterone rise with serum estradiol levels and number of follicles in GnRH antagonist/recombinant FSH-stimulated cycles. *Eur J Obstet Gynecol Reprod Biol* 2012; 162: 165-168.
4. Aflatoonian A, Davar R, Hojjat F. Elevated serum progesterone/MII oocyte ratio on the day of human chorionic gonadotropin administration can predict impaired endometrial receptivity. *Iran J Reprod Med* 2014; 12: 427.
5. Vandekerckhove F, De Baerdemaeker A, Gerris J, Tilleman K, Vansteelandt S, De Sutter P. A randomized controlled trial to evaluate the effect on oocyte yield of prolongating the follicular phase in IVF/ICSI cycles with not-elevated progesterone. *Hum Reprod* 2013; 28 (Suppl.): i311-i356.
6. Van Vaerenbergh I, Fatemi H, Blockeel C, Van Lommel L, Schuit F, Kolibianakis E, et al. Progesterone rise on HCG day in GnRH antagonist/rFSH stimulated cycles affects endometrial gene expression. *Reprod Biomed Online* 2011; 22: 263-271.
7. Bosch E, Labarta E, Crespo J, Simon C, Remohi J, Jenkins J, et al. Circulating progesterone levels and ongoing pregnancy rates in controlled ovarian stimulation cycles for in vitro fertilization: analysis of over 4000 cycles. *Hum Reprod* 2010; 25: 2092-2100.
8. Labarta E, Martínez-Conejero J, Alamá P, Horcajadas J, Pellicer A, Simón C, et al. Endometrial receptivity is affected in women with high circulating progesterone levels at the end of the follicular phase: a functional genomics analysis. *Hum Reprod* 2011; 26: 1813-1825.
9. Papanikolaou E, Pados G, Grimbizis G, Bili E, Kyriazi L, Polyzos N, et al. GnRH-agonist versus GnRH-antagonist IVF cycles: is the reproductive outcome affected by the incidence of progesterone elevation on the day of HCG triggering? A randomized prospective study. *Hum Reprod* 2012; 27: 1822-1828.
10. Vandekerckhove F, Gerris J, Vansteelandt S, De Baerdemaeker A, Tilleman K, De Sutter P. Delaying the oocyte maturation trigger by one day leads to a higher metaphase II oocyte yield in IVF/ICSI: a randomised controlled trial. *Reprod Biol Endocrinol* 2014; 12: 31.
11. Tremellen K, Lane M. Avoidance of weekend oocyte retrievals during GnRH antagonist treatment by simple advancement or delay of hCG administration does not adversely affect IVF live birth outcomes. *Hum Reprod* 2010; 25: 1219-1224.
12. Kolibianakis EM, Bourgain C, Papanikolaou EG, Camus M, Tournaye H, Van Steirteghem AC, et al. Prolongation of follicular phase by delaying hCG administration results in a higher incidence of endometrial advancement on the day of oocyte retrieval in GnRH antagonist cycles. *Hum Reprod* 2005; 20: 2453-2456.
13. Venetis C, Kolibianakis E, Papanikolaou E, Bontis J, Devroey P, Tarlatzis B. Is progesterone elevation on the day of human chorionic gonadotrophin administration associated with the probability of pregnancy in in vitro fertilization? A systematic review and meta-analysis. *Hum Reprod Update* 2007; 13: 343-355.
14. Venetis C, Kolibianakis E, Bosdou J, Tarlatzis B. Progesterone elevation and probability of pregnancy after IVF: a systematic review and meta-analysis of over 60000 cycles. *Hum Reprod Update* 2013; dmt014.
15. Mio Y, Sekijima A, Iwabe T, Onohara Y, Harada T, Terakawa N. Subtle rise in serum progesterone

- during the follicular phase as a predictor of the outcome of in vitro fertilization. *Fertil Steril* 1992; 58: 159-166.
16. Bustillo M, Stern JJ, Coulam CB. Serum progesterone at the time of human chorionic gonadotrophin does not predict pregnancy in in-vitro fertilization and embryo transfer. *Hum Reprod* 1995; 10: 2862-2867.
 17. Murad N. Ultrasound or ultrasound and hormonal determinations for in vitro fertilization monitoring. *Int J Gynecol Obstet* 1998; 63: 271-276.
 18. Kyrrou D, Kolibianakis EM, Fatemi HM, Tarlatzis BC, Tournaye H, Devroey P. Is earlier administration of human chorionic gonadotropin (hCG) associated with the probability of pregnancy in cycles stimulated with recombinant follicle-stimulating hormone and gonadotropin-releasing hormone (GnRH) antagonists? A prospective randomized trial. *Fertil Steril* 2011; 96: 1112-1115.
 19. Hauzman E, Bodri D, Guillen J, Vidal R, Coll O, Vernaeve V. Exploring different criteria by 2D or 3D ultrasound for triggering final oocyte maturation in an oocyte donation program: a randomized pilot study. *Hum Reprod* 2011; 26 (suppl.): i95-i96.
 20. Kolibianakis EM, Papanikolaou EG, Tournaye H, Camus M, Van Steirteghem AC, Devroey P. Triggering final oocyte maturation using different doses of human chorionic gonadotropin: a randomized pilot study in patients with polycystic ovary syndrome treated with gonadotropin-releasing hormone antagonists and recombinant follicle-stimulating hormone. *Fertil Steril* 2007; 88: 1382-1388.
 21. Tarlatzis B, Fauser B, Kolibianakis E, Diedrich K, Devroey P. GnRH antagonists in ovarian stimulation for IVF. *Hum Reprod Update* 2006; 12: 333-340.
 22. Mochtar M, Custers I, Koks C, Bernardus R, Verhoeve H, Mol B, et al. Timing oocyte collection in GnRH agonists down-regulated IVF and ICSI cycles: a randomized clinical trial. *Hum Reprod* 2011; 26: 1091-1096.